Package 'rnaEditr'

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Title Statistical analysis of RNA editing sites and hyper-editing
regions

Version 1.17.0

Description RNAeditr analyzes site-specific RNA editing events, as well as
hyper-editing regions. The editing frequencies can be tested against binary,
continuous or survival outcomes. Multiple covariate variables as well as
interaction effects can also be incorporated in the statistical models.

Depends R (>= 4.0)

License GPL-3

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Author Lanyu Zhang [aut, cre],
Gabriel Odom [aut],
Tiago Silva [aut],
Lissette Gomez [aut],
Lily Wang [aut]

Maintainer Lanyu Zhang <jennyzly2016@gmail.com>

Contents

AddMetaData *Add metadata columns to GRanges object.*

Description

Add metadata information to GRanges object.

Usage

```
AddMetaData(
  target_gr,
  annot_gr = NULL,
  annotType_char = c("geneSymbol", "region"),
  annotLabel_char = "symbol",
```

```
    genome = c("hg38", "hg19")
  )
```

Arguments

`target_gr`	A GRanges object that will be annotated with metadata
`annot_gr`	A GRanges object that includes the metadata information. When `annotType_char` = "geneSymbol", this argument can be left as NULL, and the gene annotation file saved in the package will be used to annotate `target_gr`. When `annotType_char` = "region", this argument must be specified, each row in `target_gr` will be annotated with rows in `annot_gr` that overlap with it.
`annotType_char`	Type of the metadata column, defaults to "geneSymbol".
`annotLabel_char`	
	Name of the metadata column, defaults to "symbol" which corresponds to default setting "geneSymbol" for argument `annotType_char`.
`genome`	Use "hg19" or "hg38" gene reference. Defaults to "hg38".

Value

A GRanges object with `seqnames`, `ranges`, `region`, and supplied metadata information.

Examples

```
data(rnaedit_df)

input_gr <- TransformToGR(
  genes_char = "PHACTR4",
  type = "symbol",
  genome = "hg19"
)

# identifies co-edited region within input_gr
coedited_gr <- AllCoeditedRegions(
  regions_gr = input_gr,
  rnaEditMatrix = rnaedit_df,
  output = "GRanges",
  method = "spearman"
)

# identify input regions for co-edited regions
AddMetaData(
  target_gr = coedited_gr,
  annot_gr = input_gr,
  annotType_char = "region",
  annotLabel_char = "inputRegion",
  genome = "hg19"
)
```

AllCloseByRegions   *Extract clusters of RNA editing sites located closely in genomic re-*
                             gions.

Description

A wrapper function to extract clusters of RNA editing sites that are located closely in genomic
regions.

Usage

```
AllCloseByRegions(
  regions_gr,
  rnaEditMatrix,
  maxGap = 50,
  minSites = 3,
  progressBar = "time"
)
```

Arguments

regions_gr   A GRanges object of input genomic regions.

rnaEditMatrix   A matrix (or data frame) of RNA editing level values on individual sites, with
                  row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
                  as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).

maxGap   An integer, genomic locations within maxGap from each other are placed into
                  the same cluster. Defaults to 50.

minSites   An integer, minimum number of RNA editing sites within each resulting cluster.
                  Defaults to 3.

progressBar   Name of the progress bar to use. There are currently five types of progress
                  bars: "time", "none", "text", "tk", and "win". Defaults to "time". See
                  [create_progress_bar](#) for more details.

Details

The algorithm of this function is based on the [clusterMaker](#) function in the bumphunter R pack-
age. Each cluster is essentially a group of site locations such that two consecutive locations in the
cluster are separated by less than maxGap.

Value

A GRanges object containing genomic regions of RNA editing sites located closely within each
input pre-defined genomic region.

See Also

TransformToGR, AllCoeditedRegions, CreateEditingTable, SummarizeAllRegions, TestAssociations, AnnotateResults

Examples

```
data(rnaedit_df)

exm_regions <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

AllCloseByRegions(
  regions_gr = exm_regions,
  rnaEditMatrix = rnaedit_df,
  maxGap = 50,
  minSites = 3,
  progressBar = "time"
)
```

AllCoeditedRegions   *Extracts contiguous co-edited genomic regions from input genomic regions .*

Description

A wrapper function to extract contiguous co-edited genomic regions from input genomic regions.

Usage

```
AllCoeditedRegions(
  regions_gr,
  rnaEditMatrix,
  output = c("GRanges", "dataframe"),
  rDropThresh_num = 0.4,
  minPairCorr = 0.1,
  minSites = 3,
  method = c("spearman", "pearson"),
  returnAllSites = FALSE,
  progressBar = "time",
  verbose = TRUE
)
```

Arguments

regions_gr	A GRanges object of input genomic regions.
rnaEditMatrix	A matrix (or data frame) of RNA editing level values on individual sites, with row names as site IDs in the form of "chrAA:XXXXXXXX", and column names as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).
output	Type of output data. Defaults to "GRanges".
rDropThresh_num	
	Threshold for minimum correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites. Please set a number between 0 and 1. Defaults to 0.4.
minPairCorr	Threshold for minimum pairwise correlation of sites within a selected cluster. To use this filter, set a number between -1 and 1 (defaults to 0.1). To select all clusters (i.e. no filter), please set this argument to -1.
minSites	Minimum number of sites to be considered as a region. Only regions with more than minSites number of sites will be returned.
method	Method for computing correlation. Defaults to "spearman".
returnAllSites	When no contiguous co-edited regions are found in an input genomic region, returnAllSites = TRUE indicates returning all the sites in the input region, while returnAllSites = FALSE indicates not returning any site from input region. Defaults to FALSE.
progressBar	Name of the progress bar to use. There are currently five types of progress bars: "time", "none", "text", "tk", and "win". Defaults to "time". See [create_progress_bar](#) for more details.
verbose	Should messages and warnings be displayed? Defaults to FALSE, but is set to TRUE when called from within SingleCoeditedRegion().

