Package 'isomiRs'

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R topics documented:

`isomiRs-package` *isomiRs*

Description

isomiRs

Author(s)

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counts *Accessors for the count matrix of a IsomirDataSeq object.*

Description

The counts slot holds the count data as a matrix of non-negative integer count values, one row for each isomiR, and one column for each sample. The normalized matrix can be obtained by using the parameter `norm=TRUE`.

Usage

```
counts.IsomirDataSeq(object, norm = FALSE)

## S4 method for signature 'IsomirDataSeq'
counts(object, norm = FALSE)

## S4 replacement method for signature 'IsomirDataSeq,matrix'
counts(object) <- value
```

Arguments

`object`	A `IsomirDataSeq` object.
`norm`	Boolean, return log2-normalized counts.
`value`	An integer matrix.

Value

base::matrix with raw or normalized count data.

Author(s)

Lorena Pantano

Examples

```
data(mirData)
head(counts(mirData))
```

dat286.long *Data frame containing mirna from Argyropoulos's paper*

Description

Argyropoulos, Christos, et al. "Modeling bias and variation in the stochastic processes of small RNA sequencing." Nucleic Acids Research (2017).

Usage

```
dat286.long
```

Format

mirna expression data in long format.

`design` *Accessors for the 'design' slot of a IsomirDataSeq object.*

Description

The design holds the R `formula` which expresses how the counts depend on the variables in `colData`. See IsomirDataSeq for details.

Usage

```
## S4 method for signature 'IsomirDataSeq'
design(object)

## S4 replacement method for signature 'IsomirDataSeq,formula'
design(object) <- value
```

Arguments

`object`	A IsomirDataSeq object.
`value`	A formula to pass to DESeq2.

Value

design for the experiment

Examples

```
data(mirData)
design(mirData) <- formula(~ 1)
```

`findTargets` *Find miRNAs target using mRNA/miRNA expression*

Description

This function creates a matrix with rows (genes) and columns (mirnas) with values indicating if miRNA-gene pair is target according putative targets and negative correlation of the expression of both molecules.

Usage

```
findTargets(mirna_rse, gene_rse, target, summarize = "group",
  min_cor = -0.6)
```

Arguments

mirna_rse	SummarizedExperiment::SummarizedExperiment with miRNA information. See details.
gene_rse	SummarizedExperiment::SummarizedExperiment with gene information. See details.
target	Matrix with miRNAs (columns) and genes (rows) target prediction values (1 if it is a target, 0 if not).
summarize	Character column name in colData(rse) to use to group samples and compare betweem miRNA/gene expression.
min_cor	Numeric cutoff for correlation value that will be use to consider a miRNA-gene pair as valid.

Value

mirna-gene matrix

Examples

```
data(isoExample)
mirna_ma <- matrix(rbinom(20*25, c(0, 1), 1), ncol = 20)
colnames(mirna_ma) <- rownames(mirna_ex_rse)
rownames(mirna_ma) <- rownames(gene_ex_rse)
corMat <- findTargets(mirna_ex_rse, gene_ex_rse, mirna_ma)
```

---

gene_ex_rse *Data frame containing gene expression data*

Description

Data frame containing gene expression data

Usage

```
gene_ex_rse
```

Format

gene expression data with 18 samples: example of a time series data

`isoCorrect` *Correct miRNA expression based on prior ligation bias information*

Description

This is the source file for fitting the linear quadratic normal family

Usage

```
isoCorrect(train, data, cycles = 5000, long = FALSE)
```

Arguments

`train`	Long data.frame to train model.
`data`	Long data.frame to correct abundance.
`cycles`	Number of cycles to reach convergency.
`long`	Boolean if input is in long format instead of standard wide format (rows:miRNAs, columns:samples).

Details

Methods adapted from:

Argyropoulos, Christos, et al. "Modeling bias and variation in the stochastic processes of small RNA sequencing." Nucleic Acids Research (2017).

