# Package 'soGGi'

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Type Package
Title Visualise ChIP-seq, MNase-seq and motif occurrence as aggregate plots Summarised Over Grouped Genomic Intervals
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Description The soGGi package provides a toolset to create genomic interval aggregate/summary plots of signal or motif occurence from BAM and bigWig files as well as PWM, rlelist, GRanges and GAlignments Bioconductor objects. soGGi allows for normalisation, transformation and arithmetic operation on and between summary plot objects as well as grouping and subsetting of plots by GRanges objects and user supplied metadata. Plots are created using the GGplot2 libary to allow user defined manipulation of the returned plot object. Coupled together, soGGi features a broad set of methods to visualise genomics data in the context of groups of genomic intervals such as genes, superenhancers and transcription factor binding events.  biocViews Sequencing, ChIPSeq, Coverage
License GPL (>= 3)
LazyLoad yes
<b>Depends</b> R (>= 3.2.0), BiocGenerics, SummarizedExperiment
Imports methods, reshape2, ggplot2, S4Vectors, IRanges, GenomeInfoDb, GenomicRanges, Biostrings, Rsamtools, GenomicAlignments, rtracklayer, preprocessCore, chipseq, BiocParallel
Collate 'allClasses.r' 'motifTools.R' 'peakTransforms.r' 'plots.R' 'soggi.R'
VignetteBuilder knitr
Suggests testthat, BiocStyle, knitr
NeedsCompilation no
R topics documented:
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c,ChIPprofile-method

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c, ChIPprofile-method Join, subset and manipulate ChIPprofile objects

#### **Description**

Join, subset and manipulate ChIPprofile objects

#### Usage

```
## S4 method for signature 'ChIPprofile'
c(x, ..., recursive = FALSE)

## S4 method for signature 'ChIPprofile'
rbind(x, ..., deparse.level = 1)

## S4 method for signature 'ChIPprofile'
cbind(x, ..., deparse.level = 1)

## S4 method for signature 'ChIPprofile,ANY,missing'
x[[i, j, ...]]

## S4 method for signature 'ChIPprofile'
x$name
```

#### **Arguments**

```
j Should be missing
... objects to be concatenated.
recursive logical. If recursive = TRUE, the function recursively descends through lists (and pairlists) combining all their elements into a vector.
deparse.level See ?base::cbind for a description of this argument.
x object from which to extract element(s) or in which to replace element(s).
```

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i

indices specifying elements to extract or replace. Indices are numeric or character vectors or empty (missing) or NULL. Numeric values are coerced to integer as by as.integer (and hence truncated towards zero). Character vectors will be matched to the names of the object (or for matrices/arrays, the dimnames): see 'Character indices' below for further details.

For [-indexing only: i, j, ... can be logical vectors, indicating elements/slices to select. Such vectors are recycled if necessary to match the corresponding extent. i, j, ... can also be negative integers, indicating elements/slices to leave out of the selection.

When indexing arrays by [ a single argument i can be a matrix with as many columns as there are dimensions of x; the result is then a vector with elements corresponding to the sets of indices in each row of i.

An index value of NULL is treated as if it were integer(0).

name

A literal character string or a name (possibly backtick quoted). For extraction, this is normally (see under 'Environments') partially matched to the names of the object.

#### Value

A ChIPprofile object

#### **Examples**

```
data(chipExampleBig)
x <- c(chipExampleBig[[1]],chipExampleBig[[2]])
y <- rbind(chipExampleBig[[1]],chipExampleBig[[2]])</pre>
```

chipExampleBig

Example ChIPprofiles

# Description

This dataset contains peaks from ChIP-signal over genes

#### Usage

```
data(chipExampleBig)
```

#### **Details**

• ChIPprofiles

#### Value

A ChIPprofile object

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ChIPprofile-class The sogs

The soggi function and ChIPprofile object.

#### **Description**

Manual for soggi and ChIPprofile object

The soggi function is the constructor for ChIPprofile objects.