Value

When output is set as "GRanges", a GRanges object with seqnames, ranges and strand of the contiguous co-edited regions will be returned. When output is set as "dataframe", a data frame with following columns will be returned:

- site : site ID.
- chr : chromosome number.
- pos : genomic position number.
- r_drop : the correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.
- keep : indicator for co-edited sites, the sites with keep = 1 belong to the contiguous and co-edited region.
- keep_contiguous : contiguous co-edited region number.
- regionMinPairwiseCor : the pairwise correlation of a subregion.
- keep_regionMinPairwiseCor : indicator for contiguous co-edited subregions, the regions with keepminPairwiseCor = 1 passed the minimum correlation and will be returned as a contiguous co-edited subregion.

See Also

[TransformToGR](), [AllCloseByRegions](), [CreateEditingTable](), [SummarizeAllRegions](), [TestAssociations](),
[AnnotateResults]()

Examples

```
data(rnaedit_df)

genes_gr <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

AllCoeditedRegions(
  regions_gr = genes_gr,
  rnaEditMatrix = rnaedit_df,
  output = "GRanges",
  method = "spearman"
)
```

AnnotateResults   *Add Annotations to site-specific or region-based analysis results.*

Description

Add annotations to site-specific or region-based analysis results from function [TestAssociations]().

Usage

```
AnnotateResults(
  results_df,
  closeByRegions_gr = NULL,
  inputRegions_gr = NULL,
  genome = c("hg38", "hg19"),
  analysis = c("region-based", "site-specific")
)
```

Arguments

results_df   An output data frame from function TestAssociations, which includes vari-
                  ables for locations and result of statistical tests for the genomic sites or regions.

closeByRegions_gr
                  An output GRanges object from function AllCloseByRegions, defaults to NULL.

inputRegions_gr
                  A GRanges object for input genomic regions, defaults to NULL.

genome   Use "hg19" or "hg38" gene reference. Defaults to "hg38".

analysis   Results type. Defaults to "region-based". When it's set to "site-specific",
                 arguments closeByRegions_gr and inputRegions_gr will not be used and set
                 to NULL automatically.

Value

A data frame with locations of the genomic sites or regions (seqnames, start, end, width), anno-
tations for locations (inputRegion, closeByRegion, symbol), test statistics (estimate, stdErr
or coef, exp_coef, se_coef), pValue and false discovery rate (fdr).

See Also

[TransformToGR](), [AllCloseByRegions](), [AllCoeditedRegions](), [CreateEditingTable](), [SummarizeAllRegions](),
[TestAssociations]()

Examples

```
data(rnaedit_df)

# get GRanges for genes
genes_gr <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

# find close-by regions within the genes
closebyRegions_gr <- AllCloseByRegions(
  regions_gr = genes_gr,
  rnaEditMatrix = rnaedit_df
)

# identify co-edited regions within the genes
coedited_gr <- AllCoeditedRegions(
  regions_gr = closebyRegions_gr,
  rnaEditMatrix = rnaedit_df,
  output = "GRanges",
  method = "spearman"
)

# summarize editing levels within each gene by maximum
summarizedRegions_df <- SummarizeAllRegions(
  regions_gr = coedited_gr,
  rnaEditMatrix = rnaedit_df,
  selectMethod = MaxSites
)

exm_pheno <- readRDS(
  system.file(
  "extdata",
  "pheno_df.RDS",
```

```
     package = 'rnaEditr',
     mustWork = TRUE
     )
 )

 # test summarized editing levels against survival outcome
 results_df <- TestAssociations(
   rnaEdit_df = summarizedRegions_df,
   pheno_df = exm_pheno,
   responses_char = "sample_type",
   covariates_char = NULL,
   respType = "binary"
 )

 AnnotateResults(
   results_df = results_df,
   closeByRegions_gr = closebyRegions_gr,
   inputRegions_gr = genes_gr,
   genome = "hg19"
 )
```

CountSamplesPerGroup   *Find minimum sample Size per group.*

Description

Find minimum sample size for each group which is decided by the combination of variables with class character or factor.

Usage

```
CountSamplesPerGroup(pheno_df, responses_char, covariates_char)
```

Arguments

pheno_df
: A data frame with phenotype and covariates, which should include all the samples in rnaEdit_df. Please make sure the input pheno_df has the variable named "sample" to indicate sample IDs.

responses_char
: A character vector of names of response variables in pheno_df. When respType is set as "survival", responses_char should have length 2. The first element must be the name of the variable with follow up time, and the second element must be the status indicator. Status indicator should be coded as 0/1(1=death), TRUE/FALSE(TRUE=death), or 1/2(2=death). Please make sure variable names are in this order. This code has not been tested for interval-censored data yet.

covariates_char
: A character vector of names of covariate variables in pheno_df.