Value

data.frame with corrected expression

Author(s)

Christos Argyropoulos and Lorena Pantano

Examples

```
options(warn = -1) # this is only for tiny example
data(mirTritation)
ma <- isoCorrect(mirTritation[mirTritation$class=="train",],
mirTritation[mirTritation$class=="test",],cycles=5,long=TRUE)
library(ggplot2)
ggplot(ma,aes(y=log2(reads), x=Dilution)) + geom_jitter()
ggplot(ma,aes(y=m, x=Dilution)) + geom_jitter()
```

isoCounts *Create count matrix with different summarizing options*

Description

This function collapses isomiRs into different groups. It is a similar concept than how to work with gene isoforms. With this function, different changes can be put together into a single miRNA variant. For instance all sequences with variants at 3' end can be considered as different elements in the table or analysis having the following naming `hsa-miR-124a-5p.iso.t3:AAA`.

Usage

```
isoCounts(ids, ref = FALSE, iso5 = FALSE, iso3 = FALSE, add = FALSE,
  subs = FALSE, seed = FALSE, minc = 1, mins = 1)
```

Arguments

`ids`	Object of class IsomirDataSeq.
`ref`	Differentiate reference miRNA from rest.
`iso5`	Differentiate trimming at 5 miRNA from rest.
`iso3`	Differentiate trimming at 3 miRNA from rest.
`add`	Differentiate additions miRNA from rest.
`subs`	Differentiate nt substitution miRNA from rest.
`seed`	Differentiate changes in 2-7 nts from rest.
`minc`	Int minimum number of isomiR sequences to be included.
`mins`	Int minimum number of samples with number of sequences bigger than `minc` counts.

Details

You can merge all isomiRs into miRNAs by calling the function only with the first parameter `isoCounts(ids)`. You can get a table with isomiRs altogether and the reference miRBase sequences by calling the function with `ref=TRUE`. You can get a table with 5' trimming isomiRS, miRBase reference and the rest by calling with `isoCounts(ids, ref=TRUE, iso5=TRUE)`. If you set up all parameters to TRUE, you will get a table for each different sequence mapping to a miRNA (i.e. all isomiRs).

Examples for the naming used for the isomiRs are at http://seqcluster.readthedocs.org/mirna_annotation.html#mirna-annotation.

Value

IsomirDataSeq object with new count table. The count matrix can be access with `counts(ids)`.

Examples

```
data(mirData)
ids <- isoCounts(mirData, ref=TRUE)
head(counts(ids))
# taking into account isomiRs and reference sequence.
ids <- isoCounts(mirData, ref=TRUE, minc=10, mins=6)
head(counts(ids))
```

isoDE *Differential expression analysis with DESeq2*

Description

This function does differential expression analysis with DESeq2::DESeq2-package using the specific formula. It will return a DESeq2::DESeqDataSet object.

Usage

```
isoDE(ids, formula = NULL, ...)
```

Arguments

ids	Object of class IsomirDataSeq.
formula	Formula used for DE analysis.
...	Options to pass to `isoCounts()` including ref, iso5, iso3, add, subs and seed parameters.

Details

First, this function collapses all isomiRs in different types. Read more at `isoCounts()` to know the different options available to collapse isomiRs.

After that, DESeq2::DESeq2-package is used to do differential expression analysis. It uses the count matrix and design experiment stored at (`counts(ids)` and `colData(ids)`) IsomirDataSeq object to construct a DESeq2::DESeqDataSet object.

Value

DESeq2::DESeqDataSet object. To get the differential expression isomiRs, use `DESeq2::results()` from DESeq2 package. This allows to ask for different contrast without calling again `isoDE()`. Read `results` manual to know how to access all the information.

Examples

```
data(mirData)
ids <- isoCounts(mirData, minc=10, mins=6)
dds <- isoDE(mirData, formula=~group)
```

isoLQNO *Differential expression between two groups using LQNO model*

Description

Differential expression between two groups using LQNO model

Usage

```
isoLQNO(counts, groups = NULL, long = FALSE)
```

Arguments

counts	Count matrix.
groups	Character vector to indicate the group of each sample.
long	Whether matrix is in long format. Default FALSE.

Details

Methods adapted from *Argyropoulos et al (2017)*.

Value

data.frame with estimates and p-values.

Author(s)

Christos Argyropoulos and Lorena Pantano

References

Argyropoulos, Christos, et al. "Modeling bias and variation in the stochastic processes of small RNA sequencing." Nucleic Acids Research (2017).

