# Package 'PICS'

October 16, 2019

Type Package

Title Probabilistic inference of ChIP-seq

version 2.28.0
Author Xuekui Zhang <xzhang@stat.ubc.ca>, Raphael Gottardo</xzhang@stat.ubc.ca>
<rgottard@fhcrc.org></rgottard@fhcrc.org>
Maintainer Renan Sauteraud <rsautera@fhcrc.org></rsautera@fhcrc.org>
<b>Depends</b> R (>= 2.14.0), BiocGenerics (>= 0.1.3)
Imports methods, stats4, IRanges, GenomicRanges, graphics, grDevices, stats, Rsamtools, GenomicAlignments, S4Vectors
Suggests ShortRead, rtracklayer, parallel
SystemRequirements GSL (GNU Scientific Library)
<b>Description</b> Probabilistic inference of ChIP-Seq using an empirical Bayes mixture model approach.
License Artistic-2.0
biocViews Clustering, Visualization, Sequencing, ChIPSeq
git_url https://git.bioconductor.org/packages/PICS
git_branch RELEASE_3_9
git_last_commit 8cf1a4a
git_last_commit_date 2019-05-02
Date/Publication 2019-10-15
R topics documented:
bam2gr makeRangedDataOutput pics pics-class picsError-class picsFDR picsList-class plot-FDR segChrRead segmentPICS segReads  12

2 bam2gr

	segReadsList																		14
	segReadsListP	Έ.																 	15
	segReadsPE .																	 	15
	setParaEM																	 	16
	setParaPrior .																	 	17
	show																	 	18
	summary																	 	18
	summarySeg																	 	19
Index																			20

bam2gr

pre-process bam files

# **Description**

Reads a bam file using Rsamtools and extract the reads for each chromosome.

# Usage

```
bam2gr(bamFile, chr=NULL, PE=FALSE, verbose=FALSE)
```

# **Arguments**

bamFile A character string, the name of the .bam file to read.

chr An optional character string. If specified, only the selected chromosome will be

returned. Speed up the computation.

PE A logical. This should be set to TRUE for paired-end sequencing data.

verbose A logical. Print additional information about the data.

## Value

Returns a GRanges of all the reads for each chromosome.

# Note

The user might encounter a memory allocation error when using bam files of bigger sizes. Splitting the file by chromosome before calling bam2gr will solve this issue.

For Paired-End data, non matched reads are discarded.

# Author(s)

Renan Sauteraud

#### See Also

segmentPICS

makeRangedDataOutput Create a RangedData object from a PICS output

# **Description**

Create a list of 'RangedData' objects from a 'pics' object. The resulting RangedData object can then be analyzed with the 'IRanges' packages and/or exported to bed/wig files with the 'rtracklayer' package.

## Usage

make Ranged Data Output (obj, type="fixed", filter=list(delta=c(0,Inf), se=c(0,Inf), sigma SqF=c(0,Inf))

## **Arguments**

obj	An object of class 'picsList' as returned by 'PICS' when running it on the IP/Control data.
type	The type of intervals to be created. The different types are 'bed', 'wig', 'ci' and 'fixed'. See details for more info.
filter	A list of filters to be used before computing the FDR. By default all regions are included, see details for more info on how to specify the filters.
length	The length to be used for the fixed type 'RangedData', see details.

## **Details**

'bed' will generate intervals from the forward peak max to the reverse peak max. 'wig' will generate a density profile for the forward and reverse reads. 'bed' and 'wig' types should be used to be exported to wig/bed files to be used with the UCSC genome browser. 'ci' corresponds to the binding site estimates +/-3\*se, while 'fixed' corresponds to the binding site estimates +/-3\*length. 'bed' and 'wig' files can be exported using the 'export' function fo the 'rtracklayer' package.

# Value

An object of type 'RangedData'.

# Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

# References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

# See Also

export

4 pics

#### **Examples**

```
## Not run:
    rdBed<-makeRangedDataOutput(pics,type="bed",filter=list(delta=c(50,Inf),se=c(0,50),sigmaSqF=c(0,22500),si
    export(rbBed,"myfile.bed")
    rdBed<-makeRangedDataOutput(pics,type="wig",filter=list(delta=c(50,Inf),se=c(0,50),sigmaSqF=c(0,22500),si
    export(rbBed,"myfile.wig")
## End(Not run)</pre>
```

pics

Estimation of binding site positions

#### **Description**

This object contains Estimation of binding site positions and has the following slots: segReadsList, dataType.

#### Usage

PICS(segReadsList,dataType=NULL, paraEM=NULL, paraPrior=NULL, nCores=1)

#### **Arguments**

segReadsList

This object contains segmentation of Genome

dataType

The type of data you are processing: specified 'TF' for transcription factor.

paraEM

A list of parameters for the EM algorithm as returned by the setParaEm function. The default parameters should be good enough for most usages.

minK: an integer, default=1. The minimum number of binding events per region. If the value is 0, the minimum number is automatically calculated.

maxK: an integer, default=15. The maximum number of binding events per region. If the value is 0, the maximum number is automatically calculated.

tol: a numeric, default=1e-4. The tolerance for the EM algorithm.

B: an integer, default=100. The maximum number of iterations to be used.

mSelect: a character string specifying the information criteria to be used when selecting the number of binding events. Default="BIC"

mergePeaks: a logical stating whether overlapping binding events should be picked. Default=TRUE

mapCorrect: a logical stating whether mappability profiles should be incorporated in the estimation, i.e. missing reads estimated. Default=TRUE

paraPrior

A list of parameters for the prior distribution as returned by the setParaPrior function. The default parameters should be good enough for most usages.

xi: an integer, default=200. The average DNA fragment size.

rho: an integer, default=1. A variance parameter for the average DNA fragment size distribution.

alpha: an integer, default=20. First hyperparameter of the inverse Gamma distribution for sigma^2 in the PICS model

pics 5

beta: an integer, default=40000. Second hyperparameter of the inverse Gamma distribution for sigma^2 in the PING model

lambda: an integer, default=0. The precision of the prior for mu used for histone data.

dMu: an integer, default=0. Our best guess for the distance between two neighboring nucleosomes.

nCores

An integer. The number of cores that should be used in parallel by the function

#### Methods

**code** signature(x = ``pics''): return the error code for each list element (i.e. candidate region) of a PICS object. If the string is empty, there were no errors.

plot signature(x = ``pics''): Plot all regions in the PICS object. This might be long, and should only be used to plot a few regions, so subset the object before plotting.

**sigmaSqR** signature(x = ``pics''): return the variance parameter of the reverse (R) distribution for each binding event.

**sigmaSqF** signature(x = ``pics''): return the variance parameter of the forward (F) distribution for each binding event.

score signature(x = ``pics''): return the score for each binding event.

**scoreF** signature(x = ``pics''): return the score of the forward (F) for each binding event.

**scoreR** signature(x = ``pics''): return the score of the forward (R) for each binding event.

maxRange signature(x = ``pics''): return the range maximum.

**minRange** signature(x = ``pics''): return the range minimal.

**K** signature(x = ``pics''): subset PICS object.

wigDensity signature(x = ``pics''): return the density for each binding event.

# Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

pics

6 pics-class

pics-class

The pics class

# **Description**

This object is used to gather all parameters from fitting PICS to a single candidate region. The objet contains the following slots: 'estimates', 'infMat', 'Nmerged', 'converge', 'chr'. 'estimates' is a list containing all parameters estimates as well as standard errors. 'infMat' is the Cholesky decomposition of the information matrix, 'converge' is a logical value indicating whether the EM algorithm has converged, while 'chr' is a character string corresponding to a candidate region's chromosome. 'Nmerged' gives the number of binding events that were merged; binding events that overlap are merged (see the cited paper below for details).

#### Accessors

The PICS package provide accessors to directly access to most of the parameters/standard errors and chromosome. In the code snippets below, 'x' is a 'pics' object.

'chromosome(x)' Gets the chromosome name of the candidate region.

'mu(x)' Gets the position estimates of all binding sites identified in the region.

'delta(x)' Gets the average fragment lengths of all binding sites identified in the region.