Value

An integer.

Examples

```
exm_pheno <- readRDS(
  system.file(
  "extdata",
  "pheno_df.RDS",
  package = 'rnaEditr',
  mustWork = TRUE
  )
)

CountSamplesPerGroup(
  pheno_df = exm_pheno,
  responses_char = "sample_type",
  covariates_char = "race"
)
```

CreateEditingTable   *Convert RNA editing matrix into a special data frame with class*
                            rnaEdit_df.

Description

Convert RNA editing matrix to a special data frame with class rnaEdit_df, which is then used to
identify differentially co-edited regions with function [TestAssociations](#).

Usage

```
CreateEditingTable(rnaEditMatrix)
```

Arguments

rnaEditMatrix   A matrix of RNA editing level values on individual sites, with row names as site
                IDs in the form of "chrAA:XXXXXXXX", and column names as sample IDs.
                Please make sure to follow the format of example dataset (data(rnaedit_df)).

Value

A dataset of class rnaEdit_df, includes variables seqnames, start, end, width and summarized
RNA editing levels in each sample.

See Also

[TransformToGR](#), [AllCloseByRegions](#), [AllCoeditedRegions](#), [SummarizeAllRegions](#), [TestAssociations](#),
[AnnotateResults](#)

Examples

```
data(rnaedit_df)
CreateEditingTable(rnaEditMatrix = rnaedit_df)[1:3, 1:5]
```

CreateOutputDF   *Create output data in the format of data frame.*

Description

Output all the contiguous co-edited subregions found by [FindCorrelatedRegions](#) function and filtered by [GetMinPairwiseCor](#) function.

Usage

```
CreateOutputDF(
  keepSites_df,
  keepContiguousSites_df,
  keepminPairwiseCor_df,
  returnAllSites = FALSE,
  verbose = TRUE
)
```

Arguments

keepSites_df   An output data frame from function MarkCoeditedSites, with variables site, keep, ind, r_drop. Please see [MarkCoeditedSites](#) for details.

keepContiguousSites_df

An output data frame from function FindCorrelatedRegions with variables site, subregion. Please see [FindCorrelatedRegions](#) for details.

keepminPairwiseCor_df

An output data frame from function GetMinPairwiseCor with variables subregion, keepminPairwiseCor and minPairwiseCor. Please see [GetMinPairwiseCor](#) for details.

returnAllSites   When no contiguous co-edited regions are found in a input genomic region, returnAllSites = TRUE indicates outputting all the sites in this input region, while returnAllSites = FALSE indicates not returning any site in this input region. Defaults to FALSE.

verbose   Should messages and warnings be displayed? Defaults to TRUE.

Value

A data frame with following columns:

- site : site ID.
- chr : chromosome number.

- pos : genomic location.
- r_drop : the correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.
- keep : indicator for co-edited sites, The sites with keep = 1 belong to the contiguous and co-edited region.
- keep_contiguous : contiguous co-edited region number.
- regionMinPairwiseCor : the minimum pairwise correlation between sites within a subregion.
- keep_regionMinPairwiseCor : indicator for contiguous co-edited subregions, The regions with keepminPairwiseCor = 1 are the ones that passed regionMinPairwiseCor filter and will be returned as a contiguous co-edited sub-region.

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

exm_sites <- MarkCoeditedSites(
  rnaEditCluster_mat = exm_data,
  method = "spearman"
)

exm_regions <- FindCorrelatedRegions(
  sites_df = exm_sites,
  featureType = "site"
)

exm_probes <- split(
  x = exm_regions$site,
  f = exm_regions$subregion
)

exm_cor <- GetMinPairwiseCor(
  rnaEditCluster_mat = exm_data,
  minPairCorr = 0.1,
  probes_ls = exm_probes,
  method = "spearman"
)

CreateOutputDF(
  keepSites_df = exm_sites,
  keepContiguousSites_df = exm_regions,
  keepminPairwiseCor_df = exm_cor$keepminPairwiseCor_df
)
```

Description

Calculates the correlation coefficient between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites in a region.

Usage

```
CreateRdrop(
  data,
  method = c("spearman", "pearson"),
  minEditFreq = 0.05,
  verbose = TRUE
)
```

Arguments

data	A data frame of RNA editing level values on individual sites, with row names as sample IDs and column names as site IDs in the form of "chrAA:XXXXXXXX".
method	Method for computing correlation. Defaults to "spearman".
minEditFreq	Threshold for minimum percentage of edited samples for a given site. The r_drop value of the sites with frequency lower than minEditFreq will be set as NA. Please set a number between 0 and 1. Defaults to 0.05.
verbose	Should messages and warnings be displayed? Defaults to TRUE.

Value

A data frame with the following columns:

- site : site ID.
- r_drop : the correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

CreateRdrop(
  data = exm_data,
```

```
    method = "spearman"
  )
```

FindCorrelatedRegions   *Find contiguous co-edited subregions.*

Description

Find contiguous co-edited subregions based on the output file from function `MarkCoeditedSites`.