Examples

```
options(warn = -1) # this is only for tiny example
data(dat286)
datRat<-subset(dat286.long,(Series=="Equi" | Series =="RatioA") & Amount=="100 fmoles")
datRat$SampleID<-factor(datRat$SampleID)
datRat$Series<-factor(datRat$Series)
res <- isoLQNO(datRat, long=TRUE)
```

IsomirDataSeq-class *Class that contains all isomiRs annotation for all samples*

Description

The IsomirDataSeq is a subclass of SummarizedExperiment used to store the raw data, intermediate calculations and results of an miRNA/isomiR analysis. This class stores all raw isomiRs data for each sample, processed information, summary for each isomiR type, raw counts, normalized counts, and table with experimental information for each sample.

Details

IsomirDataSeqFromFiles creates this object using seqbuster output files.

Methods for this objects are counts to get count matrix and isoSelect for miRNA/isomiR selection. Functions available for this object are isoCounts for count matrix creation, isoNorm for normalization, isoDE for differential expression and isoPLSDA for clustering. isoPlot helps with basic expression plot.

metadata contains two lists: rawList is a list with same length than number of samples and stores the input files for each sample; isoList is a list with same length than number of samples and stores

information for each isomiR type summarizing the different changes for the different isomiRs (trimming at 3', trimming a 5', addition and substitution). For instance, you can get the data stored in `isoList` for sample 1 and 5' changes with this code `metadata(ids)[['isoList']][[1]]$t5sum`.

The naming of isomiRs follows these rules:

- miRNA name
- type:ref if the sequence is the same than the miRNA reference. `iso` if the sequence has variations.
- `t5 tag`:indicates variations at 5 position. The naming contains two words: `direction - nucleotides`, where direction can be UPPER CASE NT (changes upstream of the 5 reference position) or LOWER CASE NT (changes downstream of the 5 reference position). `0` indicates no variation, meaning the 5 position is the same than the reference. After `direction`, it follows the nucleotide/s that are added (for upstream changes) or deleted (for downstream changes).
- `t3 tag`:indicates variations at 3 position. The naming contains two words: `direction - nucleotides`, where direction can be LOWER CASE NT (upstream of the 3 reference position) or UPPER CASE NT (downstream of the 3 reference position). `0` indicates no variation, meaning the 3 position is the same than the reference. After `direction`, it follows the nucleotide/s that are added (for downstream changes) or deleted (for upstream chanes).
- `ad tag`:indicates nucleotides additions at 3 position. The naming contains two words: `direction - nucleotides`, where direction is UPPER CASE NT (upstream of the 5 reference position). `0` indicates no variation, meaning the 3 position has no additions. After `direction`, it follows the nucleotide/s that are added.
- `mm tag`: indicates nucleotides substitutions along the sequences. The naming contains three words: `position-nucleotideATsequence-nucleotideATreference`.
- `seed tag`: same than `mm` tag, but only if the change happens between nucleotide 2 and 8.

In general nucleotides in UPPER case mean insertions respect to the reference sequence, and nucleotides in LOWER case mean deletions respect to the reference sequence.

Examples

```
path <- system.file("extra", package="isomiRs")
fn_list <- list.files(path, full.names = TRUE)
de <- data.frame(row.names=c("f1" , "f2"),
                 condition = c("newborn", "newborn"))
ids <- IsomirDataSeqFromFiles(fn_list, coldata=de)

head(counts(ids))
```

IsomirDataSeqFromFiles

`IsomirDataSeqFromFiles` *loads miRNA annotation from seqbuster tool*

Description

This function parses output of seqbuster tool to allow isomiRs/miRNAs analysis of samples in different groups such as characterization, differential expression and clustering. It creates an `IsomirDataSeq` object.

Usage

```
IsomirDataSeqFromFiles(files, coldata, rate = 0.2, canonicalAdd = TRUE,
  uniqueMism = TRUE, design = ~1L, header = TRUE, skip = 0,
  quiet = TRUE, ...)
```

Arguments

files	files with the output of seqbuster tool
coldata	data frame containing groups for each sample
rate	minimum counts fraction to consider a mismatch a real mutation
canonicalAdd	boolean only keep A/T non-template addition. All non-template nucleotides at the 3' end will be removed if they contain C/G nts.
uniqueMism	boolean only keep mutations that have a unique hit to one miRNA molecule
design	a formula to pass to DESeqDataSet
header	boolean to indicate files contain headers
skip	skip first line when reading files
quiet	boolean indicating to print messages while reading files. Default FALSE.
...	arguments provided to SummarizedExperiment including rowData.

