## Package 'skewr'

April 16, 2024

**Title** Visualize Intensities Produced by Illumina's Human Methylation 450k BeadChip

**Version** 1.34.0

Description The skewr package is a tool for visualizing the output of the Illumina Human Methylation 450k BeadChip to aid in quality control. It creates a panel of nine plots. Six of the plots represent the density of either the methylated intensity or the unmethylated intensity given by one of three subsets of the 485,577 total probes. These subsets include Type I-red, Type I-green, and Type II.The remaining three distributions give the density of the Beta-values for these same three subsets. Each of the nine plots optionally displays the distributions of the ``rs' SNP probes and the probes associated with imprinted genes as series of 'tick' marks located above the x-axis.

**Depends** R (>= 3.1.1), methylumi, wateRmelon, mixsmsn, IlluminaHumanMethylation450kmanifest

**Imports** minfi, S4Vectors (>= 0.19.1), RColorBrewer

Suggests GEOquery, knitr, minfiData

VignetteBuilder knitr

License GPL-2

LazyData true

biocViews DNAMethylation, TwoChannel, Preprocessing, QualityControl

Author Ryan Putney [cre, aut], Steven Eschrich [aut], Anders Berglund [aut]

Maintainer Ryan Putney <ryanputney@gmail.com>

git\_url https://git.bioconductor.org/packages/skewr

git\_branch RELEASE\_3\_18

git\_last\_commit f7de07a

git\_last\_commit\_date 2023-10-24

Repository Bioconductor 3.18

**Date/Publication** 2024-04-15

2 getBarcodes

## **R** topics documented:

getMethyLumiSet	
panelPlots	
preprocess	
	5
subsetProbes	

Index 10

getBarcodes

Get barcodes from idat file names

## Description

A convenience function for retrieving simple barcodes from idat file names.

## Usage

```
getBarcodes(path = getwd(), recurse = FALSE)
```

## Arguments

The path or a character vector to the directory or directories in which to find the

idat files.

recurse logical; should the function check subdirectories to derive barcodes from any

found idat files. The default is FALSE.

#### **Details**

Barcodes will be generated by all found idats in path(s). The default path is the current working directory.

#### Value

A character vector of barcodes.

## Author(s)

Ryan Putney <ryanputney@gmail.com>

#### See Also

getMethyLumiSet

getMethyLumiSet 3

#### **Examples**

```
if(require(minfiData)){
  path <- system.file("extdata/5723646052", package="minfiData")
  barcodes <- getBarcodes(path = path)
}</pre>
```

getMethyLumiSet

Read idat files and create a MethyLumiSet object

## **Description**

This a wrapper function for methylumIDAT that does not require a vector of barcodes to be provided.

#### Usage

## **Arguments**

path The path to the directory containing the idat files.

barcodes A vector of barcodes specifying which idat's to read.

norm Should normalization be done on the resulting MethyLumiSet. The default is

"none"

bg.corr logical; if TRUE, an Illumina style background subtraction will be performed

only if norm is set to 'illumina'. Otherwise, it is ignored. If background subtraction without any normalization is desired, the preprocess method must

be used.

#### **Details**

If only path is provided, all idat's found in the given folder will be pulled. If only barcodes is given, corresponding idat's will be pulled from the current working directory. Both path and barcodes may be passed for finer control. The default is to pull all idat's found in the current working directory.

#### Value

A MethyLumiSet object

#### Note

One would probably not normally want to use the preprocess option at this stage. It is more likely that a MethyLumiSet of the raw data will be desired. Then the preprocess method may be used to normalize the raw data or use background subtraction only on the raw data. See the vignette for example workflow.

4 getSNparams

#### Author(s)

Ryan Putney <ryanputney@gmail.com>

#### References

Davis S, Du P, Bilke S, Triche T, Jr. and Bootwalla M (2014). methylumi: Handle Illumina methylation data. R package version 2.12.0.

#### See Also

```
getBarcodes, preprocess
```

#### **Examples**

```
if(require('minfiData')) {
  path <- system.file("extdata/5723646052", package="minfiData")
  barcodes <- getBarcodes(path = path)
  methylumiset.raw <- getMethyLumiSet(path = path, barcodes = barcodes[1])
}</pre>
```

getSNparams

Estimate parameters for finite mixture of Skew-Normal distributions

## Description

Utilizes smsn.mix from the mixsmsn package to find the parameters for a finite mixture of skew normal distributions to model the overall distribution of signal intensities for a subset of probes on the Illumina Infinium HumanMethylation450. The probes may be subset by type and methylated or unmethylated. It can also be specified whether the SNP(rs), imprinted(idmr), or ch probes should be included or filtered out prior to parameter estimation.