Usage

```
FindCorrelatedRegions(
  sites_df,
  featureType = c("site", "cpg"),
  minSites_int = 3
)
```

Arguments

sites_df	An output data frame from function `MarkCoeditedSites`, with variables `site`, `keep`, `ind`, `r_drop`. Please see `MarkCoeditedSites` for details.
featureType	Feature type, Defaults to `"site"`.
minSites_int	An integer indicates the minimum number of sites to be considered a contiguous co-edited region.

Value

A data frame with the following columns:

- `site` : site ID.
- `subregion` : index for each output contiguous co-edited region.

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

exm_sites <- MarkCoeditedSites(
  rnaEditCluster_mat = exm_data,
  method = "spearman"
)
```

```
FindCorrelatedRegions(
  sites_df = exm_sites,
  featureType = "site"
)
```

GetMinPairwiseCor   *Calculate minimum pairwise correlation for sub-regions.*

Description

Filter the contiguous co-edited subregions found from FindCorrelatedRegions, by calculating pairwise correlations and then selecting subregions passing the minimum correlation filter.

Usage

```
GetMinPairwiseCor(
  rnaEditCluster_mat,
  minPairCorr = 0.1,
  probes_ls,
  method = c("spearman", "pearson")
)
```

Arguments

rnaEditCluster_mat

A matrix of RNA editing level values on individual sites, with row names as sample IDs and column names as site IDs in the form of "chrAA:XXXXXXXX".

minPairCorr   Minimum pairwise correlation coefficient of sites within a cluster, used as a filter. To use this filter, set a number between -1 and 1 (defaults to 0.1). To turn it off, please set the number to -1.

probes_ls   A list of regions with sites. Please note that probes in each list need to be ordered by their locations.

method   Method for computing correlation. Defaults to "spearman".

Value

A list with a list of probes passing the minPairCorr and a data frame with the following columns:

- subregion : index for each output contiguous co-edited region.

- keepminPairwiseCor : indicator for contiguous co-edited subregions, The regions with keepminPairwiseCor = 1 passed the minimum correlation and will be returned as a contiguous co-edited subregion.

- minPairwiseCor : the minimum pairwise correlation of sites within a subregion.

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

exm_sites <- list(
  "1" = c("chr1:28661656", "chr1:28661718", "chr1:28662148")
)

GetMinPairwiseCor(
  rnaEditCluster_mat = exm_data,
  minPairCorr = 0.1,
  probes_ls = exm_sites,
  method = "spearman"
)
```

GetSitesLocations   *Extract RNA editing sites located in a genomic region.*

Description

Extract and order RNA editing sites located within an input genomic region.

Usage

```
GetSitesLocations(
  region_df,
  rnaEditMatrix,
  output = c("locationsOnly", "locationsAndValues")
)
```

Arguments

region_df   A data frame with the input genomic region. Please make sure columns seqnames,
                 start, and end are included in the data frame.

rnaEditMatrix   A matrix (or data frame) of RNA editing level values on individual sites, with
                 row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
                 as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).

output   Type of output data. Defaults to "locationsOnly".

Value

When output is set to "locationsOnly", a data frame of extracted and ordered RNA editing sites with columns chr and pos will be returned.

When output is set to "locationsAndValues", a data frame of RNA editing level values from the extracted and ordered sites will be returned. Please note that site IDs will be in row names of the output data frame.

Examples

```
data (rnaedit_df)

exm_region <- data.frame(
   seqnames = "chr1",
   start =  28000000,
   end = 28826881,
   stringsAsFactors = FALSE
)

GetSitesLocations(
   region_df = exm_region,
   rnaEditMatrix = rnaedit_df,
   output = "locationsOnly"
)[1:3, ]
```

MakeModelFormula   *Make model formula.*

Description

Make model formula for different types of phenotype responses.

Usage

```
MakeModelFormula(
  responses_char,
  covariates_char = NULL,
  respType = c("binary", "continuous", "survival")
)
```

Arguments

responses_char   A character vector of the response variable.

covariates_char

              A character vector of the covariate variables.

respType   Type of outcome. Defaults to "binary".

Details

When `respType` is set as `"survival"`, `"surv_object"` is only a placeholder here , which will be defined later in `TestSingleRegion()`.

Value

A character vector of the model formula.

Examples

```
MakeModelFormula(
  responses_char = "age",
  covariates_char = c("sex", "tumor_type"),
  respType = "continuous"
)

MakeModelFormula(
  responses_char = "sample_type",
  covariates_char = c("sex", "tumor_type"),
  respType = "binary"
)

MakeModelFormula(
  responses_char = c("OS.time", "OS"),
  covariates_char = c("sex", "tumor_type"),
  respType = "survival"
)
```

MarkCoeditedSites   *Mark RNA editing sites in contiguous and co-edited region.*

Description

Mark RNA editing sites in contiguous and co-edited region by selecting sites for which r_drop values calculated from inner function [CreateRdrop](#) is greater than rDropThresh_num.

Usage

```
MarkCoeditedSites(
  rnaEditCluster_mat,
  rDropThresh_num = 0.4,
  method = c("spearman", "pearson"),
  minEditFreq = 0.05,
  verbose = TRUE
)
```

Arguments

rnaEditCluster_mat

A matrix of RNA editing level values on individual sites, with row names as sample IDs and column names as site IDs in the form of "chrAA:XXXXXXXX".

rDropThresh_num

Threshold for minimum correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites. Please set a number between 0 and 1. Defaults to 0.4.

method   Method for computing correlation. Defaults to "spearman".

minEditFreq Threshold for minimum percentage of edited samples for a given site. The `r_drop` value of the sites with frequency lower than `minEditFreq` will be set as NA. Please set a number between 0 and 1. Defaults to 0.05.

verbose   Should messages and warnings be displayed? Defaults to TRUE.

