

# Package ‘NormqPCR’

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**Version** 1.53.0

**Date** 2018-07-09

**Title** Functions for normalisation of RT-qPCR data

**Description** Functions for the selection of optimal reference genes and the normalisation of real-time quantitative PCR data.

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**Depends** R(>= 2.14.0), stats, RColorBrewer, Biobase, methods, ReadqPCR, qpcR

**biocViews** MicrotitrePlateAssay, GeneExpression, qPCR

**License** LGPL-3

**LazyLoad** yes

**LazyData** yes

**URL** [www.bioconductor.org/packages/release/bioc/html/NormqPCR.html](http://www.bioconductor.org/packages/release/bioc/html/NormqPCR.html)

**Collate** allGenerics.R deltaCt.R selectHKs.R combineTechReps.R  
combineTechRepsWithSD.R CqValues.R deltaDeltaCt.R  
stabMeasureM.R dealWithNA.R geomMean.R stabMeasureRho.R  
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NormqPCR-package	<i>Functions for normalisation of RT-qPCR data.</i>
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## Description

Functions for normalisation of real-time quantitative PCR data.

## Details

Package: NormqPCR  
 Type: Package  
 Version: 1.7.1  
 Date: 2014-08-13  
 Depends: R(>= 2.14.0), stats, RColorBrewer, Biobase, methods, ReadqPCR, qpcR  
 Imports: ReadqPCR  
 bioViews: MicrotitrePlateAssay, GeneExpression, qPCR  
 License: LGPL-3  
 LazyLoad: yes  
 LazyData: yes

require(NormqPCR)

## Author(s)

Matthias Kohl, James Perkins, Nor Izayu Abdul Rahman

Maintainer: James Perkins <jimrperkins@gmail.com>

## References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

Jo Vandesompele, Katleen De Preter, Filip Pattyn et al. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology* 2002. 3(7):research0034.1-0034.11. <http://genomebiology.com/2002/3/7/research/0034/>

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *CANCER RESEARCH* 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

Kenneth Livak, Thomase Schmittgen (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the 2<sup>^</sup>ddCt Method. *Methods* 25, 402-408, 2001 <http://www.ncbi.nlm.nih.gov/pubmed/11846609>

## Examples

```
## some examples are given in the vignette
## Not run:
library(NormqPCR)
vignette("NormqPCR")

## End(Not run)
```

---

Bladder

*Bladder dataset of Andersen et al (2004)*

---

## Description

This dataset was used in Andersen et al (2004) to demonstrate normalization of real-time quantitative RT-PCR data by geometric averaging of housekeeping genes.

## Usage

```
data(Bladder)
```

## Format

A qPCRBatch object which contains an expression matrix with the expression of 14 genes measured in 28 samples. The sample information is saved in the phenoData slot with variables

Sample.no. sample number.

Grade Grade of bladder cancer.

The following information on the measured genes is saved in the variables `Symbol` and `Gene.name` of the `featureData` slot.

ATP5B ATP synthase, H<sup>+</sup> transporting, mitochondrial F1 complex, beta polypeptide.

HSPCB Heat shock 90-kDa protein 1, beta.

S100A6 S100 calcium-binding protein A6 (calcylin).

FL0T2 Flotillin 2.

TEGT Testis enhanced gene transcript (BAX inhibitor 1).

UBB Ubiquitin B.

TPT1 Tumor protein, translationally controlled 1.

CFL1 Cofilin 1 (non-muscle).

ACTB Actin, beta.

RPS13 Ribosomal protein S13.

RPS23 Ribosomal protein S23.

GAPD Glyceraldehyde-3-phosphate dehydrogenase.

UBC Ubiquitin C.

FLJ20030 Hypothetical protein FLJ20030.

For a detailed annotation see Table 1 in Anderson et al. (2004).

### Details

The genes included in this data set were selected by screening 99 bladder sample expression profiles.

### Source

The data set was obtained from [http://www.mdl.dk/Publications\\_sup1.htm](http://www.mdl.dk/Publications_sup1.htm)

### References

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *CANCER RESEARCH* 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

### Examples

```
data(Bladder)
Bladder
head(exprs(Bladder))
pData(Bladder)
fData(Bladder)
```

---

BladderRepro

*Dataset of Andersen et al (2004)*

---

### Description

This dataset was used in Andersen et al (2004) to demonstrate normalization of real-time quantitative RT-PCR data by geometric averaging of housekeeping genes.

### Usage

```
data(BladderRepro)
```

### Format

A `qPCRBatch` object which contains an expression matrix with the expression of 8 genes measured in 26 samples. The sample information is saved in the `phenoData` slot with variables

`Sample.no.` sample number.

`Grade` Grade of bladder cancer.

The following information on the measured genes is saved in the variables `Symbol` and `Gene.name` of the `featureData` slot.