Details

This function parses the output of http://seqcluster.readthedocs.org/mirna_annotation.html for each sample to create a count matrix for isomiRs, miRNAs or isomiRs grouped in types (i.e all sequences with variations at 5' but ignoring any other type). It creates IsomirDataSeq object (see link to example usage of this class) to allow visualization, queries, differential expression analysis and clustering. To create the IsomirDataSeq, it parses the isomiRs files, and generates an initial matrix having all isomiRs detected among samples. As well, it creates a summary for each isomiR type (trimming, addition and substitution) to visualize general isomiRs distribution.

Value

IsomirDataSeq class object.

Examples

```
path <- system.file("extra", package="isomiRs")
fn_list <- list.files(path, full.names = TRUE)
de <- data.frame(row.names=c("f1" , "f2"),
                 condition = c("newborn", "newborn"))
ids <- IsomirDataSeqFromFiles(fn_list, coldata=de)

head(counts(ids))
```

isoNetwork *Clustering miRNAs-genes pairs in similar pattern expression*

Description

Clustering miRNAs-genes pairs

Usage

```
isoNetwork(mirna_rse, gene_rse, target, org, summarize = "group",
  genename = "ENSEMBL", min_cor = -0.6)
```

Arguments

mirna_rse	SummarizedExperiment::SummarizedExperiment with miRNA information. See details.
gene_rse	SummarizedExperiment::SummarizedExperiment with gene information. See details.
target	Matrix with miRNAs (columns) and genes (rows) target prediction (1 if it is a target, 0 if not).
org	AnnotationDbi::AnnotationDb obejct. For example:(org.Mm.eg.db).
summarize	Character column name in `colData(rse)` to use to group samples and compare betweem miRNA/gene expression.
genename	Character keytype of the gene names in gene_rse object.
min_cor	Numeric cutoff to consider a miRNA to regulate a target.

Details

This function will correlate miRNA and gene expression data using a specific metadata variable to group samples and detect pattern of expression that will be annotated with GO terms. mirna_rse and gene_rse can be created using the following code:

```
mi_rse = SummarizedExperiment(assays=SimpleList(norm=mirna_matrix), colData, metadata=list(sign=m
```

where, `mirna_matrix` is the normalized counts expression, `colData` is the metadata information and `mirna_keep` the list of miRNAs to be used by this function.

Value

list with network information

Examples

```
library(org.Mm.eg.db)
library(clusterProfiler)
data(isoExample)
# ego <- enrichGO(row.names(assay(gene_ex_rse, "norm")),
#                 org.Mm.eg.db, "ENSEMBL", ont = "BP")
# data = isoNetwork(mirna_ex_rse, gene_ex_rse, ma_ex,
#                   org = slot(ego, "result"))
# isoPlotNet(data)
```

isoNorm *Normalize count matrix*

Description

This function normalizes raw count matrix using `DESeq2::rlog()` function from DESeq2::DESeq2-package.

Usage

```
isoNorm(ids, formula = NULL)
```

Arguments

`ids`	Object of class IsomirDataSeq.
`formula`	Formula that will be used for normalization.

Value

IsomirDataSeq object with the normalized count matrix in a slot. The normalized matrix can be access with `counts(ids, norm=TRUE)`.

Examples

```
data(mirData)
ids <- isoCounts(mirData, minc=10, mins=6)
ids <- isoNorm(mirData, formula=~group)
head(counts(ids, norm=TRUE))
```

isoPlot *Plot the amount of isomiRs in different samples*

Description

This function plot different isomiRs proportion for each sample. It can show trimming events at both side, additions and nucleotides changes.

Usage

```
isoPlot(ids, type = "iso5", column = "condition")
```

Arguments

`ids`	Object of class IsomirDataSeq.
`type`	String (iso5, iso3, add, subs, all) to indicate what isomiRs to use for the plot. See details for explanation.
`column`	String indicating the column in `colData` to color samples.

