

Introduction to RBM package

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May 2, 2019

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1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 37

> which(myresult$permutation_p<=0.05)

[1] 32 116 121 188 191 205 223 243 262 282 287 293 306 353 376 383 396 423 450
[20] 489 509 525 550 558 570 629 635 654 669 712 837 860 879 914 951 976 997

> sum(myresult$bootstrap_p<=0.05)

[1] 12

> which(myresult$bootstrap_p<=0.05)

[1] 223 236 287 295 376 469 581 627 657 818 837 976

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 4

> which(myresult2$bootstrap_p<=0.05)

[1] 37 152 374 477

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1   3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p    3000   -none- numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 51

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 59

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 62

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 7 20 29 59 107 121 122 142 146 165 180 184 186 190 215 221 245 271 340
[20] 352 360 386 392 445 478 480 511 514 529 532 565 569 616 635 667 668 703 725
[39] 739 755 765 771 783 804 898 902 910 947 966 976 982

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 7 16 20 29 59 75 95 107 116 121 122 142 146 158 165 180 184 186 190
[20] 215 221 226 227 245 248 255 271 296 340 352 360 440 445 478 480 511 514 529
[39] 532 537 549 565 569 572 667 668 703 725 739 765 771 783 804 893 898 902 947
[58] 976 982

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 7 16 29 54 59 107 113 121 122 142 146 165 180 184 186 190 215 221 226
[20] 227 245 248 255 296 316 340 352 360 389 440 445 478 480 511 514 529 532 537
[39] 549 558 562 565 569 589 616 620 635 668 697 703 725 739 765 771 783 804 898
[58] 902 910 947 976 982

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 7

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 15

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 13

> which(con2_adjp<=0.05/3)

[1] 29 107 121 142 184 215 340 360 511 529 725 739 771 783 902

> which(con3_adjp<=0.05/3)

[1] 29 142 184 215 340 360 668 703 725 771 783 804 898

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000    -none- numeric
ordfit_pvalue 3000    -none- numeric
ordfit_beta1  3000    -none- numeric
permutation_p 3000    -none- numeric
bootstrap_p   3000    -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 52

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 49

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 53

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```
[1] 30 32 39 51 98 111 120 155 174 175 209 250 251 261 295 322 326 340 350
[20] 353 365 379 380 390 391 392 403 412 434 458 479 499 536 577 583 603 661 667
[39] 768 773 803 804 883 889 896 897 906 908 945 946 954 976
```

```
> which(myresult2_F$bootstrap_p[, 2]<=0.05)
```

```
[1] 30 32 39 56 82 98 120 155 174 175 209 250 251 261 267 295 322 335 350
[20] 353 365 379 380 390 391 392 397 403 412 434 479 499 655 661 667 773 780 803
[39] 827 847 889 896 897 906 908 945 954 961 976
```

```
> which(myresult2_F$bootstrap_p[, 3]<=0.05)
```

```
[1] 30 32 39 51 60 82 98 120 158 174 175 199 209 250 251 261 322 337 350
[20] 353 365 379 380 390 391 392 403 412 433 434 479 499 577 583 587 606 619 667
[39] 748 773 780 803 847 865 889 892 896 897 906 945 946 954 976
```

```
> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)
```

```
[1] 6
```

```
> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)
```

```
[1] 8
```

```
> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)
```

```
[1] 11
```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "C:/Users/biocbuild/bbs-3.9-bioc/tmpdir/RtmpyuvsfZ/Rinst24843ecd317/RBM/data"
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		


```
exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268
```

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 45
```

```
> sum(diff_results$permutation_p<=0.05)
```

```

[1] 56

> sum(diff_results$bootstrap_p<=0.05)

[1] 60

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 1

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 1

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2] exmdata5[, 2]
851 cg00830029 0.583625      0.5939787      0.6473961      0.6726964      0.5082024
      exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
851      0.3465747      0.6627657      0.6463451
diff_results$ordfit_t[diff_list_perm]
851
diff_results$permutation_p[diff_list_perm]
851      0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
252 cg00230502 0.1006139      0.1351787      0.1253851      0.1630492
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
252      0.1197087      0.1203616      0.1742373      0.1815548
diff_results$ordfit_t[diff_list_boot]
252
diff_results$bootstrap_p[diff_list_boot]
252      0

```