#### Usage

```
regionPlot(bamFile, testRanges, samplename = NULL, nOfWindows = 100,
   FragmentLength = 150, style = "point", distanceAround = NULL,
   distanceUp = NULL, distanceDown = NULL, distanceInRegionStart = NULL,
   distanceOutRegionStart = NULL, distanceInRegionEnd = NULL,
   distanceOutRegionEnd = NULL, paired = FALSE, normalize = "RPM",
   plotBy = "coverage", removeDup = FALSE, verbose = TRUE,
   format = "bam", seqlengths = NULL, forceFragment = NULL,
   method = "bin", genome = NULL, cutoff = 80, downSample = NULL,
   minFragmentLength = NULL, maxFragmentLength = NULL)
```

#### **Arguments**

bamFile Character vector for location of BAM file or bigWig, an rleList or PWM matrix.

testRanges GRanges object or character vector of BED file location of regions to plot.

samplename Character vector of sample name. Default is NULL.

nOfWindows Number of windows to bin regions into for coverage calculations (Default 100)

FragmentLength Integer vector Predicted or expected fragment length.

style "Point" for per base pair plot, "percentOfRegion" for normalised length and "re-

gion" for combined plot

distanceAround Distance around centre of region to be used for plotting

distanceUp Distance upstream from centre of region to be used for plotting
distanceDown Distance downstream from centre of region to be used for plotting

distanceInRegionStart

Distance into region start (5' for Watson/positive strand or notspecified strand Regions,3' for Crick/negatie strand regions) for plotting.

 ${\tt distance Out Region Start}$ 

Distance out from region start (5' for Watson/positive strand or notspecified strand Regions,3' for Crick/negatie strand regions) for plotting.

distanceInRegionEnd

Distance into region end (3' for Watson/positive strand or notspecified strand Regions,5' for Crick/negatie strand regions) for plotting.

 ${\tt distance Out Region End}$ 

Distance out from region end (3' for Watson/positive strand or notspecified strand Regions,5' for Crick/negatie strand regions) for plotting.

paired Is data paired end

normalize Calculate coverage as RPM. Presently only RPM available.

findconsensusRegions 5

plotBy Score to be used for plotting. Presently only coverage.

removeDup Remove duplicates before calculating coverage.

verbose TRUE or FALSE

format character vector of "BAM", "BigWig", "RleList" or "PWM"

seqlengths Chromosomes to be used. If missing will report all.

forceFragment Centre fragment and force consistent fragment width.

method Character vector of value "bp", "bin" or "spline". The bin method divides a re-

gion of interest into equal sized bins of number specified in nOfWindows. Coverage or counts are then summarised within these windows. The spline method creates a spline with the number of spline points as specified in nOfWindows

argument.

downSample Down sample BAM reads to this proportion of orginal.

genome BSGenome object to be used when using PWM input.

cutoff Cut-off for idnetifying motifs when using PWM input.

minFragmentLength

Remove fragments smaller than this.

maxFragmentLength

Remove fragments larger than this.

#### Value

ChIPprofile A ChIPprofile object.

#### References

See http://bioinformatics.csc.mrc.ac.uk for more details on soGGi workflows

#### **Examples**

```
data(chipExampleBig)
chipExampleBig
```

findconsensusRegions Plot coverage of points or regions.

#### **Description**

Plot coverage of points or regions.

Returns summits and summmit scores after optional fragment length prediction and read extension

#### Usage

```
findconsensusRegions(testRanges, bamFiles = NULL, method = "majority",
   summit = "mean", resizepeak = "asw", overlap = "any",
   fragmentLength = NULL, NonPrimaryPeaks = list(withinsample = "drop",
   betweensample = "mean"))
summitPipeline(reads, peakfile, fragmentLength, readlength)
```

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#### **Arguments**

testRanges Named character vector of region locations
bamFiles Named character vector of bamFile locations
method Method to select reproducible summits to merge.

summit Only mean available resizepeak Only asw available

overlap Type of overlap to consider for finding consensus sites fragmentLength Predicted fragment length. Set to NULL to auto-calculate

NonPrimaryPeaks

A list of parameters to deal with non primary peaks in consensus regions.

peakfile GRanges of genomic intervals to summit.

reads Character vector of bamFile location or GAlignments object

readlength Read length of alignments.