`CD14` CD14 antigen.

`FCN1` Ficolin (collagen/fibrinogen domain containing) 1.

`CCNG2` Cyclin G2.

`NPAS2` Neuronal PAS domain protein 2.

`UBC` Ubiquitin C.

`CFL1` Cofilin 1 (non-muscle).

`ACTB` Actin, beta.

`GAPD` Glyceraldehyde-3-phosphate dehydrogenase.

For a detailed annotation see Table 1 and Supplementary table 1 in Anderson et al. (2004).

### Details

This data set was used to check the reproducibility of the results obtained in Andersen et al (2004).

### Source

The data set was obtained from [http://www.mdl.dk/Publications\\_sup1.htm](http://www.mdl.dk/Publications_sup1.htm)

### References

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *CANCER RESEARCH* 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

**Examples**

```

data(BladderRepro)
BladderRepro
head(exprs(BladderRepro))
pData(BladderRepro)
fData(BladderRepro)

```

Colon

*Colon dataset of Andersen et al (2004)***Description**

This dataset was used in Andersen et al (2004) to demonstrate normalization of real-time quantitative RT-PCR data by geometric averaging of housekeeping genes.

**Usage**

```
data(Colon)
```

**Format**

A qPCRBatch object which contains an expression matrix with the expression of 13 genes measured in 40 samples. The sample information is saved in the phenoData slot with variables

Sample.no. sample number.

Classification Classification of colon cancer.

The following information on the measured genes is saved in the variables Symbol and Gene.name of the featureData slot.

UBC Ubiquitin C.

UBB Ubiquitin B.

SUI1 Putative translation initiation factor.

NACA Nascent-polypeptide-associated complex alpha polypeptide.

FLJ20030 Hypothetical protein FLJ20030.

CFL1 Cofilin 1 (non-muscle).

ACTB Actin, beta.

CLTC Clathrin, heavy polypeptide (Hc).

RPS13 Ribosomal protein S13.

RPS23 Ribosomal protein S23.

GAPD Glyceraldehyde-3-phosphate dehydrogenase.

TPT1 Tumor protein, translationally controlled 1.

TUBA6 Tubulin alpha 6.

For a detailed annotation see Table 1 in Anderson et al. (2004).

**Details**

The genes included in this data set were selected by screening 161 colon sample expression profiles.

**Source**

The data set was obtained from [http://www.mdl.dk/Publications\\_sup1.htm](http://www.mdl.dk/Publications_sup1.htm)

**References**

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. CANCER RESEARCH 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

**Examples**

```
data(Colon)
Colon
head(exprs(Colon))
pData(Colon)
fData(Colon)
```

---

combineTechReps

*Combines Technical Replicates*

---

**Description**

Takes expression set of qPCR values containing technical replicates and combines them.

**Usage**

```
combineTechReps(qPCRBatch, ...)

## S4 method for signature 'qPCRBatch'
combineTechReps(qPCRBatch, calc="arith")
```

**Arguments**

qPCRBatch	Expression set containing qPCR data, read in by a ReadqPCR function and containing technical reps, denoted by <code>_TechRep.n</code> suffix.
...	Extra arguments, detailed below
calc	use median, arithmetic or geometric mean for combining the values

**Details**

Takes `exprs` of qPCR values containing technical replicates and combines them using a specified centrality measure.

**Value**

qPCRBatch with same number of samples, but with less features, since all technical replicates are replaced with a single value of their means.

**Author(s)**

James Perkins <jimrperkins@gmail.com>

**References**

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**Examples**

```
path <- system.file("exData", package = "NormqPCR")
qPCR.example.techReps <- file.path(path, "qPCR.techReps.txt")
qPCRBatch.qPCR.techReps <- read.qPCR(qPCR.example.techReps)
rownames(exprs(qPCRBatch.qPCR.techReps))
combinedTechReps <- combineTechReps(qPCRBatch.qPCR.techReps)
rownames(exprs(combinedTechReps))
```

---

combineTechRepsWithSD *Combines Technical Replicates*

---

**Description**

Takes expression set of qPCR values containing technical replicates and combines them. In addition the appropriate standard deviation (SD) is computed.

**Usage**

```
combineTechRepsWithSD(qPCRBatch, ...)

## S4 method for signature 'qPCRBatch'
combineTechRepsWithSD(qPCRBatch, calc="arith")
```

**Arguments**

qPCRBatch	Expression set containing qPCR data, read in by a ReadqPCR function and containing technical reps, denoted by <code>_TechRep.n</code> suffix.
...	Extra arguments, detailed below
calc	use median, arithmetic or geometric mean for combining the values



### Details

Takes `exprs` of qPCR values containing technical replicates and combines them using a specified centrality measure.