Details

There are four different values for type parameter. To plot trimming at 5' or 3' end, use type="iso5" or type="iso3". Get a summary of all using type="all". In this case, it will plot 3 positions at both side of the reference position described at miRBase site. Each position refers to the number of sequences that start/end before or after the miRBase reference. The color indicates the sample group. The size of the point is proportional to the number of total counts. The position at y is the number of different sequences.

Same logic applies to type="add" and type="subs". However, when type="add", the plot will refer to addition events from the 3' end of the reference position. Note that this additions don't match to the precursor sequence, they are non-template additions. In this case, only 3 positions after the 3' end will appear in the plot. When type="subs", it will appear one position for each nucleotide in the reference miRNA. Points will indicate isomiRs with nucleotide changes at the given position. When type="all" a colar coordinate map will show the abundance of each isomiR type in a single plot.

Value

ggplot2::ggplot() Object showing different isomiRs changes at different positions.

Examples

```
data(mirData)
isoPlot(mirData, column="group")
```

---

isoPlotNet *Functional miRNA / gene expression profile plot*

Description

Plot analysis from isoNetwork

Usage

```
isoPlotNet(obj)
```

Arguments

obj	Output from isoNetwork().

Value

Network ggplot.

isoPlotPosition *Plot nucleotides changes at a given position*

Description

This function plot different isomiRs proportion for each sample at a given position focused on the nucleotide change that happens there.

Usage

```
isoPlotPosition(ids, position = 1, column = "condition")
```

Arguments

ids	Object of class IsomirDataSeq.
position	Integer indicating the position to show.
column	String indicating the column in colData to color samples.

Details

It shows the nucleotides changes at the given position for each sample in each group. The color indicates the sample group. The size of the point is proportional to the number of total counts of isomiRs with changes. The position at y is the number of different sequences supporting the change.

Value

`ggplot2::ggplot()` Object showing nucleotide changes at a given position.

Examples

```
data(mirData)
isoPlotPosition(mirData, column="group")
```

isoPLSDA *Partial Least Squares Discriminant Analysis for* IsomirDataSeq

Description

Use PLS-DA method with the normalized count data to detect the most important features (miRNAs/isomiRs) that explain better the group of samples given by the experimental design. It is a supervised clustering method with permutations to calculate the significance of the analysis.

Usage

```
isoPLSDA(ids, group, validation = NULL, learn = NULL, test = NULL,
 tol = 0.001, nperm = 400, refinment = FALSE, vip = 1.2)
```

Arguments

ids	Object of class IsomirDataSeq
group	Column name in colData(ids) to use as variable to explain.
validation	Type of validation, either NULL or "learntest". Default NULL.
learn	Optional vector of indexes for a learn-set. Only used when validation="learntest". Default NULL.
test	Optional vector of indices for a test-set. Only used when validation="learntest". Default NULL
tol	Tolerance value based on maximum change of cumulative R-squared coefficient for each additional PLS component. Default tol=0.001.
nperm	Number of permutations to compute the PLD-DA p-value based on R2 magnitude. Default nperm=400.
refinment	Logical indicating whether a refined model, based on filtering out variables with low VIP values.
vip	Variance Importance in Projection threshold value when a refinement process is considered. Default vip=1.2 .

Details

Partial Least Squares Discriminant Analysis (PLS-DA) is a technique specifically appropriate for analysis of high dimensionality data sets and multicollinearity (*Perez-Enciso, 2013*). PLS-DA is a supervised method (i.e. makes use of class labels) with the aim to provide a dimension reduction strategy in a situation where we want to relate a binary response variable (in our case young or old status) to a set of predictor variables. Dimensionality reduction procedure is based on orthogonal transformations of the original variables (miRNAs/isomiRs) into a set of linearly uncorrelated latent variables (usually termed as components) such that maximizes the separation between the different classes in the first few components (*Xia, 2011*). We used sum of squares captured by the model (R2) as a goodness of fit measure.

We implemented this method using the DiscriMiner::DiscriMiner-package into isoPLSDA() function. The output p-value of this function will tell about the statistical significant of the group separation using miRNA/isomiR expression data.

Read more about the parameters related to the PLS-DA directly from DiscriMiner::plsDA() function.