'sigmaSqF(x)' Gets the F peak variances of all binding sites identified in the region.

'sigmaSq $\mathbf{R}(\mathbf{x})$ ' Gets the R peak variances of all binding sites identified in the region.

'se $\mathbf{F}(\mathbf{x})$ ' Gets the standard errors of all binding site position estimates identified in the region.

'se $\mathbf{F}(\mathbf{x})$ ' Gets the standard errors of all F peak modes identified in the region.

'seR(x)' Gets the standard errors of all R peak modes identified in the region.

**score** signature(x = "pics"): return the score for each binding event.

**scoreF** signature(x = "pics"): return the score of the forward (F) for each binding event.

**scoreR** signature(x = "pics"): return the score of the forward (R) for each binding event.

#### Constructor

newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,chr) construct a new 'pics' object with the following arguments:

w The mixture weights (a vector)

**mu** The binding site positions (a vector)

**delta** The DNA fragment lengths (a vector)

**sigmaSqF** The variance parameters for the forward distribution (vector)

sigmaSqR The variance parameters for the forward distribution (vector)

**seMu** The standard errors for mu (vector)

**seMuF** The standard errors for muF (vector)

seMuR The standard errors for muR (vector)

seMuR The standard errors for muR (vector)

**score** The scores for each binding event (vector)

**Nmerged** The number of peaks that got merged (integer)

converge A logical value, TRUE, if the EM as converged

infMat The information matrix

**chr** The chromosome for the region

picsError-class 7

#### Author(s)

 $Xuekui\ Zhang,\ Arnaud\ Droit\ << arnaud\ .\ droit\ @crchuq\ .\ ualaval\ .\ ca>> and\ Raphael\ Gottardo\ << rgottard\ @fhcrc\ .\ orgottard\ arnaud\ .$ 

#### References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

```
pics picsError
```

## **Examples**

```
# Here is an example of how to construct such a region.
# Typically, you would not do this manually, you would use the pics function to return a 'picsList' that contains
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<-10
seMuF<-10
seMuR<-10
score<-5
Nmerged<-0
converge<-TRUE
chr<-"chr1"
range<-c(1000,2000)
# Contructor
#myPICS<-newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,as.integer(range),chr)</pre>
```

picsError-class

The pics class

# Description

This object is used to return an error code when the PICS function failed to return a valid set of estimates for a candidate regions. This could be due to non-convergence of the EM algorithm, a singular information matrix, or a number of reads below the limit specified by the user. All of these are typically due to too few reads in the region and do not affect the rest of the analysis, as such regions would most likely be labelled as false positives.

#### Accessors

All of the accessors defined for a 'pics' object still work for a 'picsError' object but will simply return a NULL pointer.

# Constructor

newPicsError(string) where 'string' is the error code.

8 picsFDR

#### Constructor

```
newPicsError<-function(string)
string The mixture weights (a vector)</pre>
```

## Author(s)

 $Xuekui\ Zhang,\ Arnaud\ Droit\ << arnaud\ .\ droit\ @crchuq\ .\ ualaval\ .\ ca>> and\ Raphael\ Gottardo\ << rgottard\ @fhcrc\ .\ orgottard\ arnaud\ .$ 

#### References

```
X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.
```

## See Also

pics

# **Examples**

```
# Here is an example on how to construct such a picsError object
# Typically, you would not do this manually, you would use the pics function to return a 'picsList' that contains
# Contructor
myPicsError<-newPicsError("Singular information matrix")
# Accessors
# Get the standard error of Mu
se(myPicsError)
# Get the standard error of MuF
seF(myPicsError)
# Get the scores
score(myPicsError)</pre>
```

picsFDR

Estimate the FDR.

#### **Description**

Estimate the false detection rate for an object of class pics or picsList.