#### Value

Consensus A GRanges object of consensus regions with consensus summits.

Summits A GRanges object of summits and summit scores.

groupByOverlaps Create GRangeslist from all combinations of GRanges

# Description

Create GRangeslist from all combinations of GRanges

# Usage

```
groupByOverlaps(testRanges)
```

### **Arguments**

testRanges A named list of GRanges or a named GRangesList

#### Value

groupedGRanges A named GRangesList object.

# **Examples**

```
data(ik_Example)
  gts <- groupByOverlaps(ik_Example)</pre>
```

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ik\_Example

Example Ikaros peaksets

# Description

This dataset contains peaks from Ikaros ChIP by two antibodies

# Usage

```
data(ik_Example)
```

# **Details**

• Ikpeaksets

# Value

A list containing two GRanges objects

ik\_Profiles

Example Ikaros signal over peaksets

# Description

This dataset contains signal over peaks from Ikaros ChIP by two antibodies

# Usage

```
data(ik_Profiles)
```

# **Details**

• ik\_Profiles

# Value

A ChIPprofile object

8 normaliseQuantiles

normalise

Normalise ChIPprofiles

# Description

Various normalisation methods for ChIPprofile objects

#### Usage

```
## S4 method for signature 'ChIPprofile'
normalise(object)

## S4 method for signature 'ChIPprofile, character, numeric'
normalise(object = "ChIPprofile",
    method = "rpm", normFactors = NULL)
```

#### **Arguments**

object A ChIPprofile object

method A character vector specifying normalisation method. Currently "rpm" for nor-

malising signal for BAM to total reads, "quantile" to quantile normalise across samples, "signalInRegion" to normalise to proportion of signal within intervals, "normaliseSample" to normalise across samples and "normaliseRegions" to ap-

ply a normalisation across intervals.

normFactors A numeric vector used to scale columns or rows.

#### Value

A ChIPprofile object

# Author(s)

Thomas Carroll

#### **Examples**

```
data(chipExampleBig)
normalise(chipExampleBig,method="quantile",normFactors=1)
```

normaliseQuantiles

Normalise quantile

# Description

Quantile normalisation across bins/regions.

#### Usage

```
## S4 method for signature 'ChIPprofile'
normaliseQuantiles(object)

## S4 method for signature 'ChIPprofile'
normaliseQuantiles(object = "ChIPprofile")
```

#### **Arguments**

object

A ChIPprofile object

#### Value

A ChIPprofile object containing normalised data

#### Author(s)

Thomas Carroll

#### **Examples**

```
data(chipExampleBig)
normaliseQuantiles(chipExampleBig)
```

```
{\it Ops, ChiPprofile, ChiPprofile-method} \\ {\it Arithmetic operations}
```

# **Description**

Arithmetic operations

#### Usage

```
## S4 method for signature 'ChIPprofile, ChIPprofile'
Ops(e1, e2)

## S4 method for signature 'ChIPprofile, numeric'
Ops(e1, e2)

## S4 method for signature 'numeric, ChIPprofile'
Ops(e1, e2)

## S4 method for signature 'ChIPprofile'
mean(x, ...)

## S4 method for signature 'ChIPprofile'
log2(x)

## S4 method for signature 'ChIPprofile'
log(x, base = exp(1))
```

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#### **Arguments**

e1	ChIPprofile object
e2	ChIPprofile object
Х	objects.
	further arguments passed to methods.
base	a positive or complex number: the base with respect to which logarithms are computed. Defaults to $e=\exp(1)$ .

#### Value

A ChIPprofile object of result of arithmetic operation.

