

Longitudinal Analysis of Cancer Evolution with LACE

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Overview. LACE (Longitudinal Analysis of Cancer Evolution) is an algorithmic framework that processes single-cell somatic mutation profiles from cancer samples collected at different time points and in distinct experimental settings, in order to describe the evolutionary history of the tumor. The tool can return an high resolution picture of clones' prevalence and their variations, e.g., because of therapies.

LACE can be employed to process single-cell mutational profiles as generated by calling variants from the increasingly available scRNA-seq data, such as the ones obtained by SMARTseq2 protocol.

The output of the method is a longitudinal tree that best fits the input data, modelling both phylogenetic constraints and sc-RNAseq specific noise. Moreover, the package provides a suite of functions to visualize and explore the results.

In this vignette, we give an overview of the package by presenting its main functions.

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1 Using the LACE R package

We now present an example of longitudinal analysis of cancer evolution with LACE using single-cell data obtained from Rambow, Florian, et al. "Toward minimal residual disease-directed therapy in melanoma." Cell 174.4 (2018): 843-855. The data comprises point mutations for four time points: (1) before treatment, (2) 4 days treatment, (3) 28 days treatment and finally (4) 57 days treatment.

We first load the data.

```
library("LACE")
data(longitudinal_sc_variants)
names(longitudinal_sc_variants)

## [1] "T1_before_treatment" "T2_4_days_treatment" "T3_28_days_treatment"
## [4] "T4_57_days_treatment"
```

The input data (D) can be either a list as shown above or a SummarizedExperiment object. In the latter case, the object needs to include genomic data (0: mutation absent, 1: mutation detected or NA: missing information) in assay field. Following guidelines, the rows of such matrix are variants, while columns are single cells concatenated for all the time points. Furthermore, a rowData field needs to be specified and must be a data.frame with a column named "TimePoint" reporting the label of the experiment for each single cell; we highlight that the ordering of the labels in rowData must match the one of the columns in the assay field. We next show an example.

```
library("SummarizedExperiment", attach.required=FALSE)
T1 = t(longitudinal_sc_variants[[1]])
T2 = t(longitudinal_sc_variants[[2]])
T3 = t(longitudinal_sc_variants[[3]])
T4 = t(longitudinal_sc_variants[[4]])
concat_time_point = cbind(T1,T2,T3,T4)
TimePointLabes = c(rep("T1", ncol(T1)),
                   rep("T2", ncol(T2)),
                   rep("T3", ncol(T3)),
                   rep("T4", ncol(T4)))
longitudinal_SE = SummarizedExperiment(assays = concat_time_point,
                                       colData = data.frame(TimePoint = TimePointLabes))

print(longitudinal_SE)

## class: SummarizedExperiment
## dim: 6 475
## metadata(0):
## assays(1): ''
## rownames(6): ARPC2_2_218249894_C_T CCT8_21_29063389_G_A ...
```

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```
## PRAME_22_22551005_T_A RPL5_1_92837514_C_G
## rowData names(0):
## colnames(475): SRR7424153 SRR7424154 ... SRR7424823 SRR7424824
## colData names(1): TimePoint
```

We setup the main parameter in order to perform the inference. First of all, as the three data points may potentially provide sequencing for an unbalanced number of cells, we weight each time point as follows $w_s = (1 - \frac{n_s}{n_T}) / (y - 1)$ in order to account for this. In the formula, e.g., the weight for time point s (w_s) is calculated based on the number of cells observed in the time point (n_s) and the total number of cells in the three time points (n_T). The denominator ($y - 1$, with y being the number of time points, i.e., 3 in our case) aims at normalizing the weights to sum to one.

```
lik_weights = c(0.2308772, 0.2554386, 0.2701754, 0.2435088)
```

The second main parameter to be defined as input is represented by the false positive and false negative error rates, i.e., alpha and beta. We can specify a different rate per time point as a list of rates. When multiple sets of rates are provided, LACE performs a grid search in order to estimate the best set of error rates.