## Usage

```
picsFDR(picsIP,picsCont,filter=list(delta=c(0,Inf),se=c(0,Inf),sigmaSqF=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),sigmaSqR=c(0,Inf),s
```

# **Arguments**

picsIP An object of class pics or picsList containing the informations for the IP

reads.

picsCont An object of class pics or picsList containing the informations for the control

reads.

filter filterA list of ranges for filtering regions based on PICS parameters. By default

filter is set to 'NULL' and all regions are used.

delta Length of the binding sites.

picsList-class 9

se Standard error.sigmaSqF Forward peak variancesigmaSqR Reverse peak variance

#### Value

A 3 columns data. frame with the following columns: FDR, score, N.

#### Author(s)

Xuekui Zhang

#### See Also

picsList pics

picsList-class

The pics class

#### **Description**

This object is used to gather all parameters from fitting PICS to multiple candidate regions (as returned by the 'segmentReads' function). The objet contains the following slots: 'List', 'paraPrior', 'paraEM', 'minReads', 'N', 'Nc'. 'List' is a list of 'pics' or 'picsError' objects. 'paraPrior' is a list containing the hyperparameters used for the prior, 'paraEM' is a list of convergence parameters for the EM, 'minReads' is a list containing the minimum number of reads used to fit a region with 'PICS', 'N' is the total number of reads in the ChIP samples while 'Nc' is the total number of reads in the control sample.

#### **Arguments**

object

An object of class pics.

#### Accessors

The PICS package provide accessors to directly access to most of the parameters/standard errors and chromosomes. In the code snippets below, 'x' is a 'picsList' object. For all accessors, the 'picsError' objects are omitted, so that the accessors only return values for the 'pics' objects (i.e. all valid binding events).

'chromosome(x)' Gets the chromosome names of all candidate regions.

'mu(x)' Gets the position estimates of all binding sites identified in all candidate regions.

'delta(x)' Gets the average fragment lengths of all binding sites identified in all candidate regions.

'sigma $\operatorname{SqF}(x)$ ' Gets the F peak variances of all binding sites identified in all candidate regions.

'sigmaSqR(x)' Gets the R peak variances of all binding sites identified in all candidate regions.

'seF(x)' Gets the standard errors of all binding site position estimates identified in all candidate regions.

 $\mathbf{seF}(\mathbf{x})$  Gets the standard errors of all F peak modes identified in all candidate regions.

'seR(x)' Gets the standard errors of all R peak modes identified in all candidate regions.

'score(x)' Gets the scores of all binding events identified in all candidate regions.

10 picsList-class

#### Constructor

```
newPicsList(List, paraEM, paraPrior, minReads, N, Nc)

List The mixture weights (a vector)

paraEM The binding site positions (a vector)

paraPrior The DNA fragment lengths (a vector)

N The variance parameters for the forward distribution (vector)
```

Nc The variance parameters for the forward distribution (vector)

## Methods

```
[ signature(x = ``pics''): subset PICS object.
```

# Methods

```
length signature(x = ``pics''): subset PICS object.
```

#### Constructor

newPicsList<-function(List, paraEM, paraPrior, minReads, N, Nc) constructs a new 'picsList' object with the following arguments.

## newPicsList

```
w The mixture weights (a vector)
mu The binding site positions (a vector)
delta The DNA fragment lengths (a vector)
sigmaSqF The variance parameters for the forward distribution (vector)
sigmaSqR The variance parameters for the reverse distribution (vector)
seMu The standard errors for mu (vector)
seMuF The standard errors for muF (vector)
seMuR The standard errors for muR (vector)
seOur The scores for each binding event (vector)
Nmerged The number of peaks that were merged (integer)
converge A logical value, TRUE, if the EM as converged
infMat The information matrix
chr The chromosome for the region
```

## Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

plot-FDR 11

#### See Also

pics

# **Examples**

```
# Here is an example of how to construct such a region
# Typically, you would not do this manually, you would use the pics function to return a 'picsList' that contains
w<-1
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<-10
seMuF<-10
seMuR<-10
score<-5
Nmerged<-0
converge<-TRUE
infMat<-matrix(0)</pre>
chr<-"chr1"
range<-c(1000,2000)
# Contructor
#myPICS1<-newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,as.integer(reference)
#myPICS2<-newPics(w,mu+1000,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,as.integ</pre>
#minReads<-list(perPeak=2,perRegion=5)</pre>
#paraPrior<-list(xi=200,rho=1,alpha=20,beta=40000)</pre>
#paraEM<-list(minK=1,maxK=15,tol=10e-6,B=100)</pre>
#N<-100
#Nc<-200
#mynewPicsList<-newPicsList(list(myPICS1,myPICS2), paraEM, paraPrior, minReads, as.integer(100), as.integer(
# Accessors
# Get the standard error of Mu
#se(mynewPicsList)
# Get the standard error of MuF
#seF(mynewPicsList)
# Get the scores
#score(mynewPicsList)
```

