

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 33

> which(myresult$permutation_p<=0.05)

[1] 16 45 54 63 157 169 176 185 272 301 304 328 331 368 434 461 468 473 597
[20] 621 651 659 722 742 744 773 811 831 868 902 948 958 974

> sum(myresult$bootstrap_p<=0.05)

[1] 9

> which(myresult$bootstrap_p<=0.05)

[1] 61 153 185 230 288 368 409 710 742

> permutation_adjp <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adjp<=0.05)

[1] 5

> bootstrap_adjp <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adjp<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 42

> which(myresult2$bootstrap_p<=0.05)

[1] 17 43 44 47 89 91 98 105 119 156 189 221 226 230 263 331 414 419 436
[20] 446 477 513 520 543 580 584 607 609 639 673 674 679 731 763 778 786 802 849
[39] 864 872 890 921

> bootstrap2_adjp <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adjp<=0.05)

[1] 2

```

- Examples using the `RBM_F` function: `normdata_F` simulates a standardized gene expression data and `unifdata_F` simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```
> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)
```

	Length	Class	Mode
ordfit_t	3000	-none-	numeric
ordfit_pvalue	3000	-none-	numeric
ordfit_beta1	3000	-none-	numeric
permutation_p	3000	-none-	numeric
bootstrap_p	3000	-none-	numeric

```
> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 68

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 52

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 60

> which(myresult_F$permutation_p[, 1]<=0.05)
```

[1]	69	82	87	103	120	135	139	147	149	161	174	187	196	199	226
[16]	229	233	242	247	261	287	289	296	297	332	337	346	349	356	376
[31]	381	417	426	427	442	461	476	511	514	547	561	569	574	586	587
[46]	590	627	634	651	703	713	729	744	770	785	788	809	829	831	867
[61]	868	902	912	927	976	987	991	1000							

```
> which(myresult_F$permutation_p[, 2]<=0.05)
```

[1]	69	82	87	120	135	139	147	149	161	174	196	226	229	233	242	247	261	287	296
[20]	346	349	354	356	376	426	432	442	465	476	514	547	561	569	574	586	587	627	651
[39]	703	729	744	785	809	829	831	839	868	927	969	976	987	991					

```
> which(myresult_F$permutation_p[, 3]<=0.05)
```

[1]	72	82	120	135	147	149	161	174	196	199	226	233	242	247	261	287	296	324	332
[20]	337	346	349	356	376	399	426	427	442	461	476	511	514	547	561	569	574	586	587
[39]	627	648	703	713	744	770	785	788	809	813	829	831	839	868	892	907	908	966	969
[58]	976	987	991																

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 7

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 6

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 8

> which(con2_adjp<=0.05/3)

[1] 82 174 356 376 785 987

> which(con3_adjp<=0.05/3)

[1] 82 120 174 247 332 356 785 809

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p   3000   -none-  numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 50

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 69

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 58

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 20 26 29 101 134 148 169 194 250 263 268 288 309 317 321 333 335 383 391
[20] 484 493 529 532 539 547 551 557 582 637 649 650 668 696 730 736 763 785 808
[39] 814 817 818 831 856 876 879 906 928 929 930 933

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 7 20 26 29 48 94 101 107 148 169 194 202 219 225 250 263 288 309 317
[20] 321 335 378 383 391 404 426 484 486 493 529 532 533 539 542 547 551 557 577
[39] 585 637 649 650 668 671 685 693 696 721 736 746 763 766 798 814 817 818 824
[58] 831 856 876 879 886 906 928 929 930 950 985 998

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 7 20 29 94 101 148 169 194 197 202 250 263 288 309 317 321 333 335 373
[20] 378 383 391 479 484 486 493 526 529 539 551 557 571 649 650 668 685 696 721
[39] 730 736 763 766 814 817 818 824 831 876 879 906 928 929 930 933 950 966 976
[58] 978

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 6

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 5

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 9

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "C:/Users/biocbuild/bbs-3.10-bioc/tmpdir/RtmpOUcNbr/Rinst294c327d601/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1   Min.   :0.01058   Min.   :0.01187   Min.   :0.009103
cg00002426: 1   1st Qu.:0.04111   1st Qu.:0.04407   1st Qu.:0.041543
cg00003994: 1   Median :0.08284   Median :0.09531   Median :0.087042
cg00005847: 1   Mean    :0.27397   Mean    :0.28872   Mean    :0.283729
cg00006414: 1   3rd Qu.:0.52135   3rd Qu.:0.59032   3rd Qu.:0.558575
cg00007981: 1   Max.    :0.97069   Max.    :0.96937   Max.    :0.970155
(Other)      :994                NA's     :4
exmdata4[, 2]      exmdata5[, 2]      exmdata6[, 2]      exmdata7[, 2]
Min.   :0.01019   Min.   :0.01108   Min.   :0.01937   Min.   :0.01278
1st Qu.:0.04092   1st Qu.:0.04059   1st Qu.:0.05060   1st Qu.:0.04260
Median :0.09042   Median :0.08527   Median :0.09502   Median :0.09362
Mean    :0.28508   Mean    :0.28482   Mean    :0.27348   Mean    :0.27563
3rd Qu.:0.57502   3rd Qu.:0.57300   3rd Qu.:0.52099   3rd Qu.:0.52240
Max.    :0.96658   Max.    :0.97516   Max.    :0.96681   Max.    :0.95974
                NA's     :1
exmdata8[, 2]
Min.   :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000   -none- numeric
ordfit_pvalue 1000   -none- numeric
ordfit_beta0   1000   -none- numeric
ordfit_beta1   1000   -none- numeric
permutation_p 1000   -none- numeric
bootstrap_p    1000   -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 45

```

```

> sum(diff_results$permutation_p<=0.05)

[1] 39

> sum(diff_results$bootstrap_p<=0.05)

[1] 63

> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adjp<=0.05)

[1] 0

> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adjp<=0.05)

[1] 1

> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adjp<=0.05)

[1] 3

> diff_list_perm <- which(perm_adjp<=0.05)
> diff_list_boot <- which(boot_adjp<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
245 cg00224508 0.04479948  0.04972043  0.04152814  0.04189373
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
245  0.04208405  0.05284988  0.03775905  0.03955271
      diff_results$ordfit_t[diff_list_perm]
245                                1.962457
      diff_results$permutation_p[diff_list_perm]
245                                0

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
146 cg00134539 0.6110132  0.5332178  0.4599934  0.4678742
851 cg00830029 0.5836250  0.5939787  0.6473961  0.6726964
979 cg00945507 0.1343225  0.2385460  0.3474976  0.2890334
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
146  0.6719151  0.6313738  0.4792961  0.4542830
851  0.5082024  0.3465747  0.6627657  0.6463451
979  0.1184851  0.1665385  0.3071842  0.2662474

```

```

diff_results$ordfit_t[diff_list_boot]
146          5.394750
851         -2.841244
979         -4.750997
diff_results$bootstrap_p[diff_list_boot]
146          0
851          0
979          0

```