

segmentSeq: methods for identifying small RNA loci from high-throughput sequencing data

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1 Introduction

High-throughput sequencing technologies allow the production of large volumes of short sequences, which can be aligned to the genome to create a set of *matches* to the genome. By looking for regions of the genome which to which there are high densities of matches, we can infer a segmentation of the genome into regions of biological significance. The methods we propose allows the simultaneous segmentation of data from multiple samples, taking into account replicate data, in order to create a consensus segmentation. This has obvious applications in a number of classes of sequencing experiments, particularly in the discovery of small RNA loci and novel mRNA transcriptome discovery.

We approach the problem by considering a large set of potential *segments* upon the genome and counting the number of tags that match to that segment in multiple sequencing experiments (that may or may not contain replication). We then adapt the empirical Bayesian methods implemented in the baySeq package [1] to establish, for a given segment, the likelihood that the count data in that segment is similar to background levels, or that it is similar to the regions to the left or right of that segment. We then rank all the potential segments in order of increasing likelihood of similarity and reject those segments for which there is a high likelihood of similarity with the background or the regions to the left or right of the segment. This gives us a large list of overlapping segments. We reduce this list to identify non-overlapping loci by choosing, for a set of overlapping segments, the segment which has the lowest likelihood of similarity with either background or the regions to the left or right of that segment and rejecting all other segments that overlap with this segment. For fuller details of the method, see Hardcastle *et al.* [2].

2 Preparation

We begin by loading the segmentSeq package.

```
> library(segmentSeq)
```

Note that because the experiments that segmentSeq is designed to analyse are usually massive, we should use (if possible) parallel processing as implemented by the parallel package. If using this approach, we need to begin by define a *cluster*. The following command will use eight processors on a single machine; see the help page for 'makeCluster' for more information. If we don't want to parallelise, we can proceed anyway with a NULL cluster.

```
> if(require("parallel"))
+ {
+   numCores <- min(8, detectCores())
+   cl <- makeCluster(numCores)
+ } else {
+   cl <- NULL
+ }
```

The readGeneric function is able to read in tab-delimited files which have appropriate column names, and create an alignmentData object. Alternatively, if the appropriate column names are not present, we can specify which columns to use for the data. In either case, to use this function we pass a character vector of files, together with information on which data are to be treated as replicates to the function. We also need to define the lengths of

the chromosome and specify the chromosome names as a character. The data here, drawn from text files in the 'data' directory of the segmentSeq package are taken from the first million bases of an alignment to chromosome 1 and the first five hundred thousand bases of an alignment to chromosome 2 of *Arabidopsis thaliana* in a sequencing experiment where libraries 'SL9' and 'SL10' are replicates, as are 'SL26' and 'SL32'. Libraries 'SL9' and 'SL10' are sequenced from an Argonaute 6 IP, while 'SL26' and 'SL32' are an Argonaute 4 IP.

A similar function, readBAM performs the same operation on files in the BAM format. Please consult the help page for further details.

```
> chrlens <- c(1e6, 2e5)
> datadir <- system.file("extdata", package = "segmentSeq")
> libfiles <- c("SL9.txt", "SL10.txt", "SL26.txt", "SL32.txt")
> libnames <- c("SL9", "SL10", "SL26", "SL32")
> replicates <- c("AG06", "AG06", "AG04", "AG04")
> aD <- readGeneric(files = libfiles, dir = datadir,
+                   replicates = replicates, libnames = libnames,
+                   chrs = c(">Chr1", ">Chr2"), chrlens = chrlens,
+                   polyLength = 10, header = TRUE, gap = 200)
> aD
```

An object of class "alignmentData"
13765 rows and 4 columns

Slot "libnames":
[1] "SL9" "SL10" "SL26" "SL32"

Slot "replicates":
[1] AG06 AG06 AG04 AG04
Levels: AG04 AG06

Slot "alignments":
GRanges object with 13765 ranges and 2 metadata columns:

	seqnames	ranges	strand	tag	multireads
	<Rle>	<IRanges>	<Rle>	<character>	<numeric>
[1]	>Chr1	[265, 284]	-	AAATGAAGATAAACCATCCA	1
[2]	>Chr1	[405, 427]	-	AAGGAGTAAGAATGACAATAAAT	1
[3]	>Chr1	[406, 420]	-	AAGAATGACAATAAAA	1
[4]	>Chr1	[600, 623]	+	AAGGATTGGTGGTTTGAAGACACA	1
[5]	>Chr1	[665, 688]	+	ATCCTTGTAGCACACATTTGGCA	1
...
[13761]	>Chr2	[179972, 179993]	+	ATGAATGGCTCTCTCTAGCGGA	1
[13762]	>Chr2	[179978, 180000]	-	GAGATTCTCCGCTAGAGAGAGCC	1
[13763]	>Chr2	[179999, 180022]	-	ATTAATATTAATTCATCGGGAAGA	1
[13764]	>Chr2	[180002, 180022]	-	ATTAATATTAATTCATCGGGA	1
[13765]	>Chr2	[180014, 180037]	+	AATATTAATGTATTTGTGGAAAA	1

seqinfo: 2 sequences from an unspecified genome

Slot "data":
Matrix with 13765 rows.