plot-FDR

FDR plot for PICS

# **Description**

This method plots an FDR curve showing the FDR as a function of the PICS scores.

# Usage

```
## S4 method for signature 'picsList,picsList'
plot(x, y, filter=NULL, h=.1, ...)
```

12 segmentPICS

## **Arguments**

X	A picsList object as returned by the function PICS run on the treatment data.
у	A picsList object as returned by the function PICS run on the control data.
filter	A list of ranges for filtering regions based on PICS parameters. By default filter is set to 'NULL' and all regions are used.
h	A value between 0 and 1, representing the desired FDR. This simply draws a horizontal line at the given value.
	Further graphical parameters passed to the generic function plot.

# Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq," Biometrics, iss. In press, 2010.

# See Also

**PICS** 

segChrRead	Segmentation of paired-end sequencing data	

# Description

These two functions are part of the segmentation step for paired-end sequencing data and are exported to be used in PING package.

segme	entPICS	Segment the genome into candidate regions

# Description

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

# Usage

```
segmentPICS(data, dataC=NULL, map=NULL, minReads=2, minReadsInRegion=3,
    jitter=FALSE, dataType="TF", maxLregion=0, minLregion=100)
```

segmentPICS 13

## **Arguments**

data A linkS4class{GRanges} object containing the IP reads. See details for more

information on how to set up the data.

dataC A linkS4class{GRanges} object containing the control reads. Set to NULL by

default, i.e. no control.

map A 'RangedData' object containing the mappability profiles. Set to NULL by

default, i.e. no profiles.

minReads The minimum number of F/R reads to be present in the sliding window.

minReadsInRegion

The minimum number of F/R reads to be present in the region.

jitter A logical value stating whether some noise should be added to the read locations.

This is recommended if the read positions have lots of duplicates.

dataType Type of experiment. "TF" or "H".

maxLregion The maximum length.

minLregion The minimum length.

#### Value

An object of class segReadsList containing the results for all regions pre-processed.

seg<-segmentPICS(dataIP, dataC=dataCont, map=map, minReads=1)</pre>

#### Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

#### References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009.

# See Also

```
segReadsList
```

# **Examples**

```
# Read data
path<-system.file("extdata",package="PICS")
## Note that the col name for the chromosome needs to be space and not chr
dataIP<-read.table(file.path(path, "Treatment_tags_chr21_sort.bed"),header=TRUE,colClasses=c("factor","integ
dataIP<-as(dataIP, "GRanges")

dataCont<-read.table(file.path(path, "Input_tags_chr21_sort.bed"),header=TRUE,colClasses=c("factor","integer
dataCont<-as(dataCont, "GRanges")

map<-read.table(file.path(path, "mapProfileShort"),header=TRUE,colClasses=c("factor","integer","integer","NU
map<-as(map, "GRanges")</pre>
```

14 segReadsList

segReads

Segment the genome into candidate regions

# **Description**

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

# Methods

```
map signature(x = ``pics''): subset PICS object.
```

## Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

pics

segReadsList

Segment the genome into candidate regions

# **Description**

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

# Methods

```
[ signature(x = ``pics''): subset gadem object.
[[ signature(x = ``pics''): subset gadem object.
```

#### Methods

```
length signature(x = ``pics''): subset PICS object.
```

## Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

segReadsListPE 15

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

pics

segReadsListPE

List of segReadsPE objects

# Description

A list of segReadsPE. The class also store information related to the segmentation process, keeping a trace of the parameters used and the proportion of forward and reverse reads for the input and the control.

