

# Additional plots for: Independent filtering increases power for detecting differentially expressed genes, Bourgon et al., PNAS (2010)

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*genefilter* version 1.64.0 (Last revision )

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## 1 Introduction

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This vignette illustrates use of some functions in the *genefilter* package that provide useful diagnostics for independent filtering [1]:

- `kappa_p` and `kappa_t`
- `filtered_p` and `filtered_R`
- `filter_volcano`
- `rejection_plot`

## 2 Data preparation

---

Load the ALL data set and the *genefilter* package:

```
library("genefilter")
library("ALL")
data("ALL")
```

Reduce to just two conditions, then take a small subset of arrays from these, with 3 arrays per condition:

```
bcell <- grep("^B", as.character(ALL$BT))
moltyp <- which(as.character(ALL$mol.biol) %in%
               c("NEG", "BCR/ABL"))
ALL_bcrneg <- ALL[, intersect(bcell, moltyp)]
ALL_bcrneg$mol.biol <- factor(ALL_bcrneg$mol.biol)
n1 <- n2 <- 3
set.seed(1969)
use <- unlist(tapply(1:ncol(ALL_bcrneg),
                    ALL_bcrneg$mol.biol, sample, n1))
subsample <- ALL_bcrneg[,use]
```

We now use functions from *genefilter* to compute overall standard deviation filter statistics as well as standard two-sample *t* and related statistics.

```
S <- rowSds( exprs( subsample ) )
temp <- rowttests( subsample, subsample$mol.biol )
d <- temp$dm
p <- temp$p.value
t <- temp$statistic
```

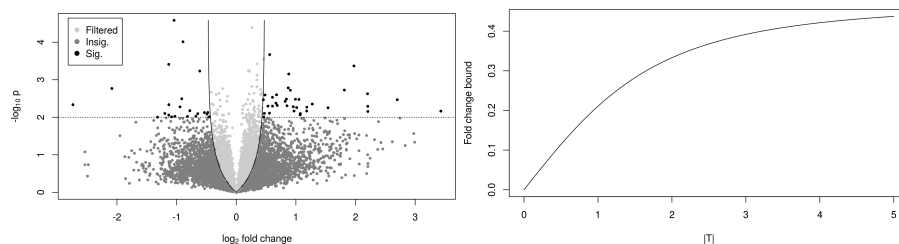
### 3 Filtering volcano plot

Filtering on overall standard deviation and then using a standard *t*-statistic induces a lower bound of fold change, albeit one which varies somewhat with the significance of the *t*-statistic. The `filter_volcano` function allows you to visualize this effect.

The output is shown in the left panel of Fig. 1.

The `kappa_p` and `kappa_t` functions, used to make the volcano plot, compute the fold change bound multiplier as a function of either a *t*-test *p*-value or the *t*-statistic itself. The actual induced bound on the fold change is  $\kappa$  times the filter's cutoff on the overall standard deviation. Note that fold change bounds for values of  $|T|$  which are close to 0 are not of practical interest because we will not reject the null hypothesis with test statistics in this range.

The plot is shown in the right panel of Fig. 1.



**Figure 1:** Left panel: plot produced by the `filter_volcano` function  
Right panel: graph of the `kappa_t` function.

## 4 Rejection count plots

### 4.1 Across $p$ -value cutoffs

The `filtered_p` function permits easy simultaneous calculation of unadjusted or adjusted  $p$ -values over a range of filtering thresholds ( $\theta$ ). Here, we return to the full “BCR/ABL” versus “NEG” data set, and compute adjusted  $p$ -values using the method of Benjamini and Hochberg, for a range of different filter stringencies.

```
table(ALL_bcrneg$mol.biol)
```

```
##
## BCR/ABL    NEG
##      37     42
```