# **Examples**

```
data(chipExampleBig)
chipExampleBig[[1]] + chipExampleBig[[2]]
```

orientBy

Set strand by overlapping or nearest anchor GRanges

# Description

Set strand by overlapping or nearest anchor GRanges

#### Usage

```
orientBy(testRanges, anchorRanges)
```

# Arguments

testRanges The GRanges object to anchor.

anchorRanges A GRanges object by which to anchor strand orientation.

#### Value

newRanges A GRanges object.

# **Examples**

```
data(ik_Example)
strand(ik_Example[[1]]) <- "+"
anchoredGRanges <- orientBy(ik_Example[[2]],ik_Example[[1]])</pre>
```

plotRegion 11

# **Description**

A function to plot regions

# Usage

```
## S4 method for signature 'ChIPprofile'
plotRegion(object,
gts,sampleData,groupData,summariseBy,
colourBy,lineBy,groupBy,
plotregion,outliers,freeScale)

## S4 method for signature 'ChIPprofile'
plotRegion(object = "ChIPprofile", gts = NULL,
    sampleData = NULL, groupData = NULL, summariseBy = NULL,
    colourBy = NULL, lineBy = NULL, groupBy = NULL, plotregion = "full",
    outliers = NULL, freeScale = FALSE)
```

# **Arguments**

_	
object	A ChIPprofile object
gts	A list of character vectors or GRangesList
plotregion	region to plot. For combined plots with style "region", may be "start" or "end" to show full resolution of plot of edges.
groupData	Dataframe of metadata for groups
sampleData	Dataframe of metadata for sample
summariseBy	Column names from GRanges elementmetadata. Formula or character vector of column names to use to collapse genomic ranges to summarised profiles. summariseBy can not be used injustion with groups specified by gts argument.
colourBy	Character vector or formula of either column names from colData(object) containing sample metadata or character vector "group" to colour by groups in gts
lineBy	Character vector or formula of either column names from colData(object) containing sample metadata or character vector "group" to set line type by groups in gts
groupBy	Character vector or formula of either column names from colData(object) containing sample metadata or character "group" to colour by groups in gts
outliers	A numeric vector of length 1 containing proportion from limits to windsorise.]
freeScale	TRUE or FALSE to set whether ggplot 2 facets have their own scales. Useful for comparing multiple samples of differing depths without normalisation. Default is FALSE.

# Value

A gg object from ggplot2

pwmToCoverage

#### Author(s)

Thomas Carroll

# **Examples**

```
data(chipExampleBig)
plotRegion(chipExampleBig[[2]])
```

pwmCov

Example motif coverage

# Description

This dataset contains an rlelist of motif coverage

# Usage

```
data(pwmCov)
```

#### **Details**

• pwmCov

#### Value

A rlelist of motif coverage

pwmToCoverage

PWM hits and motif scores as an RLElist

# Description

Creates rlelist of pwm hits.

Motif score as an RLElist

# Usage

```
pwmToCoverage(pwm, genome, min = "70%", removeRand = FALSE,
    chrsOfInterest = NULL)

makeMotifScoreRle(pwm, regions, genome, extend, removeRand = FALSE,
    strandScore = "mean", atCentre = FALSE)
```

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#### **Arguments**

pwm A PWM matrix object. genome A BSgenome object

min pwm score (as percentage of maximum score) cutoff

removeRand Remove contigs with rand string

chrsOfInterest Chromosomes to use

regions GRanges object to include in pwm rlelist

extend bps to extend regions by

strandScore Method for averaging strand. Options are max, mean, sum, bothstrands

atCentre TRUE/FALSE. TRUE assigns score onto 1bp position at centre of motif. FALSE

assigns every basepair the sum of scores of all overlapping motifs.

#### Value

A RLElist of motif density per base pair to be used as input to main soggi function.

#### Author(s)

Thomas Carroll

#### **Examples**

```
data(pwmCov)
data(singleGRange)
```

singleGRange

A single GRange

# Description

This dataset contains an rlelist of motif coverage

#### Usage

data(singleGRange)

#### **Details**

• singleGRange

#### Value

A single GRanges used in motif coverage example/

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