## Methods

```
[ signature(x = ``pics''): subset gadem object.
[[ signature(x = ``pics''): subset gadem object.
```

## **Extends**

Class segReadsList, directly.

# Author(s)

Xuekui Zhang

## See Also

segReadsPE segReadsList

segReadsPE

Class to store post-segmentation result

# Description

This class stores the information of the segmentation performed by segmentPING. It is used as the input of the PING function.

# Extends

Class segReadsList, directly.

## Author(s)

Xuekui Zhang

16 setParaEM

## See Also

segReads segReadsListPE

setParaEM	Function that returns a list of parameters for the EM algorithm that can be used as an argument of PICS.
	control insert an arran of 1102.

# Description

This function takes from 0 to 7 EM algorithm parameters as argument, check if they are valid and returns a list to be used in a call to PICS.

# Usage

setParaEM(minK=1,maxK=15,tol=1e-4,B=100,mSelect="BIC",mergePeaks=TRUE,mapCorrect=TRUE,dataType=

# Arguments

minK	An integer. The minimum number of binding events per region. If the value is 0, the minimum number is automatically calculated.
maxK	An integer. The maximum number of binding events per region. If the value is 0, the maximum number is automatically calculated.
tol	A numeric. The tolerance for the EM algorithm.
В	An integer. The maximum number of iterations to be used.
mSelect	A character string specifying the information criteria to be used when selecting the number of binding events.
mergePeaks	A logical stating whether overlapping binding events should be picked.
mapCorrect	A logical stating whether mappability profiles should be incorporated in the estimation, i.e: missing reads estimated.
dataType	A character. If a dataType is set, the algorithm will use the default parameters for this type of data (all the previous arguments will be ignored).

# Value

Returns a list of parameters to be used in PICS.

# Author(s)

Renan Sauteraud

# See Also

PICS

setParaPrior 17

setParaPrior	Function that returns a list of parameters that can be used as an argument of PICS.

# Description

This function takes from 0 to 6 parameters as argument, check if they are valid and returns a list to be used in a call to PICS.

# Usage

```
setParaPrior(xi=200,rho=1,alpha=20,beta=40000,lambda=0,dMu=0, dataType=NULL, PExi=0)
```

# Arguments

xi	An integer. The average DNA fragment size.
rho	An integer. A variance parameter for the average DNA fragment size distribution.
alpha	An integer. First hyperparameter of the inverse Gamma distribution for sigma^2 in the PICS model
beta	An integer. Second hyperparameter of the inverse Gamma distribution for sigma^2 in the PICS model
lambda	An integer. The precision of the prior for mu used for histone data.
dMu	An integer. Our best guess for the distance between two neighboring nucleosomes.
dataType	A character string. If a valid dataType is specified, use our suggested parameters. "MNase" or "sonicated"
PExi	A numeric. With paired end data, 'xi' can be calculated directly from the reads. If PExi is set, it will overwrite the xi determined by the dataType.

# Value

Returns a list of 6 parameters to be used in PICS.

# Author(s)

Renan Sauteraud

# See Also

PICS

# **Examples**

```
# set prior for PICS data
paraPrior<-setParaPrior()
# set prior for sonicated data using our selected default parameters
paraPrior<-setParaPrior(dataType="sonicated")</pre>
```

18 summary

show show PICS

# Description

This methods show the objects of PICS

# Usage

```
## S4 method for signature 'pics'
show(object)
## S4 method for signature 'picsError'
show(object)
## S4 method for signature 'picsList'
show(object)
## S4 method for signature 'segReads'
show(object)
## S4 method for signature 'segReadsList'
show(object)
```

# **Arguments**

object Object returned from pics.

## **Details**

List of the slots include in the object

# Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

# See Also

summary

summary

summary PICS

# Description

This methods summarized 'pics', 'picsList', 'seg' or 'segList' objects.

summarySeg 19

# Usage

```
## S4 method for signature 'pics'
summary(object)
## S4 method for signature 'picsList'
summary(object)
## S4 method for signature 'segReads'
summary(object)
## S4 method for signature 'segReadsList'
summary(object)
```

# **Arguments**

object

Object returned from pics.

## Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@crchuq.ualaval.ca>> and Raphael Gottardo <<rgottard@fhcrc.org

#### See Also

show

summarySeg

Summarize a segReadsList object.

# Description

Returns info about a segReadsList object in a data.frame containing the following informations: chr: chromosome id NF: number of forward reads NR: number of reverse reads L: length of segment min: start location of segments max: end location of segments

# Usage

```
summarySeg(seg)
```

# **Arguments**

seg

An object of class segReadsList

#### Value

A six columns data. frame.

# Author(s)

Xuekui Zhang

# See Also

```
segReadsList
```

# Index

*Topic <b>classes</b>	[[,segReadsListPE,ANY,ANY-method
segReadsListPE, 15	(segReadsListPE), 15
segReadsPE, 15	%in%, ANY, segReads-method(segReads), 14
*Topic data	
makeRangedDataOutput, 3	as.list,AlignedRead-method(pics), $4$
*Topic functions	
bam2gr, 2	bam2gr, 2
picsFDR, 8	
segmentPICS, 12	candidate.region(segChrRead), 12
setParaEM, 16	chromosome (pics-class), 6
setParaPrior, 17	chromosome, data. frame-method
summarySeg, 19	(pics-class), 6
*Topic graphs	chromosome, pics-method (pics-class), 6
plot-FDR, 11	chromosome,picsError-method
*Topic models	(picsError-class), 7
pics, 4	chromosome,picsList-method
pics-class, 6	(picsList-class), 9
picsError-class, 7	code (pics), 4
picsList-class, 9	code, pics-method (pics), 4
segReads, 14	code, picsError-method (pics), 4
segReadsList, 14	code, picsList-method (pics), 4
*Topic <b>print</b>	coerce (pics), 4
show, 18	coerce, AlignedRead, GenomeData-method
summary, 18	(pics), 4
*Topic <b>process</b>	coerce, data.frame, GenomeData-method
makeRangedDataOutput, 3	<pre>(pics), 4 coerce, data.frame, GRanges-method</pre>
[,picsList,ANY,ANY-method	(pics), 4
(picsList-class), 9	coerce,data.frame,picsList-method
[,picsList-method(picsList-class),9	(pics), 4
[,segReadsList,ANY,ANY-method	coerce,picsList,data.frame-method
(segReadsList), 14	(pics), 4
[,segReadsList-method(segReadsList), 14	coerce,picsList,RangedData-method
[,segReadsListPE,ANY,ANY-method	(pics), 4
(segReadsListPE), 15	coerce, RangedData, GenomeData-method
[,segReadsListPE-method	(pics), 4
(segReadsListPE), 15	(р163), 1
[[,picsList,ANY,ANY-method	delta(pics-class), 6
(picsList-class), 9	delta, data. frame-method (pics-class), 6
[[,picsList-method(picsList-class),9	delta, pics-method (pics-class), 6
[[,segReadsList,ANY,ANY-method	delta,picsError-method
(segReadsList), 14	(picsError-class), 7
[[,segReadsList-method(segReadsList),	delta,picsList-method(picsList-class)
1/1	0