The arithmetic mean (`calc="arith"`) is combined with the classical standard deviation. In case of the geometric mean (`calc="geom"`) the classical standard deviation of the log-values is exponentiated. The median (`calc="median"`) is calculated in connection with the MAD.

### Value

`qPCRBatch` with same number of samples, but with less features, since all technical replicates are replaced with a single value of their means. In addition the slot `assayData` includes a matrix with SD values which can be accessed via `se.exprs`.

### Author(s)

Matthias Kohl <Matthias.Kohl@stamats.de>

### References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### See Also

[combineTechReps](#)

### Examples

```
path <- system.file("exData", package = "NormqPCR")
qPCR.example.techReps <- file.path(path, "qPCR.techReps.txt")
qPCRBatch.qPCR.techReps <- read.qPCR(qPCR.example.techReps)
rownames(exprs(qPCRBatch.qPCR.techReps))
combinedTechReps <- combineTechRepsWithSD(qPCRBatch.qPCR.techReps)
rownames(exprs(combinedTechReps))
exprs(combinedTechReps)
se.exprs(combinedTechReps)
```

### Description

This function computes normalized relative quantities (NRQs) for a `qPCRBatch`.

**Usage**

```
ComputeNRQs(qPCRBatch, ...)  
## S4 method for signature 'qPCRBatch'  
ComputeNRQs(qPCRBatch, hkgs)
```

**Arguments**

qPCRBatch	an object of class <a href="#">qPCRBatch</a> .
hkgs	Names of reference/housekeeping genes.
...	other parameters to be passed to downstream methods.

**Details**

Allows the user to normalized relative quantities as defined in Hellemanns et al. (2007).

**Value**

Object of class "qPCRBatch".

**Author(s)**

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

**References**

Jan Hellemans, Geert Mortier, Anne De Paepe, Frank Speleman and Jo Vandesompele (2007). qBase relative quantification framework and software for management and automated analysis of real-time quantitative PCR data. *Genome Biology*, 8:R19

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**See Also**

[qPCRBatch-class](#)

**Examples**

```
## Example data  
path <- system.file("exData", package = "ReadqPCR")  
qPCR.example <- file.path(path, "qPCR.example.txt")  
Cq.data <- read.qPCR(qPCR.example)  
  
## combine technical replicates  
Cq.data1 <- combineTechRepsWithSD(Cq.data)  
  
## add efficiencies  
Efs <- file.path(path, "Efficiencies.txt")  
Cq.effs <- read.table(file = Efs, row.names = 1, header = TRUE)
```

```

rownames(Cq.effs) <- featureNames(Cq.data1)
effs(Cq.data1) <- as.matrix(Cq.effs[,"efficiency",drop = FALSE])
se.effs(Cq.data1) <- as.matrix(Cq.effs[,"SD.efficiency",drop = FALSE])

##
res <- ComputeNRQs(Cq.data1, hkgs = c("gene_az", "gene_gx"))
## NRQs
exprs(res)
## SD of NRQs
se.exprs(res)

```

CqValues

*Compute Cq value and amplification efficiency***Description**

This function calculates Cq value and amplification efficiency for a `CyclesSet`. It is based on function `pcrbatch` of package **qpcR**.

**Usage**

```

CqValues(object, ...)
## S4 method for signature 'CyclesSet'
CqValues(object, Effmethod = "expfit", group = NULL,
          model = 15, check = "uni2", checkPAR = parKOD(),
          remove = "none", exclude = NULL, type = "cpD2",
          labels = NULL, norm = FALSE, baseline = NULL,
          basefac = 1, smooth = NULL,
          smoothPAR = list(span = 0.1),
          factor = 1, opt = FALSE,
          optPAR = list(sig.level = 0.05, crit = "ftest"),
          plot = FALSE, verbose = FALSE, ...)

```

**Arguments**

<code>object</code>	an object of class <code>CyclesSet</code> .
<code>Effmethod</code>	a character vector defining the methods for computing amplification efficiency.
<code>group</code>	a vector containing the grouping for possible replicates.
<code>model</code>	the model to be used for all runs. Default model is 15.
<code>check</code>	the method for kinetic outlier detection in <code>KOD</code> . Method "uni2" is set as default which is a test on sigmoidal structure.
<code>checkPAR</code>	parameters to be supplied to the check method. See <code>parKOD</code> .
<code>remove</code>	indicates which runs to be removed. Either none of them, those which failed to fit or from the outlier methods.
<code>exclude</code>	indicates samples to be excluded from calculation, either "" for samples with missing column names or a regular expression defining columns (samples); see 'Details' and 'Examples' in <code>modlist</code> .