```
alpha = list()
alpha[[1]] = c(0.02, 0.01, 0.01, 0.01)
alpha[[2]] = c(0.10, 0.05, 0.05, 0.05)
beta = list()
beta[[1]] = c(0.10, 0.05, 0.05, 0.05)
beta[[2]] = c(0.10, 0.05, 0.05, 0.05)
head(alpha)

## [[1]]
## [1] 0.02 0.01 0.01 0.01
##
## [[2]]
## [1] 0.10 0.05 0.05 0.05

head(beta)

## [[1]]
## [1] 0.10 0.05 0.05 0.05
##
## [[2]]
## [1] 0.10 0.05 0.05 0.05
```

We can now perform the inference as follows. Notice that `D` can be either the list longitudinal sc variants or the SummarizedExperiment longitudinal SE.

```
inference = LACE(D = longitudinal_sc_variants,
  lik_w = lik_weights,
  alpha = alpha,
  beta = beta,
  keep_equivalent = TRUE,
  num_rs = 5,
  num_iter = 10,
  n_try_bs = 5,
```

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```
num_processes = NA,
seed = 12345,
verbose = FALSE)

## [1] "B is already binary"
## [1] "Root is parent of Clone_1"
## [1] "Root is parent of Clone_2"
## [1] "Root is parent of Clone_3"
## [1] "Root is parent of Clone_4"
## [1] "Root is parent of Clone_5"
## [1] "Root is parent of Clone_6"
## [1] "Root is parent of Clone_1"
## [1] "Root is parent of Clone_2"
## [1] "Root is parent of Clone_3"
## [1] "Root is parent of Clone_4"
## [1] "Root is parent of Clone_5"
## [1] "Root is parent of Clone_6"
## [1] "nodes in B are sprouting from a single node"
## [1] "B is a full-rank matrix."
## [1] "Root Root 0"
## [1] "Root Clone_1 1"
## [1] "Root Clone_2 0"
## [1] "Root Clone_3 0"
## [1] "Root Clone_4 0"
## [1] "Root Clone_5 0"
## [1] "Root Clone_6 0"
## [1] "Clone_1 Clone_1 1"
## [1] "Clone_1 Clone_2 1"
## [1] "Clone_1 Clone_3 0"
## [1] "Clone_1 Clone_4 0"
## [1] "Clone_1 Clone_5 0"
## [1] "Clone_1 Clone_6 0"
## [1] "Clone_2 Clone_2 1"
## [1] "Clone_2 Clone_3 1"
## [1] "Clone_2 Clone_4 1"
## [1] "Clone_2 Clone_5 0"
## [1] "Clone_2 Clone_6 0"
## [1] "Clone_3 Clone_3 1"
## [1] "Clone_3 Clone_4 1"
## [1] "Clone_3 Clone_5 0"
## [1] "Clone_3 Clone_6 0"
## [1] "Clone_4 Clone_4 1"
## [1] "Clone_4 Clone_5 1"
## [1] "Clone_4 Clone_6 1"
## [1] "Clone_5 Clone_5 1"
## [1] "Clone_5 Clone_6 1"
## [1] "Clone_6 Clone_6 1"
##      [,1]
## [1,]    0
## [2,]    1
## [3,]    1
```