INDEX 21

density,picsList-method	picsList-class, 9
(picsList-class), 9	<pre>plot,pics,segReads-method(pics),4</pre>
	<pre>plot,picsError,segReads-method(pics),4</pre>
FDR (plot-FDR), 11	<pre>plot,picsList,picsList-method</pre>
	(plot-FDR), 11
K (pics), 4	plot,picsList,segReadsList-method
K, pics-method (pics), 4	(pics), 4
K, picsError-method (pics), 4	plot-FDR, 11
K,picsList-method(pics),4	prot TDN, TT
length (picsList-class), 9	score (pics), 4
length,picsList-method	score, data.frame-method (pics-class), 6
(picsList-class), 9	score, pics-method (pics-class), 6
length,segReadsList-method	score,picsError-method
(segReadsList), 14	(picsError-class), 7
(368,164432136), 11	<pre>score,picsList-method(picsList-class),</pre>
makeRangedDataOutput, 3	9
map (segReads), 14	scoreForward (pics), 4
map, segReads-method (segReads), 14	scoreForward, data.frame-method (pics), 4
map, segReadsList-method (segReads), 14	scoreForward, pics-method (pics), 4
maxRange (pics), 4	scoreForward, picsError-method (pics), 4
maxRange, pics-method (pics), 4	
maxRange, picsError-method (pics), 4	scoreForward, picsList-method (pics), 4
maxRange, picsList-method (pics), 4	scoreReverse (pics), 4
minRange (pics), 4	scoreReverse, data.frame-method (pics), 4
	scoreReverse, pics-method (pics), 4
minRange, pics-method (pics), 4	scoreReverse, picsError-method (pics), 4
minRange, picsError-method (pics), 4	<pre>scoreReverse,picsList-method(pics), 4</pre>
minRange, picsList-method (pics), 4	se (pics-class), 6
mu (pics-class), 6	se, data. frame-method (pics-class), $6$
mu, data.frame-method (pics-class), 6	se,pics-method(pics-class), 6
mu, pics-method (pics-class), 6	se,picsError-method(picsError-class),7
mu, picsError-method (picsError-class), 7	<pre>se,picsList-method(picsList-class),9</pre>
<pre>mu,picsList-method(picsList-class), 9</pre>	seF (pics-class), 6
no Dio (nico alora) (	seF, data.frame-method(pics-class), 6
newPics (pics-class), 6	seF, pics-method (pics-class), 6
newPics, pics-method (pics-class), 6	<pre>seF,picsError-method(picsError-class),</pre>
newPicsError (picsError-class), 7	7
newPicsError,picsError-method	<pre>seF,picsList-method(picsList-class),9</pre>
(picsError-class), 7	segChrRead, 12
newPicsList (picsList-class), 9	segmentPICS, 2, 12
newPicsList,picsList-method	segReads, 14, <i>16</i>
(picsList-class), 9	segReads-class (segReads), 14
DICC 12	segReadsGeneric (segmentPICS), 12
PICS, 12	segReadsList, 13, 14, 15, 19
PICS (pics), 4	
pics, 4, 5, 7–9, 11, 14, 15, 18, 19	segReadsList-class (segReadsList), 14
pics-class, 6	segReadsListPE, 15, 16
pics-class (pics), 4	<pre>segReadsListPE-class(segReadsListPE),</pre>
picsError, 7	15
picsError (picsError-class), 7	segReadsPE, 15, 15
picsError-class, 7	segReadsPE-class (segReadsPE), 15
picsFDR, 8	seR (pics-class), 6
picsList, 9	seR, data.frame-method(pics-class), 6
picsList (picsList-class), 9	seR, pics-method (pics-class), 6

22 INDEX

```
seR, picsError-method (picsError-class),
seR,picsList-method(picsList-class), 9
setParaEM, 16
setParaPrior, 17
show, 18, 19
show, pics-method (show), 18
show, picsError-method (show), 18
show, picsList-method (show), 18
show, segReads-method (show), 18
show, segReadsList-method (show), 18
show-methods (show), 18
sigmaSqF (pics), 4
sigmaSqF, data.frame-method(pics), 4
sigmaSqF, pics-method (pics), 4
sigmaSqF,picsError-method(pics), 4
sigmaSqF,picsList-method(pics), 4
sigmaSqR (pics), 4
sigmaSqR,data.frame-method(pics),4
sigmaSqR, pics-method (pics), 4
sigmaSqR,picsError-method(pics), 4
sigmaSqR,picsList-method(pics), 4
summary, 18, 18
summary, pics-method (summary), 18
summary, picsList-method (summary), 18
summary, segReads-method (summary), 18
summary, segReadsList-method (summary),
        18
summary-methods (summary), 18
summarySeg, 19
w (pics-class), 6
w, data.frame-method(pics-class), 6
w, pics-method (pics-class), 6
w,picsError-method(picsError-class), 7
w,picsList-method(picsList-class), 9
wigDensity (pics), 4
wigDensity, pics-method (pics), 4
wigDensity, picsError-method (pics), 4
wigDensity, picsList-method (pics), 4
```