type	the point on the amplification curve which is used for efficiency estimation; see <a href="#">efficiency</a> .
labels	a vector containing labels which define replicate groups. See more details in <a href="#">pcrbatch</a> and <a href="#">ratiobatch</a> .
norm	a logical value which determines whether the raw data should be normalized within [0, 1] before model fitting or not.
baseline	type of baseline subtraction. More details in <a href="#">efficiency</a> .
basefac	a factor when using averaged baseline cycles, such as 0.95.
smooth	the curve smoothing method. See more details in <a href="#">pcrbatch</a> .
smoothPAR	parameters to be supplied to smoothing method in <code>smooth</code> .
factor	a multiplication factor for the fluorescence response values.
opt	a logical value which determines whether model selection should be applied to each model or not.
optPAR	parameters to be supplied for model selection in <a href="#">mselect</a> .
plot	a logical value. If TRUE, the single runs are plotted from the internal <code>modlist</code> for diagnostics.
verbose	a logical value. If TRUE, fitting and tagging results will be displayed in the console.
...	other parameters to be passed to downstream methods.

### Details

Allows the user to compute Cq value and amplification efficiency. In addition, all values generated during the computations are saved. This function has four choices of methods for computing amplification efficiency values which are the methods provided by package **qpcR**.

More details on technical replication and normalization is given in the vignette `NormqPCR`.

### Value

Object of class "qPCRBatch".

### Author(s)

Nor Izayu Abdul Rahman, Matthias Kohl <Matthias.Kohl@stamats.de>

### References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orenge, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### See Also

[pcrbatch](#), [CyclesSet-class](#), [qPCRBatch-class](#)

## Examples

```
## Read in the raw qPCR data from file "LC480_Example.txt"
path <- system.file("exData", package = "ReadqPCR")
LC480.example <- file.path(path, "LC480_Example.txt")
cycData <- read.LC480(file = LC480.example)

## Read in the sample information data from file "LC480_Example_SampleInfo.txt".
LC480.SamInfo <- file.path(path, "LC480_Example_SampleInfo.txt")
samInfo <- read.LC480SampleInfo(LC480.SamInfo)

## Merge information
cycData1 <- merge(cycData, samInfo)

## Compute Cq values
## 1) use sigmoidal model
res1 <- CqValues(cycData1, Effmethod = "sigfit")
res1
effs(res1)
se.effs(res1)

## 2) fit exponential model (default)
res2 <- CqValues(cycData1, Effmethod = "expfit")
res2
effs(res2)
se.effs(res2)

## 3) use window of linearity
res3 <- CqValues(cycData1, Effmethod = "sliwin")
res3
effs(res3)
se.effs(res3)

## 4) linear regression of efficiency
res4 <- CqValues(cycData1, Effmethod = "LRE")
res4
effs(res4)
se.effs(res4)
```

---

deltaCt

*Perform normalization with a given housekeeping gene*

---

## Description

Normalise qPCR eset using a given housekeeping gene as control, then perform differential expression analysis using the delta delta Ct method

## Usage

```
deltaCt(qPCRBatch, ...)
```

```
## S4 method for signature 'qPCRBatch'
deltaCt(qPCRBatch, hkgs, combineHkgs=FALSE, calc="arith")
deltaCq(qPCRBatch, hkgs, combineHkgs=FALSE, calc="arith")
```

### Arguments

qPCRBatch	qPCR-specific expression set, containing qPCR data.
...	Extra arguments, detailed below
hkgs	String containing the name of the name of the housekeeping gene which will be used to normalise the rest of the genes.
combineHkgs	Logical - if TRUE, then as long as more than one housekeeper given for argument hkgs, it will combine the housekeepers by finding the geometric mean. Housekeepers can be found using geNorm or NormFinder algorithms.
calc	use arithmetic or geometric mean.

### Details

Takes expression set of qPCR values and normalises them using a housekeeping gene. Returns a qPCRBatch with exprs set of the same dimensions but with the given hkg value subtracted.

### Value

qPCRBatch with exprs set of the same dimensions but with the given hkg value subtracted.

### Author(s)

James Perkins <jimrperkins@gmail.com>

### References

Kenneth Livak, Thomase Schmittgen (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the  $2^{-\Delta\Delta C_t}$  Method. *Methods* 25, 402-408, 2001 <http://www.ncbi.nlm.nih.gov/pubmed/11846609>

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### See Also

selectHkgs, deltaDeltaCq

### Examples

```
path <- system.file("exData", package = "NormqPCR")
taqman.example <- file.path(path, "example.txt")
qPCRBatch.taqman <- read.taqman(taqman.example)
hkgs<-"Actb-Rn00667869_m1"
qPCRBatch.norm <- deltaCq(qPCRBatch = qPCRBatch.taqman, hkgs = hkgs, calc="arith")
head(exprs(qPCRBatch.norm))
```

---

deltaDeltaCt	<i>Perform normalization and differential expression with given house-keeping gene</i>
--------------	--

---

### Description

Normalise qPCRBatch RT-qPCR data using housekeeping genes as control, then perform differential expression analysis using the delta delta Cq method.