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```
## [4,] 1
## [5,] 1
## [6,] 1
## [7,] 1
##      [,1]
## [1,] 0
## [2,] 1
## [3,] 1
## [4,] 1
## [5,] 1
## [6,] 1
## [7,] 1
## [1] TRUE
## [1] "B represents a forest"
## [1] "B has a forest like structure"
## [1] "Root Root 0"
## [1] "Root Clone_1 1"
## [1] "Root Clone_2 0"
## [1] "Root Clone_3 0"
## [1] "Root Clone_4 0"
## [1] "Root Clone_5 0"
## [1] "Root Clone_6 0"
## [1] "Clone_1 Clone_1 1"
## [1] "Clone_1 Clone_2 1"
## [1] "Clone_1 Clone_3 0"
## [1] "Clone_1 Clone_4 0"
## [1] "Clone_1 Clone_5 0"
## [1] "Clone_1 Clone_6 0"
## [1] "Clone_2 Clone_2 1"
## [1] "Clone_2 Clone_3 1"
## [1] "Clone_2 Clone_4 1"
## [1] "Clone_2 Clone_5 0"
## [1] "Clone_2 Clone_6 0"
## [1] "Clone_3 Clone_3 1"
## [1] "Clone_3 Clone_4 1"
## [1] "Clone_3 Clone_5 0"
## [1] "Clone_3 Clone_6 0"
## [1] "Clone_4 Clone_4 1"
## [1] "Clone_4 Clone_5 1"
## [1] "Clone_4 Clone_6 1"
## [1] "Clone_5 Clone_5 1"
## [1] "Clone_5 Clone_6 1"
## [1] "Clone_6 Clone_6 1"
##      [,1]
## [1,] 0
## [2,] 1
## [3,] 1
## [4,] 1
## [5,] 1
## [6,] 1
## [7,] 1
```

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```

##      [,1]
## [1,]    0
## [2,]    1
## [3,]    1
## [4,]    1
## [5,]    1
## [6,]    1
## [7,]    1
## [1] TRUE
## [1] "B represents a forest"
## [1] "Root is parent of Clone_1"
## [1] "Root is parent of Clone_2"
## [1] "Root is parent of Clone_3"
## [1] "Root is parent of Clone_4"
## [1] "Root is parent of Clone_5"
## [1] "Root is parent of Clone_6"
## [1] "Root is parent of Clone_1"
## [1] "Root is parent of Clone_2"
## [1] "Root is parent of Clone_3"
## [1] "Root is parent of Clone_4"
## [1] "Root is parent of Clone_5"
## [1] "Root is parent of Clone_6"
## [1] "B is a binary matrix"
## [1] "B is a square matrix"
## [1] "B is a full rank matrix"
## [1] "B is a forest "
## [1] "B has single root"
## [1] "all checks done!"
## [1] "continue..."
## [1] "clonal tree:"
##
##      Root  ARPC2_2_218249894_C_T  PRAME_22_22551005_T_A
## Root      0                      1                      0
## ARPC2_2_218249894_C_T      0                      0                      1
## PRAME_22_22551005_T_A      0                      0                      0
## HNRNPC_14_21211843_C_T      0                      0                      0
## COL1A2_7_94422978_C_A      0                      0                      0
## RPL5_1_92837514_C_G      0                      0                      0
## CCT8_21_29063389_G_A      0                      0                      0
##
##      HNRNPC_14_21211843_C_T  COL1A2_7_94422978_C_A
## Root      0                      0
## ARPC2_2_218249894_C_T      0                      0
## PRAME_22_22551005_T_A      1                      1
## HNRNPC_14_21211843_C_T      0                      0
## COL1A2_7_94422978_C_A      0                      0
## RPL5_1_92837514_C_G      0                      0
## CCT8_21_29063389_G_A      0                      0
##
##      RPL5_1_92837514_C_G  CCT8_21_29063389_G_A
## Root      0                      0
## ARPC2_2_218249894_C_T      0                      0
## PRAME_22_22551005_T_A      0                      0
## HNRNPC_14_21211843_C_T      0                      0