#### Usage

## Arguments

MethyLumiSet	A MethyLumiSet object
allele	Should parameter estimation be done on the methylated or unmethylated signal intensities
type	Use the signal intensities for which probe type
snps	logical; should the rs probes be included in the dataset. The default is TRUE
idmr	logical; should the probes of imprinted gene loci be included in the dataset. The default is TRUE
ch	logical; should the ch probes be included in the dataset. The default is FALSE

panelPlots 5

#### Value

A Skew.normal object as returned by smsn.mix from the mixsmsn package with the means and modes of the components added.

#### Author(s)

Ryan Putney <ryanputney@gmail.com>

#### References

Pidsley R, Wong CCY, Volta M, Lunnon K, Mill J, Schalwyk LC(2013). A data-driven approach to preprocessing Illumina 450k methylation array data. BMC Genomics, 14:293.

Prates MO, Cabral CRB, Lachos VH (2013).mixsmsn: Fitting Finite Mixture of Scale Mixture of Skew-Normal Distributions. Journal of Statistical Software, 54(12), 1-20. http://www.jstatsoft.org/v54/i12/

#### See Also

subsetProbes

## Examples

```
if(require('wateRmelon')) {
  data(melon)
  mixes.raw.meth.II <- getSNparams(melon[,1], 'M', 'II')
}</pre>
```

panelPlots

Plot the distributions of the probe intensities and the components of the skew-normal mixture model

#### **Description**

Creates a panel of nine plots. Six of the plots represent the density of either the methylated intensity or the unmethylated intensity given by one of three subsets of the 485,577 total probes. These subsets include Type I-red, Type I-green, and Type II.The remaining three distributions give the density of the beta-values for these same three subsets. Each of the nine plots optionally displays the distributions of the "rs" SNP probes and the probes associated with imprinted genes(Pidsley,2013) as a series of 'tick' marks located above the x-axis.

#### Usage

6 panelPlots

#### **Arguments**

MethyLumiSet The MethyLumiSet object from which the mixture models were derived

typeIRedModels A list of the Type I-red mixture models listed in the following order: methy-

lated models followed by unmethylated models

typeIGreenModels

A list of the Type I-green mixture models listed in the following order: methy-

lated models followed by unmethylated models

typeIIModels A list of the Type II mixture models listed in the following order: methylated

models followed by unmethylated models

plot Should the output consist of panel plots—one panel per sample or a single panel if

samp.num is specified; or should the function output separate plots corresponding to the frames, given by frame.nums, for a single sample. The default is

"panel". If set to "frames", samp.num must be specified

samp.num If plotting for a single sample is desired, for which sample. The number given

simply refers to the MethyLumiSet column that corresponds to the sample of

interest

frame.nums if plot is set to "frame", then frame.nums is a vector that specifies which frames

of the panel to plot. The default is to plot all nine frames. The frames are numbered from 1 to 9 in column-major order starting with the top left. For

example, to plot the four corners, use frame.nums=c(1,3,7,9)

norm A character string which will be displayed as part of the main title for each plot.

Useful in indicated which normalization method was used for the modeled and

plotted data

idmr logical; should the intensities of the idmr probes be plotted as a series of tick-

marks above the x-axis. The default is TRUE

snps logical; should the intensities of the rs probes be plotted as a series of tick-marks

above the x-axis. The default is TRUE

#### Value

No return value. Only plots are generated.

## Note

Please refer to the vignette for an example workflow.

#### Author(s)

Ryan Putney <ryanputney@gmail.com>

#### References

Prates MO, Cabral CRB, Lachos VH (2013).mixsmsn: Fitting Finite Mixture of Scale Mixture of Skew-Normal Distributions. Journal of Statistical Software, 54(12), 1-20. http://www.jstatsoft.org/v54/i12/

preprocess 7

#### See Also

```
getSNparams
```

#### **Examples**

```
if(require('minfiData')) {
  path <- system.file("extdata/5723646052", package="minfiData")
  methylumiset.raw <- getMethyLumiSet(path = path)
  mixes.raw.meth.I.red <- getSNparams(methylumiset.raw, 'M', 'I-red')
  mixes.raw.meth.I.green <- getSNparams(methylumiset.raw, 'M', 'I-green')
  mixes.raw.meth.II <- getSNparams(methylumiset.raw, 'M', 'II')
  mixes.raw.unmeth.I.red <- getSNparams(methylumiset.raw, 'U', 'I-red')
  mixes.raw.unmeth.I.green <- getSNparams(methylumiset.raw, 'U', 'I-green')
  mixes.raw.unmeth.II <- getSNparams(methylumiset.raw, 'U', 'II')
  mixes.I.red <- list(mixes.raw.meth.I.red, mixes.raw.unmeth.I.red)
  mixes.I.green <- list(mixes.raw.meth.I.green, mixes.raw.unmeth.I.green)
  mixes.II <- list(mixes.raw.meth.II, mixes.raw.unmeth.II)
  panelPlots(methylumiset.raw, mixes.I.red, mixes.I.green, mixes.II)
}</pre>
```