Value

A base::list with the following elements: R2Matrix (R-squared coefficients of the PLS model), components (of the PLS, similar to PCs in a PCA), vip (most important isomiRs/miRNAs), group (classification of the samples), p.value and R2PermutationVecto obtained by the permutations.

If the option refinment is set to TRUE, then the following elements will appear: R2RefinedMatrix and componentsRefinedModel (R-squared coefficients of the PLS model only using the most important miRNAs/isomiRs). As well, p.valRefined and R2RefinedPermutationVector with p-value and R2 of the permutations where samples were randomized. And finally, p.valRefinedFixed and R2RefinedFixedPermutationVector with p-value and R2 of the permutations where miRNAs/isomiRs were randomized.

References

Perez-Enciso, Miguel and Tenenhaus, Michel. Prediction of clinical outcome with microarray data: a partial least squares discriminant analysis (PLS-DA) approach. Human Genetics. 2003.

Xia, Jianguo and Wishart, David S. Web-based inference of biological patterns, functions and pathways from metabolomic data using MetaboAnalyst. Nature Protocols. 2011.

Examples

```
data(mirData)
# Only miRNAs with > 10 reads in all samples.
ids <- isoCounts(mirData, minc=10, mins=6)
ids <- isoNorm(ids, formula=~group)
pls.ids = isoPLSDA(ids, "group", nperm = 2)
cat(paste0("pval:",pls.ids$p.val))
cat(paste0("components:",pls.ids$components))
```

`isoPLSDAplot` *Plot components from isoPLSDA analysis (pairs plot)*

Description

Plot the most significant components that come from `isoPLSDA()` analysis together with the density of the samples scores along those components.

Usage

```
isoPLSDAplot(pls, n = 2)
```

Arguments

`pls`	Output from `isoPLSDA()` function.
`n`	Number of components to plot.

Details

The function `isoPLSDAplot` helps to visualize the results from `isoPLSDA()`. It will plot the samples using the significant components (t1, t2, t3 ...) from the PLS-DA analysis and the samples score distribution along the components. It uses `GGally::ggpairs()` for the plot.

Value

`GGally::ggpairs()` plot showing the scores for each sample using isomiRs/miRNAs expression to explain variation.

base::data.frame object with a first column refering to the sample group, and the following columns refering to the score that each sample has for each component from the PLS-DA analysis.

Examples

```
data(mirData)
# Only miRNAs with > 10 reads in all samples.
ids <- isoCounts(mirData, minc=10, mins=6)
ids <- isoNorm(ids, formula=~group)
pls.ids <- isoPLSDA(ids, "group", nperm = 2)
isoPLSDAplot(pls.ids)
```

`isoSelect` *Method to select specific miRNAs from an IsomirDataSeq object.*

Description

This method allows to select a miRNA and all its isomiRs from the count matrix.

Usage

```
isoSelect.IsomirDataSeq(object, mirna, minc = 10)

## S4 method for signature 'IsomirDataSeq'
isoSelect(object, mirna, minc = 10)
```

Arguments

`object`	A IsomirDataSeq object.
`mirna`	String referring to the miRNA to show.
`minc`	Minimum number of isomiR reads needed to be included in the table.

Value

S4Vectors::DataFrame with count information. The `row.names` show the isomiR names, and each of the columns shows the counts for this isomiR in that sample. Mainly, it will return the count matrix only for isomiRs belonging to the miRNA family given by the `mirna` parameter. IsomiRs need to have counts bigger than `minc` parameter at least in one sample to be included in the output. Annotation of isomiRs follows these rules:

- miRNA name
- mismatches
- additions
- 5 trimming events
- 3 trimming events

Author(s)

Lorena Pantano

Examples

```
data(mirData)
# To select isomiRs from let-7a-5p miRNA
# and with 10000 reads or more.
isoSelect(mirData, mirna="hsa-let-7a-5p", minc=10000)
```

isoTop — *Heatmap of the top expressed isomiRs*

Description

This function creates a heatmap with the top N isomiRs/miRNAs. It uses the matrix under `counts(ids)` to get the top expressed isomiRs/miRNAs using the average expression value and plot a heatmap with the raw counts for each sample.