```

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```

## COL1A2_7_94422978_C_A          1          1
## RPL5_1_92837514_C_G           0          0
## CCT8_21_29063389_G_A         0          0
##      Root ARPC2_2_218249894_C_T PRAME_22_22551005_T_A HNRNPC_14_21211843_C_T
## Root      1          0          0          0
## Clone_1    1          1          0          0
## Clone_2    1          1          1          0
## Clone_3    1          1          1          1
## Clone_4    1          1          1          0
## Clone_5    1          1          1          0
## Clone_6    1          1          1          0
##      COL1A2_7_94422978_C_A RPL5_1_92837514_C_G CCT8_21_29063389_G_A
## Root      0          0          0
## Clone_1    0          0          0
## Clone_2    0          0          0
## Clone_3    0          0          0
## Clone_4    1          0          0
## Clone_5    1          1          0
## Clone_6    1          0          1
## [1] "No incongruences found due to the chronological order"
## [1] "first time mutation occurrences based the chronological order of samples:"
##      Root Clone_1 Clone_2 Clone_3 Clone_4 Clone_5 Clone_6
##      1      1      1      1      1      1      1
## [1] "adjacent matrix with first time occurrences:"
##      Root_t1 Clone_1_t1 Clone_2_t1 Clone_3_t1 Clone_4_t1 Clone_5_t1 Clone_6_t1
## Root_t1      0          1          0          0          0          0          0
## Clone_1_t1    0          0          1          0          0          0          0
## Clone_2_t1    0          0          0          1          1          0          0
## Clone_3_t1    0          0          0          0          0          0          0
## Clone_4_t1    0          0          0          0          0          1          1
## Clone_5_t1    0          0          0          0          0          0          0
## Clone_6_t1    0          0          0          0          0          0          0
## [1] "time=" "1"
## [1] "1st Root_t1->Root_t1=1, prev_Root_t1=0.143835616438356: Root_t1->Root_t2=2"
## [1] "1st Root_t1->Root_t2=2, prev_Root_t2=0.162162162162162: Root_t2->Root_t3=2"
## [1] "1st Root_t1->Clone_1_t1=1, prev_Clone_1_t1=0.150684931506849: Clone_1_t1->Clone_1_t2=2"
## [1] "1st Clone_1_t1->Clone_1_t2=2, prev_Clone_1_t2=0.225225225225225: Clone_1_t2->Clone_1_t3=2"
## [1] "1st Clone_1_t1->Clone_2_t1=1, prev_Clone_2_t1=0.184931506849315: Clone_2_t1->Clone_2_t2=2"
## [1] "1st Clone_2_t1->Clone_2_t2=2, prev_Clone_2_t2=0.243243243243243: Clone_2_t2->Clone_2_t3=2"
## [1] "1st Clone_2_t1->Clone_3_t1=1, prev_Clone_3_t1=0.0753424657534247: Clone_3_t1->Clone_3_t2=2"
## [1] "1st Clone_2_t1->Clone_4_t1=1, prev_Clone_4_t1=0.205479452054795: Clone_4_t1->Clone_4_t2=2"
## [1] "1st Clone_3_t1->Clone_3_t2=2, prev_Clone_3_t2=0.108108108108108: Clone_3_t2->Clone_3_t3=2"
## [1] "1st Clone_4_t1->Clone_4_t2=2, prev_Clone_4_t2=0.135135135135135: Clone_4_t2->Clone_4_t3=2"
## [1] "1st Clone_4_t1->Clone_5_t1=1, prev_Clone_5_t1=0.143835616438356: Clone_5_t1->Clone_5_t2=2"
## [1] "1st Clone_4_t1->Clone_6_t1=1, prev_Clone_6_t1=0.0958904109589041: Clone_6_t1->Clone_6_t2=2"
## [1] "Clone_5_t1->Clone_5_t2=2, prev_Clone_5_t2=0: Clone_5_t1->Clone_5_t2=3"
## [1] "Clone_5_t1->Clone_5_t2=3, prev_Clone_5_t2=0: Clone_5_t2->Clone_5_t3=3"
## [1] "1st Clone_6_t1->Clone_6_t2=2, prev_Clone_6_t2=0.126126126126126: Clone_6_t2->Clone_6_t3=2"
## [1] "time=" "2"
## [1] "1st Root_t2->Root_t3=2, prev_Root_t3=0.133333333333333: Root_t3->Root_t4=2"
## [1] "1st Clone_1_t2->Clone_1_t3=2, prev_Clone_1_t3=0.166666666666667: Clone_1_t3->Clone_1_t4=2"

```

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```
## [1] "1st Clone_2_t2->Clone_2_t3=2, prev_Clone_2_t3=0.17777777777777778: Clone_2_t3->Clone_2_t4=2"
## [1] "1st Clone_3_t2->Clone_3_t3=2, prev_Clone_3_t3=0.05555555555555556: Clone_3_t3->Clone_3_t4=2"
## [1] "1st Clone_4_t2->Clone_4_t3=2, prev_Clone_4_t3=0.22222222222222222: Clone_4_t3->Clone_4_t4=2"
## [1] "1st Clone_5_t2->Clone_5_t3=3, prev_Clone_5_t3=0.03333333333333333: Clone_5_t3->Clone_5_t4=2"
## [1] "1st Clone_6_t2->Clone_6_t3=2, prev_Clone_6_t3=0.21111111111111111: Clone_6_t3->Clone_6_t4=2"
## [1] "time=" "3"
## [1] "time=" "4"
## [1] "time=" "3"
## [1] "time=" "2"
## [1] "time=" "1"
## [1] "idx"      "Clone_1"
## [1] "idx"      "Clone_2"
## [1] "idx"      "Clone_3"
## [1] "idx"      "Clone_4"
## [1] "idx"      "Clone_5"
## [1] "idx"      "Clone_6"
## [1] "/private/tmp/RtmpffRpiY/Rbuildald1238c6e8c/LACE/vignettes"
## [1] "info"
## [1] ""
```