Details

`r_drop` statistic is used to identify co-edited sites. An outlier site (keep = 0) in a genomic region typically has low correlation with the rest of the sites in a genomic region. The sites with `r_drop` value greater than `rDropThresh_num` are marked to have keep = 1. Please see [CreateRdrop](#) for more details.

Value

A data frame with the following columns:

- `site` : site ID.

- `r_drop` : The correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.

- `keep` : indicator for co-edited sites, The sites with keep = 1 belong to the contiguous and co-edited region.

- `keep_contiguous` : contiguous co-edited region number

- `site` : site ID.

- `keep` : indicator for co-edited sites, The sites with keep = 1 belong to the contiguous and co-edited region.

- `ind` : index for the sites.

- `r_drop` : the correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.

See Also

[CreateRdrop](#)

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

MarkCoeditedSites(
  rnaEditCluster_mat = exm_data,
  method = "spearman"
)
```

OrderSitesByLocation   *Order RNA editing sites by their genomic locations.*

Description

Split RNA editing sites locations into chromosomes and positions, and then order the sites by their genomic locations.

Usage

```
OrderSitesByLocation(sites_char, output = c("dataframe", "vector"))
```

Arguments

sites_char   A character vector of RNA editing sites. site IDs should be in the form of "chrAA:XXXXXXXX".

output   Type of output data. Defaults to "dataframe".

Value

Depends on the output type. When output is set as "vector", a character vector of ordered input RNA editing sites will be returned. When output is set as "dataframe", a data frame of ordered RNA editing sites with following columns will be returned:

- site : site ID.
- chr : chromosome number.
- pos : genomic location.

Examples

```
exm_sites <- c(
  "chr22:41327462", "chr22:24969087",
  "chr22:29538891", "chr22:45736763"
)

OrderSitesByLocation(
  sites_char = exm_sites,
  output = "dataframe"
)
```

PlotEditingCorrelations

Plotting correlations of RNA editing levels within a region.

Description

Plotting correlations of RNA editing levels within a region.

Usage

```
PlotEditingCorrelations(region_gr, rnaEditMatrix, ...)
```

Arguments

region_gr   A GRanges object of a region.

rnaEditMatrix A matrix (or data frame) of RNA editing level values on individual sites, with
               row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
               as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).

...   Dots for additional internal arguments, see corrplot for details.

Value

(Invisibly) returns a reordered correlation matrix.

Examples

```
data(rnaedit_df)

genes_gr <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

exm_regions <- AllCoeditedRegions(
  regions_gr = genes_gr,
```

```
    rnaEditMatrix = rnaedit_df,
    output = "GRanges",
    method = "spearman"
)

PlotEditingCorrelations(
    region_gr = exm_regions[1],
    rnaEditMatrix = rnaedit_df
)
```

RegionSummaryMethod   *Methods to summarize RNA editing levels from multiple sites within a single region.*

Description

Summarize RNA editing sites in a single region by taking maximum, mean, median or first principal component.

Usage

```
MaxSites(rnaEditMatrix, ...)

MeanSites(rnaEditMatrix, ...)

MedianSites(rnaEditMatrix, ...)

PC1Sites(rnaEditMatrix, ...)
```

Arguments

rnaEditMatrix   A matrix (or data frame) of RNA editing level values on individual sites, with
                 row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
                 as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).
...   Dots for additional internal arguments (currently unused).

Value

A named numeric vector of summarized RNA editing levels with sample IDs as names.

Examples

```
    data(rnaedit_df)
    MedianSites(rnaEditMatrix = rnaedit_df)[1:3]
```

---

rnaedit_df	*Example breast cancer RNA editing dataset.*

Description

A subset of the TCGA breast cancer RNA editing dataset for 272 edited sites on genes PHACTR4, CCR5, METTL7A and a few randomly sampled sites for 221 subjects.

Usage

```
rnaedit_df
```

Format

A data frame containing RNA editing levels for 272 sites (in the rows) for 221 subjects (in the columns). Row names are site IDs and column names are sample IDs.

Source

Synapse database ID: syn2374375.

SingleCloseByRegion	*Extracts clusters of RNA editing sites located closely in a single genomic region.*

Description

Extracts clusters of RNA editing sites located closely in an input genomic region.

Usage

```
SingleCloseByRegion(region_df, rnaEditMatrix, maxGap = 50, minSites = 3)
```

Arguments

region_df	A data frame with the input genomic region. Please make sure columns seqnames, start, and end are included in the data frame.
rnaEditMatrix	A matrix (or data frame) of RNA editing level values for individual sites, with row names as site IDs in the form of "chrAA:XXXXXXXX", and column names as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).
maxGap	An integer, genomic locations within maxGap from each other are placed into the same cluster. Defaults to 50.
minSites	An integer, minimum number of edited sites for a cluster to be selected for output. Defaults to 3.

Details

The algorithm of this function is based on the clusterMaker function in the bumphunter R package. Each cluster is essentially a group of sites such that two consecutive sites in the cluster are separated by less than maxGap.

Value

A GRanges object containing genomic locations of RNA editing sites located closely within the single input pre-defined genomic region.

Examples