Usage

```
isoTop(ids, top = 20)
```

Arguments

`ids`	Object of class IsomirDataSeq.
`top`	Number of isomiRs/miRNAs used.

Value

heatmap with top expressed miRNAs

Examples

```
data(mirData)
isoTop(mirData)
```

LQNO — *LQNO distribution*

Description

Linear quadratic family that assumes the following relation for the *variance* of the normal distribution `Var = mu*(1+s*mu)`. regression on mu and on the sigma (log and identity links)

Usage

```
dLQNO(x, mu = 1, sigma = 1, log = FALSE)
pLQNO(q, mu = 1, sigma = 1, lower.tail = TRUE, log.p = FALSE)
qLQNO(p, mu = 1, sigma = 1, lower.tail = TRUE, log.p = FALSE)
rLQNO(n, mu = 1, sigma = 1)
LQNO(mu.link="log", sigma.link="log")

dLQNO(x, mu = 1, sigma = 1, log = FALSE)

pLQNO(q, mu = 1, sigma = 1, lower.tail = TRUE, log.p = FALSE)

qLQNO(p, mu = 1, sigma = 1, lower.tail = TRUE, log.p = FALSE)

rLQNO(n, mu = 1, sigma = 1)
```

Arguments

mu.link	Type of transformation
sigma.link	Type of transformation
x	Vector of quantiles.
mu	Vector of means.
sigma	Vector of standard deviations.
log	Logical; if TRUE, probabilities p are given as log(p).
q	Vector of quantiles.
lower.tail	Logical; if TRUE (default), probabilities are P(X < x) otherwise, P(X > x).
log.p	Logical; if TRUE, probabilities p are given as log(p).
p	Vector of probabilities.
n	Number of observations. If length(n) > 1, the length is taken to be the number required.

Details

Methods adapted from:

Argyropoulos, Christos, et al. "Modeling bias and variation in the stochastic processes of small RNA sequencing." Nucleic Acids Research (2017).

Value

LQNO function

Author(s)

Christos Argyropoulos

---

ma_ex *Data frame containing gene-mirna relationship*

Description

Data frame containing gene-mirna relationship

Usage

```
ma_ex
```

Format

A data frame with rows sames as gene_ex_rse and columns same as mirna_ex_rse.

mirData *Example of IsomirDataSeq with human brain miRNA counts data*

Description

This data set is the object return by `IsomirDataSeqFromFiles`. It contains miRNA count data from 14 samples: 7 control individuals (pc) and 7 patients with Parkinson's disease in early stage (Pantano et al, 2016). Use `colData` to see the experiment design.

Usage

```
data("mirData")
```

Format

a `IsomirDataSeq` class.

Author(s)

Lorena Pantano, 2016-04-07

Source

Data is available from GEO dataset under accession number GSE97285

Every sample was analyzed with seqbuster tool, see `http://seqcluster.readthedocs.org/mirna_annotation.html` for more details. You can get same files running the small RNA-seq pipeline from `https://github.com/chapmanb/bcbio-nextgen`.

bcbio_nextgen was used for the full analysis.

```
library(isomiRs) files = list.files(file.path(root_path),pattern = "mirbase-ready",recursive = T,f
metadata_fn =  list.files(file.path(root_path),pattern = "summary.csv$",recursive = T, full.names
metadata = read.csv(metadata_fn, row.names="sample_id") condition = names(metadata)[1]
mirData <- IsomirDataSeqFromFiles(files[rownames(design)], metadata)
```

References

Pantano L, Friedlander MR, Escaramis G, Lizano E et al. Specific small-RNA signatures in the amygdala at premotor and motor stages of Parkinson's disease revealed by deep sequencing analysis. Bioinformatics 2016 Mar 1;32(5):673-81. PMID: 26530722

mirna_ex_rse *Data frame containing mirna expression data*

Description

Data frame containing mirna expression data

Usage

```
mirna_ex_rse
```

Format

mirna expression data with 18 samples: example of a time series data

---

`mirTritation` *Data frame containing mirna from Argyropoulos's paper*

Description

Argyropoulos, Christos, et al. "Modeling bias and variation in the stochastic processes of small RNA sequencing." Nucleic Acids Research (2017).

Usage

```
mirTritation
```

Format

mirna expression data in long format. Train and test data to use with isoCorrect

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