We notice that the inference resulting on the command above should be considered only as an example; the parameters `num rs`, `num iter` and `n try bs` representing the number of steps performed during the inference are downscaled to reduce execution time. We refer to the Manual for discussion on default values. We provide within the package results of inferences performed with correct parameters as `RData`.

```
data(inference)
print(names(inference))

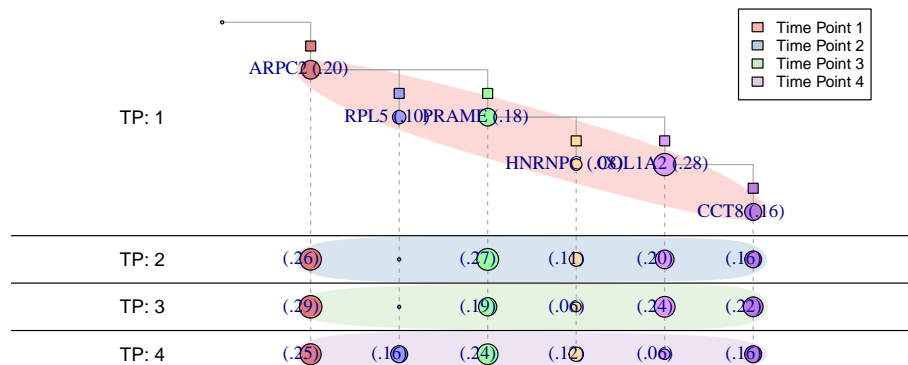
## [1] "B"                  "C"                  "corrected_genotypes"
## [4] "clones_prevalence"  "relative_likelihoods" "joint_likelihood"
## [7] "clones_summary"     "equivalent_solutions" "error_rates"
```

LACE returns a list of nine elements as results. Namely, B and C provide respectively the maximum likelihood longitudinal tree and cells attachments; corrected genotypes the corrected genotypes, clones prevalence, the estimated prevalence of any observed clone; relative likelihoods and joint likelihood the estimated likelihoods for each time point and the weighted likelihood; clones summary provide a summary of association of mutations to clones. In equivalent solutions, solutions (B and C) with likelihood equivalent to the best solution are returned; notice that in the example we disabled this feature by setting `equivalent_solutions` parameter to `FALSE`. Finally, error rates provide the best error rates (alpha and beta) as estimated by the grid search.