```
data(rnaedit_df)

exm_region <- data.frame(
  seqnames = "chr1",
  start =  28691093,
  end = 28826881,
  stringsAsFactors = FALSE
)

SingleCloseByRegion(
  region_df = exm_region,
  rnaEditMatrix = rnaedit_df,
  maxGap = 50,
  minSites = 3
)
```

SingleCoeditedRegion   *Extracts contiguous co-edited genomic regions from a single genomic region.*

Description

Extracts contiguous co-edited genomic regions from an input genomic region.

Usage

```
SingleCoeditedRegion(
  region_df,
  rnaEditMatrix,
  output = c("GRanges", "dataframe"),
  rDropThresh_num = 0.4,
  minPairCorr = 0.1,
  minSites = 3,
  method = c("spearman", "pearson"),
  minEditFreq = 0.05,
```

```
    returnAllSites = FALSE,
    verbose = TRUE
)
```

Arguments

region_df	A data frame with the input genomic region. Please make sure columns seqnames, start, and end are included in the data frame.
rnaEditMatrix	A matrix (or data frame) of RNA editing level values on individual sites, with row names as site IDs in the form of "chrAA:XXXXXXXX", and column names as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).
output	Type of output data, can be "GRanges" or "dataframe". Defaults to "GRanges".
rDropThresh_num	
	Threshold for minimum correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites. Please set a number between 0 and 1. Defaults to 0.4.
minPairCorr	Minimum pairwise correlation coefficient of a cluster is used as a filter to select clusters for output. Only clusters with all pairwise correlations between sites more than minPairCorr will be selected for output. To use this filter, set this argument to a number between -1 and 1 (defaults to 0.1). To turn it off, please set the argument to -1.
minSites	Minimum number of sites to be considered a region. Only regions with more than minSites number of sites will be returned.
method	Method for computing correlations. Defaults to "spearman".
minEditFreq	Threshold for minimum percentage of samples for a given site. The r_drop value of the sites with frequency lower than minEditFreq will be set as NA. Please set a number between 0 and 1. Defaults to 0.05.
returnAllSites	When no co-edited region is found in an input genomic region, returnAllSites = TRUE indicates outputting all the sites from the input region, while returnAllSites = FALSE indicates not returning any site from the input region. Defaults to FALSE.
verbose	Should messages and warnings be displayed? Defaults to TRUE, but is set to FALSE when called from within AllCoeditedRegions().

Value

When output is set to "GRanges", a GRanges object with seqnames, ranges and strand of the contiguous co-edited regions will be returned.

When output is set to "dataframe", a data frame with following columns will be returned:

- site : site ID.
- chr : chromosome.
- pos : genomic location.
- r_drop : the correlation between RNA editing levels of one site and the mean RNA editing levels of the rest of the sites.

- keep : indicator for co-edited sites, The sites with keep = 1 belong to the contiguous and co-edited region.

- keep_contiguous : contiguous co-edited region number.

- regionMinPairwiseCor : the minimum pairwise correlation of a co-edited region.

- keep_regionMinPairwiseCor : equals 1 for contiguous co-edited subregions. The regions with keepminPairwiseCor = 1 are the ones that passed the regionMinPairwiseCor filter and will be returned as a co-edited sub-region.

Examples

```
data(rnaedit_df)

exm_region <- data.frame(
  seqnames = "chr1",
  start =  28691093,
  end = 28826881,
  stringsAsFactors = FALSE
)

SingleCoeditedRegion(
  region_df = exm_region,
  rnaEditMatrix = rnaedit_df,
  minPairCorr = 0.25,
  output = "dataframe",
  method = "spearman"
)
```

SitesToRegion   *Create output data in the format of GRanges.*

Description

Output contiguous co-edited subregions found by FindCorrelatedRegions function and filtered by GetMinPairwiseCor function.

Usage

```
SitesToRegion(
  sitesSubregion_df,
  sitesAreOrdered = TRUE,
  keepminPairwiseCor_df,
  returnAllSites = FALSE,
  verbose = TRUE
)
```

Arguments

sitesSubregion_df

> An output data frame from function FindCorrelatedRegions with variables
> `site`, `subregion`. Please see [FindCorrelatedRegions](#) for details.

sitesAreOrdered

> Are the sites in `sitesSubregion_df` ordered by location? Defaults to FALSE.

keepminPairwiseCor_df

> An output data frame from function GetMinPairwiseCor with variables `subregion`,
> `keepminPairwiseCor` and `minPairwiseCor`. Please see [GetMinPairwiseCor](#)
> for details.

returnAllSites   When no contiguous co-edited regions are found in a input genomic region,
> `returnAllSites = TRUE` indicates outputting all the sites in this input region,
> while `returnAllSites = FALSE` indicates not returning any site in this input
> region. Defaults to FALSE.

verbose   Should messages and warnings be displayed? Defaults to TRUE.

Value

A GRanges object with `seqnames`, `ranges` and `strand` of the contiguous co-edited regions.

Examples