### Usage

```
deltaDeltaCt(qPCRBatch,...)
## S4 method for signature 'qPCRBatch'
deltaDeltaCt(qPCRBatch, maxNACase=0, maxNAControl=0, hkgs, contrastM,
             case, control, paired=TRUE, hkgCalc="arith", statCalc="arith")
deltaDeltaCq(qPCRBatch, maxNACase=0, maxNAControl=0, hkgs, contrastM,
             case, control, paired=TRUE, hkgCalc="arith", statCalc="arith")
```

### Arguments

qPCRBatch	qPCR-specific expression set, containing qPCR data.
...	Extra arguments, detailed below
maxNACase	Maximum number of NA values allowed before a detector's reading is discarded for samples designated as case.
maxNAControl	Maximum number of NA values allowed before a detector's reading is discarded for samples designated as control.
hkgs	String containing the name of the housekeeping gene which will be used to normalise the rest of the genes.
contrastM	A binary matrix which designates case and control samples.
case	The name of the column in contrastM that corresponds to the case samples.
control	The name of the column in contrastM that corresponds to the control samples.
paired	Logical - if TRUE the detectors and housekeepers in the same sample will be paired for calculating standard deviation, effectively meaning we will be calculating standard deviation of the differences. If FALSE, there will be no pairing, and standard deviation will be pooled between the detector and housekeepers.
hkgCalc	String - either "arith" or "geom", details how the different housekeeper genes should be combined - either by using the arithmetic or geometric mean.
statCalc	String - either "arith" or "geom", details how genes should be combined - either by using the arithmetic or geometric mean.

### Details

Takes expression set of qPCR values and normalises them using different housekeeping genes. Returns separate sets of values for each housekeeping gene given.

**Value**

matrix with columns containing the detector ids,  $2^{\Delta Cq}$  values for the sample of interest and the callibrator sample, alongside their respective standard deviations, the  $2^{\Delta\Delta Cq}$  values and the minimum and maximum values (ie the values that are 1 sd away )

**Author(s)**

James Perkins <jimrperkins@gmail.com>

**References**

Kenneth Livak, Thomase Schmittgen (2001). Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the  $2^{\Delta\Delta Cq}$  Method. *Methods* 25, 402-408, 2001 <http://www.ncbi.nlm.nih.gov/pubmed/11846609>

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**See Also**

selectHKs, deltaCq

**Examples**

```
path <- system.file("exData", package = "NormqPCR")
taqman.example <- file.path(path, "example.txt")
qPCRBatch.taqman <- read.taqman(taqman.example)
hkg <- "Actb-Rn00667869_m1"

contM <- cbind(c(0,0,1,1,0,0,1,1),c(1,1,0,0,1,1,0,0))
colnames(contM) <- c("interestingPhenotype", "wildTypePhenotype")
rownames(contM) <- sampleNames(qPCRBatch.taqman)

ddCq.taqman <- deltaDeltaCq(qPCRBatch = qPCRBatch.taqman, maxNAcase=1,
                           maxNAControl=1, hkg=hkg, contrastM=contM,
                           case="interestingPhenotype",
                           control="wildTypePhenotype",
                           statCalc="geom", hkgCalc="arith")

head(ddCq.taqman)
```

---

geNorm

*Data set of Vandesompele et al (2002)*

---

**Description**

This data set was used in Vandesompele et al (2002) to demonstrate normalization of real-time quantitative RT-PCR data by geometric averaging of housekeeping genes.



**Usage**

```
data(geNorm)
```

**Format**

A qPCRBatch object which contains an expression matrix with 85 observations on the following 10 variables which stand for expression data of ten potential reference/housekeeping genes

ACTB actin, beta

B2M beta-2-microglobulin

GAPD glyceraldehyde-3-phosphate dehydrogenase

HMBS hydroxymethylbilane synthase

HPRT1 hypoxanthine phosphoribosyltransferase 1

RPL13A ribosomal protein L13a

SDHA succinate dehydrogenase complex subunit A

TBP TATA box binding protein

UBC ubiquitin C

YWHAZ tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, zeta polypeptide

**Details**

The row names of this data set indicate the various human tissues which were investigated.