We can plot the inferred model using the function `longitudinal.tree.plot`.

```
clone_labels = c("ARPC2", "PRAME", "HNRNPC", "COL1A2", "RPL5", "CCT8")
longitudinal.tree = longitudinal.tree.plot(inference = inference,
                                           labels_show = "clones",
                                           clone_labels = clone_labels,
                                           legend_position = "topright")
```


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2 sessionInfo()

- R version 4.2.0 RC (2022-04-19 r82224), x86_64-apple-darwin17.0
- Locale: C/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
- Running under: macOS Mojave 10.14.6
- Matrix products: default
- BLAS:
 - /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRblas.0.dylib
- LAPACK:
 - /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRlapack.dylib
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: LACE 2.0.0, SummarizedExperiment 1.26.0, knitr 1.38
- Loaded via a namespace (and not attached): AnnotationDbi 1.58.0, Biobase 2.56.0, BiocFileCache 2.4.0, BiocGenerics 0.42.0, BiocManager 1.30.17, BiocStyle 2.24.0, Biostrings 2.64.0, DBI 1.1.2, DT 0.22, DelayedArray 0.22.0, GenomeInfoDb 1.32.0, GenomeInfoDbData 1.2.8, GenomicRanges 1.48.0, IRanges 2.30.0, KEGGREST 1.36.0, Matrix 1.4-1, MatrixGenerics 1.8.0, R6 2.5.1, RColorBrewer 1.1-3, RCurl 1.98-1.6, RSQLite 2.2.12, Rcpp 1.0.8.3, RcppTOML 0.1.7, RcppZigurat 0.1.6, Rfast 2.0.6, S4Vectors 0.34.0, XML 3.99-0.9, XVector 0.36.0, assertthat 0.2.1, biomaRt 2.52.0, bit 4.0.4, bit64 4.0.5, bitops 1.0-7, blob 1.2.3, bsplus 0.1.3, cachem 1.0.6, callr 3.7.0, cli 3.3.0, codetools 0.2-18, compiler 4.2.0, configr 0.3.5, crayon 1.5.1, curl 4.3.2, data.table 1.14.2, data.tree 1.0.0, dbplyr 2.1.1, digest 0.6.29, doParallel 1.0.17, dplyr 1.0.8, ellipsis 0.3.2, evaluate 0.15, fansi 1.0.3, fastmap 1.1.0, filelock 1.0.2, foreach 1.5.2, fs 1.5.2, generics 0.1.2, glue 1.6.2, grid 4.2.0, highr 0.9, hms 1.1.1, htmltools 0.5.2, htmlwidgets 1.5.4, httpuv 1.6.5, httr 1.4.2, igraph 1.3.1, ini 0.3.1, iterators 1.0.14, jsonlite 1.8.0, later 1.3.0, lattice 0.20-45, learnr 0.10.1, lifecycle 1.0.1, lubridate 1.8.0, magrittr 2.0.3, markdown 1.1, matrixStats 0.62.0, memoise 2.0.1, mime 0.12, parallel 4.2.0, pillar 1.7.0, pkgconfig 2.0.3, png 0.1-7, prettyunits 1.1.1, processx 3.5.3, progress 1.2.2, promises 1.2.0.1, ps 1.7.0, purrr 0.3.4, rappdirs 0.3.3, readr 2.1.2, rlang 1.0.2, rmarkdown 2.14, rprojroot 2.0.3, shiny 1.7.1, shinyBS 0.61.1, shinyFiles 0.9.1, shinydashboard 0.7.2, shinyjs 2.1.0, shinythemes 1.2.0,

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sortable 0.4.5, stats4 4.2.0, stringi 1.7.6, stringr 1.4.0, tibble 3.1.6, tidyr 1.2.0,
tidyselect 1.1.2, tools 4.2.0, tzdb 0.3.0, utf8 1.2.2, vctrs 0.4.1, withr 2.5.0, xfun 0.30,
xml2 1.3.3, xtable 1.8-4, yaml 2.3.5, zlibbioc 1.42.0