```
data(t_rnaedit_df)

ordered_cols <- OrderSitesByLocation(
  sites_char = colnames(t_rnaedit_df),
  output = "vector"
)
exm_data <- t_rnaedit_df[, ordered_cols]

exm_sites <- MarkCoeditedSites(
  rnaEditCluster_mat = exm_data,
  method = "spearman"
)

exm_regions <- FindCorrelatedRegions(
  sites_df = exm_sites,
  featureType = "site"
)

exm_sites <- split(
  x = exm_regions$site,
  f = exm_regions$subregion
)

exm_cor <- GetMinPairwiseCor(
  rnaEditCluster_mat = exm_data,
  minPairCorr = 0.1,
  probes_ls = exm_sites,
  method = "spearman"
```

```
)

SitesToRegion(
  sitesSubregion_df = exm_regions,
  keepminPairwiseCor_df = exm_cor$keepminPairwiseCor_df
)
```

SummarizeAllRegions   *Summarize RNA editing levels from multiple sites in regions.*

Description

A wrapper function to summarize RNA editing levels from multiple sites in regions.

Usage

```
SummarizeAllRegions(
  regions_gr,
  rnaEditMatrix,
  selectMethod = MedianSites,
  progressBar = "time",
  ...
)
```

Arguments

regions_gr   A GRanges object of input genomic regions.

rnaEditMatrix   A matrix (or data frame) of RNA editing level values for individual sites, with
                row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
                as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).

selectMethod   Method for summarizing regions. Available options are "MaxSites", "MeanSites",
                "MedianSites", "PC1Sites". Please see [RegionSummaryMethod](#) for more de-
                tails.

progressBar   Name of the progress bar to use. There are currently five types of progress
                bars: "time", "none", "text", "tk", and "win". Defaults to "time". See
                [create_progress_bar](#) for more details.

...   Dots for additional internal arguments (currently unused).

Value

A data frame of the class rnaEdit_df, includes variables seqnames, start, end, width and sum-
marized RNA editing levels in each sample.

See Also

[TransformToGR](#), [AllCloseByRegions](#), [AllCoeditedRegions](#), [CreateEditingTable](#), [TestAssociations](#),
[AnnotateResults](#)

Examples

```
data(rnaedit_df)

genes_gr <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

exm_regions <- AllCoeditedRegions(
  regions_gr = genes_gr,
  rnaEditMatrix = rnaedit_df,
  output = "GRanges",
  method = "spearman"
)

SummarizeAllRegions(
  regions_gr = exm_regions,
  rnaEditMatrix = rnaedit_df
)[1:3, 1:6]
```

SummarizeSingleRegion   *Summarizes RNA editing levels from multiple sites in a single region.*

Description

Summarizes RNA editing levels from multiple sites in an input region.

Usage

```
SummarizeSingleRegion(
  region_df,
  rnaEditMatrix,
  selectMethod = MedianSites,
  ...
)
```

Arguments

region_df   A data frame with the input genomic region. Please make sure columns seqnames,
                start, and end are included in the data frame.

rnaEditMatrix   A matrix (or data frame) of RNA editing level values for individual sites, with
                row names as site IDs in the form of "chrAA:XXXXXXXX", and column names
                as sample IDs. Please make sure to follow the format of example dataset (data(rnaedit_df)).

selectMethod   Method for summarizing regions. Available options are "MaxSites", "MeanSites",
                "MedianSites", "PC1Sites". Please see RegionSummaryMethod for more de-
                tails.

...   Dots for additional internal arguments (currently unused).

Value

A named numeric vector of summarized RNA editing levels with sample IDs as column names.

Examples

```
data(rnaedit_df)

exm_region <- data.frame(
  seqnames = "chr1",
  start =  28691093,
  end = 28826881,
  stringsAsFactors = FALSE
)

SummarizeSingleRegion(
  region_df = exm_region,
  rnaEditMatrix = rnaedit_df
)[1:3]
```

TestAssociations   *Test associations between phenotype and RNA editing levels.*

Description

A wrapper function to test associations between phenotype and RNA editing levels in single-site analysis or summarized RNA editing levels in region-based analysis.

Usage

```
TestAssociations(
  rnaEdit_df,
  pheno_df,
  responses_char,
  covariates_char = NULL,
  respType = c("binary", "continuous", "survival"),
  progressBar = "time",
  orderByPval = TRUE
)
```

Arguments

rnaEdit_df   A data frame with class rnaEdit_df, which is a output from function [CreateEditingTable]()
                or function [SummarizeAllRegions](). This data frame should include RNA
                editing level values, with row names as site IDs or region IDs, and column names
                as sample IDs.

pheno_df   A data frame with phenotype and covariates, which should include all the sam-
                  ples in rnaEdit_df. Please make sure the input pheno_df has the variable
                  named "sample" to indicate sample IDs.

responses_char   A character vector of names of response variables in pheno_df. When respType
                  is set as "survival", responses_char should have length 2. The first element
                  must be the name of the variable with following up time, and the second ele-
                  ment must be status indicator. Status indicator should be coded as 0/1(1=death),
                  TRUE/FALSE(TRUE=death), or 1/2(death). Please make sure variable names
                  are in this order. We have not tested this code on interval-censored data; use at
                  your own risk. See [Surv](#) for more details.

covariates_char
                  A character vector of names of covariate variables in pheno_df.

respType   Type of outcome. Defaults to "binary".

progressBar   Name of the progress bar to use. There are currently five types of progress
                  bars: "time", "none", "text", "tk", and "win". Defaults to "time". See
                  [create_progress_bar](#) for more details.

orderByPval   Sort co-edited regions by model p-value or not? Defaults to TRUE.

Value

A data frame with locations of the genomic regions or sites (seqnames, start, end, width),
test statistics (estimate, stdErr or coef, exp_coef, se_coef), pValue and false discovery rate
(fdr).

See Also

[TransformToGR](#), [AllCloseByRegions](#), [AllCoeditedRegions](#), [CreateEditingTable](#), [SummarizeAllRegions](#),
[AnnotateResults](#)

Examples