**BM** 9 normal bone-marrow samples

**POOL** 9 normal human tissues from pooled organs (heart, brain, fetal brain, lung, trachea, kidney, mammary gland, small intestine and uterus)

**FIB** 20 short-term cultured normal fibroblast samples from different individuals

**LEU** 13 normal leukocyte samples

**NB** 34 neuroblastoma cell lines (independently prepared in different labs from different patients)

**Source**

The data set was obtained from <http://genomebiology.com/content/supplementary/gb-2002-3-7-research0034-s1.txt>

**References**

Jo Vandesompele, Katleen De Preter, Filip Pattyn et al. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology* 2002. 3(7):research0034.1-0034.11. <http://genomebiology.com/2002/3/7/research/0034/>

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**Examples**

```
data(geNorm)
str(exprs(geNorm.qPCRBatch))
sampleNames(geNorm.qPCRBatch)
```

---

geomMean	<i>Geometric Mean</i>
----------	-----------------------

---

**Description**

Computation of the geometric mean.

**Usage**

```
geomMean(x, na.rm = TRUE)
```

**Arguments**

x	numeric vector of non-negative Reals
na.rm	a logical value indicating whether NA values should be stripped before the computation proceeds.

**Details**

The computation of the geometric mean is done via  $\text{prod}(x)^{(1/\text{length}(x))}$ .

**Value**

geometric mean

**Note**

A first version of this function appeared in package SLqPCR.

**Author(s)**

Matthias Kohl <Matthias.Kohl@stamats.de>

**References**

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**Examples**

```
x <- rlnorm(100)
geomMean(x)
```

---

makeAllNAs	<i>Make all Cq values NA</i>
------------	------------------------------

---

### Description

Make all Cq values for a given detector NA when the number of NAs for that detector is above a given threshold

### Usage

```
makeAllNAs(qPCRBatch, ...)  
  
## S4 method for signature 'qPCRBatch'  
makeAllNAs(qPCRBatch, contrastM, sampleMaxM)
```

### Arguments

qPCRBatch	Expression set containing qPCR data.
...	Extra arguments, detailed below
contrastM	Contrast Matrix like that used in <code>limma</code> . Columns represent the different samples types, rows are the different samples, with a 1 or 0 in the matrix indicating which sample types the different samples belong to.
sampleMaxM	Sample Max Matrix. Columns represent the different sample types. There is one value per column, which represents the max number of NAs allowed for that sample type.

### Details

Make all NAs when number of NAs above a given threshold

### Value

qPCRBatch object with a new `exprs` slot, everything else equal

### Author(s)

James Perkins <jimrperkins@gmail.com>

### References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**Examples**

```

# read in the data
path <- system.file("exData", package = "NormqPCR")
taqman.example <- file.path(path, "example.txt")
qPCRBatch.taqman <- read.taqman(taqman.example)
exprs(qPCRBatch.taqman)["Cc120.Rn00570287_m1",] # values before

# make contrastM
a <- c(0,0,1,1,0,0,1,1) # one for each sample type, with 1 representing
b <- c(1,1,0,0,1,1,0,0) # position of sample type in the samplenames vector
contM <- cbind(a,b)
colnames(contM) <- c("case","control") # then give the names of each sample type
rownames(contM) <- sampleNames(qPCRBatch.taqman) # and the rows of the matrix
contM

# make sampleMaxM
sMaxM <- t(as.matrix(c(3,3))) # now make the sample max matrix
colnames(sMaxM) <- c("case","control") # make sure these line up with samples
sMaxM

# function
qPCRBatch.taqman.replaced <- makeAllNAs(qPCRBatch.taqman, contM, sMaxM)
exprs(qPCRBatch.taqman.replaced)["Cc120.Rn00570287_m1",]

```

---

makeAllNewVal

*Make all Cq values NA*


---

**Description**

Make all Cq values for a given detector NA when the number of NAs for that detector is above a given threshold

**Usage**

```

makeAllNewVal(qPCRBatch, ...)

## S4 method for signature 'qPCRBatch'
makeAllNewVal(qPCRBatch, contrastM, sampleMaxM, newVal)

```

**Arguments**

qPCRBatch	Expression set containing qPCR data.
...	Extra arguments, detailed below
contrastM	Contrast Matrix like that used in <code>limma</code> . Columns represent the different samples types, rows are the different samples, with a 1 or 0 in the matrix indicating which sample types the different samples belong to.

sampleMaxM	Sample Max Matrix. Columns represent the different sample types. There is one value per column, which represents the max number of NAs allowed for that sample type.
newVal	New value to give the values in the group where the NAs are above the threshold.

### Details

Make all a given value when number of NAs above a given threshold, with different thresholds for the different sample classes, using sMaxM and contM to provide this information, as detailed below.