preprocess

Normalize a MethyLumiSet object using some popular choices

#### Description

This is a wrapper function that allows normalizing of a MethyLumiSet using either a BeadStudio approximation, SWAN, or dasen. If desired, background correction only may be performed on the raw data.

#### Usage

## Arguments

MethyLumiSet A MethyLumiSet object

norm The normalization method to be used

bg.corr If TRUE, background subtarction using negative controls is performed. Ignored

unless norm equals 'illumina' or 'none'

#### **Details**

Both Illumina style normalization via controls and the background correct method are handled by methylumi. The SWAN and dasen normalization methods are both performed by wateRmelon

8 subsetProbes

#### Value

A MethyLumiSet

#### Author(s)

Ryan Putney <ryanputney@gmail.com>

#### References

Davis S, Du P, Bilke S, Triche T, Jr. and Bootwalla M (2014). methylumi: Handle Illumina methylation data. R package version 2.12.0.

Maksimovic J, Gordon L, Oshlack A (2012). SWAN: Subset Quantile Within-Array Normalization for Illumina Infinium HumanMethylation450 BeadChips. Genome Biology, 13:R44.

Pidsley R, Wong CCY, Volta M, Lunnon K, Mill J, Schalwyk LC(2013). A data-driven approach to preprocessing Illumina 450k methylation array data. BMC Genomics, 14:293.

Schalkwyk LC, Pidsley R, Wong CC, Touleimat N, Defrance M, Teschendorff A and Maksimovic J (2013). wateRmelon: Illumina 450 methylation array normalization and metrics. R package version 1.5.1.

#### See Also

getMethyLumiSet

#### **Examples**

```
if(require('wateRmelon')) {
  data(melon)
  melon.dasen <- preprocess(melon, norm = 'dasen')
}</pre>
```

subsetProbes

Conveniently subset probes by type and retrieve the methylated or unmethylated intensities

#### **Description**

Thus function accepts a MethyLumiSet object generated by methylumi or a MethylSet object generated by minfi. It will subset the probes by type—"I-red", "I-green", or "II"—and return a matrix of the methylated, "M", or unmethylated, "U" signal intensities. It is also possible to include or filter out probes according to whether they are CpG sites(cg), SNPs(rs), imprinted(idmr) gene sites, or non-CpG loci(ch).

#### Usage

subsetProbes 9

## Arguments

object	A MethyLumiSet or MethylSet object
allele	Should methylated or unmethylated data for the probes be returned.
type	May be "I-red", "I-green", or "II".
cg	Logical; Should the returned dataset contain the CpG probes. The default is $\ensuremath{TRUE}$
snps	Logical; Should the returned dataset conain the rs probes. The default is TRUE
idmr	Logical; should the returned dataset include probes that interrogate imprinted gene sites as given by Pidsley et al.(2013). The default is TRUE
ch	Logical; should the returned dataset include the non-CpG (ch) probes. The default if FALSE

## Value

A matrix

## Author(s)

Ryan Putney <ryanputney@gmail.com>

#### References

Pidsley R, Wong CCY, Volta M, Lunnon K, Mill J, Schalwyk LC(2013). A data-driven approach to preprocessing Illumina 450k methylation array data. BMC Genomics, 14:293.

## See Also

getSNparams

## **Examples**

```
if(require('wateRmelon')) {
  data(melon)
  melon.meth.II <- subsetProbes(melon, 'M', 'II')
}</pre>
```

# **Index**

```
* IO
    {\tt getMethyLumiSet}, \color{red} {\tt 3}
* aplot
    panelPlots, 5
* manip
    preprocess, 7
     subsetProbes, 8
* models
     getSNparams, 4
* utilities
    getBarcodes, 2
getBarcodes, 2, 4
{\tt getMethyLumiSet}, 2, 3, 8
getSNparams, 4, 7, 9
panelPlots, 5
preprocess, 4, 7
subsetProbes, 5, 8
```