```
data(rnaedit_df)

genes_gr <- TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

exm_regions <- AllCoeditedRegions(
  regions_gr = genes_gr,
  rnaEditMatrix = rnaedit_df,
  output = "GRanges",
  method = "spearman"
)

sum_regions <- SummarizeAllRegions(
  regions_gr = exm_regions,
  rnaEditMatrix = rnaedit_df,
```

```
    selectMethod = MaxSites
  )

  exm_pheno <- readRDS(
    system.file(
    "extdata",
    "pheno_df.RDS",
    package = 'rnaEditr',
    mustWork = TRUE
    )
  )

  TestAssociations(
    rnaEdit_df = sum_regions,
    pheno_df = exm_pheno,
    responses_char = "sample_type",
    covariates_char = NULL,
    respType = "binary"
  )
```

TestSingleRegion   *Test associations between phenotype and RNA editing levels.*

Description

Test associations between phenotype and RNA editing levels in a single site or summarized RNA editing levels in a single region.

Usage

```
TestSingleRegion(
  rnaEdit_num,
  modelPrep_ls,
  respType = c("binary", "continuous", "survival")
)
```

Arguments

rnaEdit_num   A named numeric vector of (summarized) RNA editing level values with sample IDs as names.

modelPrep_ls   A list includes modelFormula_char which is created by function MakeModelFormula, pheno_df which is the input phenotype data frame in TestAssociations, and minSize (minimum sample size per group to use regular logistic regression) which is created by function CountSamplesPerGroup when respType is "binary".

respType   Type of outcome. Defaults to "binary".

Details

minSize is used by function `TestSingleRegion` to decide on whether to use regular logistic regression or Firth corrected logistic regression (`"https://www.jstor.org/stable/2336755"`).

Value

a dataframe with test statistics (`estimate`, `stdErr`, `pValue` or `coef`, `exp_coef`, `se_coef`, `pValue`).

Examples

```
data(rnaedit_df)

exm_pheno <- readRDS(
  system.file(
  "extdata",
  "pheno_df.RDS",
  package = 'rnaEditr',
  mustWork = TRUE
  )
)

exm_model <- list(
  modelFormula_char = "age_at_diagnosis ~ rnaEditSummary",
  pheno_df = exm_pheno,
  minSize = NULL
)

TestSingleRegion(
  rnaEdit_num = unlist(rnaedit_df[2,]),
  modelPrep_ls = exm_model,
  respType = "continuous"
)
```

TransformToGR   *Transform gene symbols or region ranges into GRanges object.*

Description

Transform a character vector of gene symbols or region ranges into a GRanges object.

Usage

```
TransformToGR(
  genes_char,
  type = c("symbol", "region"),
  genome = c("hg38", "hg19")
)
```

Arguments

genes_char   A character vector of gene symbols or region ranges. If you select type to be `"symbol"`, then please make sure your input of genes_char is in the format of c("ABCB10", "PEX26"). If you select type to be `"region"`, then please make sure your input of genes_char is in the format of c("chr1:33772367-33791699", "chr22:18555686-18573797").

type   What is the type of genes_char. Can be `"symbol"` (default) or `"region"`.

genome   Use `"hg19"` or `"hg38"` gene reference. Defaults to `"hg38"`. It's only used when type is set to `"symbol"`

Details

`TransformToGR()` uses the hg19/hg38 genes to associate gene symbols with their genomic region ranges. The pre-processed dataset is saved in inst/extdata in this package.

Users who wish to add gene symbols to the GRanges created using function `TransformToGR()` can use function AddMetaData(). Please see [AddMetaData](#) for details.

Value

A GRanges object with seqnames, ranges and strand.

See Also

[AllCloseByRegions](#), [AllCoeditedRegions](#), [CreateEditingTable](#), [SummarizeAllRegions](#), [TestAssociations](#), [AnnotateResults](#)

Examples

```
TransformToGR(
  genes_char = c("PHACTR4", "CCR5", "METTL7A"),
  type = "symbol",
  genome = "hg19"
)

TransformToGR(
  genes_char = c("chr22:18555686-18573797", "chr22:36883233-36908148"),
  type = "region",
  genome = "hg19"
)
```

---

t_rnaedit_df *Transposed breast cancer example dataset.*

Description

A subset of the TCGA breast cancer RNA editing dataset for 20 randomly selected RNA editing sites and 50 randomly selected subjects from example dataset rnaedit_df. Please note that this is only a computational testing dataset for inner functions of this package. To test main functions, please use dataset rnaedit_df instead.

Usage

    t_rnaedit_df

Format

A data frame containing RNA editing levels for 50 subjects (in the rows) at 20 edited sites (in the columns). Row names are sample IDs and column names are site IDs.

Source

Synapse database ID: syn2374375.

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