### Value

qPCRBatch object with a new exprs slot, everything else equal

### Author(s)

James Perkins <jimrperkins@gmail.com>

### References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### Examples

```
# read in the data
path <- system.file("exData", package = "NormqPCR")
taqman.example <- file.path(path, "example.txt")
qPCRBatch.taqman <- read.taqman(taqman.example)
exprs(qPCRBatch.taqman)["Cc120.Rn00570287_m1",] # values before

# make contrastM
a <- c(0,0,1,1,0,0,1,1) # one for each sample type, with 1 representing
b <- c(1,1,0,0,1,1,0,0) # position of sample type in the samplenames vector
contM <- cbind(a,b)
colnames(contM) <- c("case","control") # then give the names of each sample type
rownames(contM) <- sampleNames(qPCRBatch.taqman) # and the rows of the matrix
contM

# make sampleMaxM
sMaxM <- t(as.matrix(c(3,3))) # now make the sample max matrix
colnames(sMaxM) <- c("case","control") # make sure these line up with samples
sMaxM

# function
qPCRBatch.taqman.replaced <- makeAllNewVal(qPCRBatch.taqman, contM, sMaxM)
exprs(qPCRBatch.taqman.replaced)["Cc120.Rn00570287_m1",]
```

---

replaceAboveCutOff      *Replace Cq values with new value*

---

### Description

Replace Cq values above a given threshold with a new value

### Usage

```
replaceAboveCutOff(qPCRBatch, ...)
```

```
## S4 method for signature 'qPCRBatch'
replaceAboveCutOff(qPCRBatch, newVal=NA, cutOff=38)
```

### Arguments

qPCRBatch	Expression set containing qPCR data.
...	Extra arguments, detailed below
newVal	The new value with which to replace the values above the cutoff
cutOff	the minimal threshold above which the values will be replaced

### Details

Replaces values in the exprs slot of the qPCRBatch object that are above a threshold value with a new number

### Value

qPCRBatch object with a new exprs slot

### Author(s)

James Perkins <jimrperkins@gmail.com>

### References

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### Examples

```
path <- system.file("exData", package = "NormqPCR")
taqman.example <- file.path(path, "example.txt")
qPCRBatch.taqman <- read.taqman(taqman.example)
exprs(qPCRBatch.taqman)["Cc120.Rn00570287_m1",]
qPCRBatch.taqman.replaced <- replaceAboveCutOff(qPCRBatch.taqman, newVal = NA, cutOff = 35)
exprs(qPCRBatch.taqman.replaced)["Cc120.Rn00570287_m1",]
```

---

replaceNAs	<i>Replace NAs with a given value</i>
------------	---------------------------------------

---

**Description**

Replace NAs with a given value

**Usage**

```
replaceNAs(qPCRBatch, ...)  
  
## S4 method for signature 'qPCRBatch'  
replaceNAs(qPCRBatch, newNA)
```

**Arguments**

qPCRBatch	Expression set containing qPCR data.
...	Extra arguments, detailed below
newNA	The new value to replace the NAs with

**Details**

Replaces NA values in the exprs slot of the qPCRBatch object with a given number

**Value**

qPCRBatch object with a new exprs slot

**Author(s)**

James Perkins <jimrperkins@gmail.com>

**References**

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**Examples**

```
path <- system.file("exData", package = "NormqPCR")  
taqman.example <- file.path(path, "example.txt")  
qPCRBatch.taqman <- read.taqman(taqman.example)  
qPCRBatch.taqman.replaced <- replaceNAs(qPCRBatch.taqman, newNA = 40)  
exprs(qPCRBatch.taqman.replaced)["Cc120.Rn00570287_m1",]
```

---

selectHKs

*Selection of reference/housekeeping genes*


---

## Description

This function can be used to determine a set of reference/housekeeping (HK) genes for gene expression experiments

## Usage

```
selectHKs(qPCRBatch, ...)

## S4 method for signature 'matrix'
selectHKs(qPCRBatch, group, method = "geNorm", minNrHKs = 2, log = TRUE, Symbols,
          trace = TRUE, na.rm = TRUE)

## S4 method for signature 'qPCRBatch'
selectHKs(qPCRBatch, group, method = "geNorm", minNrHKs = 2, log = TRUE, Symbols,
          trace = TRUE, na.rm = TRUE)
```

## Arguments

qPCRBatch	matrix or qPCRBatch, containing the data (expression matrix) in the exprs slot
...	Extra arguments, detailed below
group	optional factor not used by all methods, hence may be missing
method	method to compute most stable genes
minNrHKs	minimum number of HK genes that should be considered
log	logical: is data on log-scale
Symbols	gene symbols
trace	logical, print additional information
na.rm	a logical value indicating whether NA values should be stripped before the computation proceeds.

## Details

This function can be used to determine a set of reference/housekeeping (HK) genes for gene expression experiments. The default method "geNorm" was proposed by Vandesompele et al. (2002).

Currently, the geNorm method by Vandesompele et al. (2002) and the NormFinder method of Andersen et al. (2004) are implemented.

Vandesompele et al. (2002) propose a cut-off value of 0.15 for the pairwise variation. Below this value the inclusion of an additional housekeeping gene is not required.