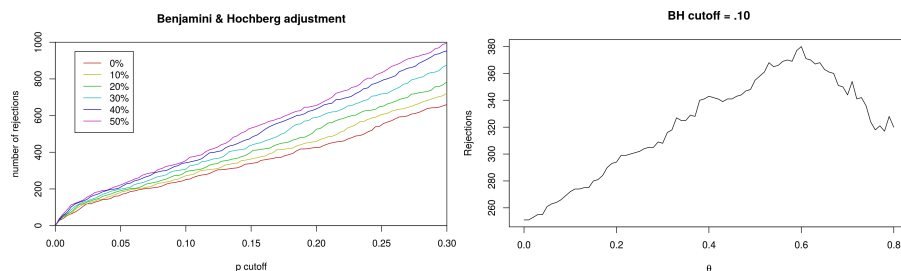
```
S2 <- rowVars(exprs(ALL_bcrneg))
p2 <- rowttests(ALL_bcrneg, "mol.biol")$p.value
theta <- seq(0, .5, .1)
p_bh <- filtered_p(S2, p2, theta, method="BH")
```

```
head(p_bh)
```

```
##      0%    10%   20%   30%   40%   50%
## [1,] 0.919 0.894 0.862 0.828    NA    NA
## [2,] 0.959 0.946 0.930 0.906 0.887 0.871
## [3,] 0.702    NA    NA    NA    NA    NA
## [4,] 0.981 0.975 0.968 0.957    NA    NA
## [5,] 0.951 0.935 0.912 0.884    NA    NA
## [6,] 0.634 0.590 0.544 0.495 0.450 0.410
```

The `rejection_plot` function takes sets of  $p$ -values corresponding to different filtering choices — in the columns of a matrix or in a list — and shows how rejection count ( $R$ ) relates to the choice of cutoff for the  $p$ -values. For these data, over a reasonable range of FDR cutoffs, increased filtering corresponds to increased rejections.

```
rejection_plot(p_bh, at="sample",
               xlim=c(0, .3), ylim=c(0,1000),
               main="Benjamini & Hochberg adjustment")
```



**Figure 2:** Left panel: plot produced by the `rejection_plot` function  
Right panel: graph of `theta`.

The plot is shown in the left panel of Fig. 2.

## 4.2 Across filtering fractions

If we select a fixed cutoff for the adjusted  $p$ -values, we can also look more closely at the relationship between the fraction of null hypotheses filtered and the total number of discoveries. The `filtered_R` function wraps `filtered_p` and just returns rejection counts. It requires a  $p$ -value cutoff.

```
theta <- seq(0, .80, .01)
R_BH <- filtered_R(alpha=.10, S2, p2, theta, method="BH")
```

```
head(R_BH)
## 0% 1% 2% 3% 4% 5%
## 251 251 253 255 255 261
```

Because overfiltering (or use of a filter which is inappropriate for the application domain) discards both false and true null hypotheses, very large values of  $\theta$  reduce power in this example:

```
plot(theta, R_BH, type="l",
      xlab=expression(theta), ylab="Rejections",
      main="BH cutoff = .10"
      )
```

The plot is shown in the right panel of Fig. 2.

## Session information

- R version 3.5.1 Patched (2018-07-12 r74967), x86\_64-pc-linux-gnu
- Running under: Ubuntu 16.04.5 LTS
- Matrix products: default
- BLAS: /home/biocbuild/bbs-3.8-bioc/R/lib/libRblas.so
- LAPACK: /home/biocbuild/bbs-3.8-bioc/R/lib/libRlapack.so
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, utils
- Other packages: ALL 1.23.0, Biobase 2.42.0, BiocGenerics 0.28.0, DESeq 1.34.0, RColorBrewer 1.1-2, genefilter 1.64.0, knitr 1.20, lattice 0.20-35, locfit 1.5-9.1, pasilla 1.9.0
- Loaded via a namespace (and not attached): AnnotationDbi 1.44.0, BiocManager 1.30.3, BiocStyle 2.10.0, DBI 1.0.0, IRanges 2.16.0, Matrix 1.2-14, RCurl 1.95-4.11, RSQLite 2.1.1, Rcpp 0.12.19, S4Vectors 0.20.0, XML 3.98-1.16, annotate 1.60.0, backports 1.1.2, bit 1.1-14, bit64 0.9-7, bitops 1.0-6, blob 1.1.1, codetools 0.2-15, compiler 3.5.1, digest 0.6.18, evaluate 0.12, geneplotter 1.60.0,

grid 3.5.1, highr 0.7, htmltools 0.3.6, magrittr 1.5, memoise 1.1.0, rmarkdown 1.10,  
rprojroot 1.3-2, splines 3.5.1, stats4 3.5.1, stringi 1.2.4, stringr 1.3.1, survival 2.43-1,  
tools 3.5.1, xtable 1.8-3, yaml 2.2.0

## References

- [1] Richard Bourgon, Robert Gentleman and Wolfgang Huber. Independent filtering increases power for detecting differentially expressed genes.