**Value**

If method = "geNorm" a list with the following components is returned

ranking	ranking of genes from best to worst where the two most stable genes cannot be ranked
variation	pairwise variation during stepwise selection
meanM	average expression stability M

If method = "NormFinder" a list with the following components is returned

ranking	ranking of genes from best to worst where the two most stable genes cannot be ranked
rho	stability measure rho of Andersen et al. (2004)

**Author(s)**

Matthias Kohl <Matthias.Kohl@stamats.de>

**References**

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

Jo Vandesompele, Katleen De Preter, Filip Pattyn et al. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology* 2002. 3(7):research0034.1-0034.11. <http://genomebiology.com/2002/3/7/research/0034/>

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *CANCER RESEARCH* 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

**Examples**

```
data(geNorm)
tissue <- as.factor(c(rep("BM", 9), rep("FIB", 20), rep("LEU", 13),
                    rep("NB", 34), rep("POOL", 9)))
res.BM <- selectHKs(geNorm.qPCRBatch[,tissue == "BM"], method = "geNorm",
                  Symbols = featureNames(geNorm.qPCRBatch), minNrHK = 2,
                  log = FALSE)
```

---

stabMeasureM	<i>Gene expression stability value M</i>
--------------	--

---

### Description

Computation of the gene expression stability value M for real-time quantitative RT-PCR data. For more details we refer to Vandesompele et al. (2002).

### Usage

```
stabMeasureM(x, log = TRUE, na.rm = TRUE)
```

### Arguments

x	matrix or data.frame containing real-time quantitative RT-PCR data
log	logical: is data on log-scale
na.rm	a logical value indicating whether NA values should be stripped before the computation proceeds.

### Details

The gene expression stability value M is defined as the average pairwise normalization factor; i.e., one needs to specify data from at least two genes. For more details see Vandesompele et al. (2002). Note this dispatches on a transposed expression matrix, not a qPCRBatch object since it is only called from within the selectHKs method.

### Value

numeric vector with gene expression stability values

### Author(s)

Matthias Kohl <Matthias.Kohl@stamats.de>

### References

Jo Vandesompele, Katleen De Preter, Filip Pattyn et al. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology* 2002. 3(7):research0034.1-0034.11. <http://genomebiology.com/2002/3/7/research/0034/>

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

### See Also

selectHKs

**Examples**

```
data(geNorm)
tissue <- as.factor(c(rep("BM", 9), rep("FIB", 20), rep("LEU", 13),
                    rep("NB", 34), rep("POOL", 9)))
res.BM <- selectHKs(geNorm.qPCRBatch[,tissue == "BM"], method = "geNorm",
                  Symbols = featureNames(geNorm.qPCRBatch), minNrHK = 2, log = FALSE)
```

---

stabMeasureRho

*Gene expression stability value rho*


---

**Description**

Computation of the gene expression stability value rho for real-time quantitative RT-PCR data. For more details we refer to Andersen et al. (2004).

**Usage**

```
stabMeasureRho(x,...)

## S4 method for signature 'x'
stabMeasureRho(x, group, log = TRUE, na.rm = TRUE, returnAll = FALSE)
```

**Arguments**

x	matrix containing real-time quantitative RT-PCR data, or qPCRBatch object
...	Extra arguments, detailed below
group	grouping factor, either a factor vector or a phenoData column called "Group"
log	logical: is data on log-scale
na.rm	a logical value indicating whether NA values should be stripped before the computation proceeds.
returnAll	logical, return additional information.

**Details**

The gene expression stability value rho is computed. For more details see Andersen et al. (2004).

**Value**

numeric vector with gene expression stability values

If returnAll == TRUE a list with the following components is returned

rho	stability measure rho of Andersen et al. (2004)
d	used by selectHKs
v	used by selectHKs

**Author(s)**

Matthias Kohl <Matthias.Kohl@stamats.de>

**References**

Claus Lindbjerg Andersen, Jens Ledet Jensen and Torben Falck Orntoft (2004). Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *CANCER RESEARCH* 64, 5245-5250, August 1, 2004. <http://cancerres.aacrjournals.org/cgi/content/full/64/15/5245>

Perkins, JR, Dawes, JM, McMahon, SB, Bennett, DL, Orengo, C, Kohl, M (2012). ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics*, **13**, 1:296.

**See Also**

selectHKs

**Examples**

```
data(Colon)
Class <- pData(Colon)[,"Classification"]
res.Colon <- stabMeasureRho(Colon, group = Class, log = FALSE)
sort(res.Colon) # cf. Table 3 in Andersen et al (2004)

data(Bladder)
Grade <- pData(Bladder)[,"Grade"]
res.Bladder <- stabMeasureRho(Bladder, group = Grade, log = FALSE)
sort(res.Bladder) # cf. Table 3 in Andersen et al (2004)
```

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