	SL9	SL10	SL26	SL32
1	1	0	0	0
2	0	0	0	2
3	0	1	0	0
4	0	1	0	0
5	7	1	0	0
...
13761	2	7	0	0
13762	0	1	0	0
13763	0	1	0	0

```
13764  0    1    0    0
13765  1    0    0    0
```

```
Slot "libsizes":
[1] 4447 6531 9666 6675
```

Next, we process this alignmentData object to produce a segData object. This segData object contains a set of potential segments on the genome defined by the start and end points of regions of overlapping alignments in the alignmentData object. It then evaluates the number of tags that hit in each of these segments.

```
> sD <- processAD(aD, gap = 100, cl = cl)
> sD
```

```
An object of class "segData"
14444 rows and 4 columns
```

```
Slot "replicates":
[1] AG06 AG06 AG04 AG04
Levels: AG04 AG06
```

```
Slot "coordinates":
GRanges object with 14444 ranges and 0 metadata columns:
```

	seqnames	ranges	strand
	<Rle>	<IRanges>	<Rle>
[1]	>Chr1	[265, 284]	*
[2]	>Chr1	[405, 427]	*
[3]	>Chr1	[600, 623]	*
[4]	>Chr1	[600, 688]	*
[5]	>Chr1	[600, 830]	*
...
[14440]	>Chr2	[179708, 179872]	*
[14441]	>Chr2	[179708, 180037]	*
[14442]	>Chr2	[179738, 179872]	*
[14443]	>Chr2	[179738, 180037]	*
[14444]	>Chr2	[179923, 180037]	*

```
-----
seqinfo: 2 sequences from an unspecified genome
```

```
Slot "locLikelihoods" (stored on log scale):
Matrix with 0 rows.
<0 x 0 matrix>
```

```
Slot "data":
Matrix with 0 rows. Matrix with 0 rows.
SL9 SL10 SL26 SL32
```

```
Slot "libsizes":
[1] 4447 6531 9666 6675
```

We can now construct a segment map from these potential segments.

Segmentation by heuristic methods

A fast method of segmentation can be achieved by exploiting the bimodality of the densities of small RNAs in the potential segments. In this approach, we assign each potential segment to one of two clusters for each replicate group, either as a segment or a null based on the density of sequence tags within that segment. We then combine these clusterings for each replicate group to gain a consensus segmentation map.

```
> hS <- heuristicSeg(sD = sD, aD = aD, RKPM = 1000, largeness = 1e8, getLikes = TRUE, cl = cl)
```

.....

Segmentation by empirical Bayesian methods

A more refined approach to the problem uses an existing segment map (or, if not provided, a segment map defined by the `hS` function) to acquire empirical distributions on the density of sequence tags within a segment. We can then estimate posterior likelihoods for each potential segment as being either a true segment or a null. We then identify all potential segments in the with a posterior likelihood of being a segment greater than some value '`lociCutoff`' and containing no subregion with a posterior likelihood of being a null greater than '`nullCutoff`'. We then greedily select the longest segments satisfying these criteria that do not overlap with any other such segments in defining our segmentation map.

```
> classSegs <- classifySeg(sD = sD, aD = aD, cD = hS, cl = cl)
```

.....

```
> classSegs
```

GRanges object with 251 ranges and 0 metadata columns:

	seqnames	ranges	strand
	<Rle>	<IRanges>	<Rle>
[1]	>Chr1	[1, 664]	*
[2]	>Chr1	[665, 967]	*
[3]	>Chr1	[968, 17054]	*
[4]	>Chr1	[17055, 18728]	*
[5]	>Chr1	[18729, 27656]	*
...
[247]	>Chr2	[169231, 178343]	*
[248]	>Chr2	[178344, 178636]	*
[249]	>Chr2	[178637, 179707]	*
[250]	>Chr2	[179708, 180037]	*
[251]	>Chr2	[180038, 200000]	*

seqinfo: 2 sequences from an unspecified genome

An object of class "lociData"

251 rows and 4 columns

Slot "replicates"

AG06 AG06 AG04 AG04

Slot "groups":

[[1]]

[1] AG06 AG06 AG04 AG04

Levels: AG04 AG06

Slot "data":

	AG06.1	AG06.2	AG04.1	AG04.2
[1,]	1	2	0	2
[2,]	54	45	65	83
[3,]	2	3	0	0
[4,]	682	621	1405	1103
[5,]	0	3	0	0

246 more rows...

Slot "annotation":

data frame with 0 columns and 251 rows

Slot "locLikelihoods" (stored on log scale):

Matrix with 251 rows.

```

      AG04      AG06
1    0.24502  0.11169
2    0.95322  0.97731
3    0.034197 0.032662
4    0.98437  0.99646
5    0.036519 0.048366
...      ...      ...
247  0.29841  0.063213
248  0.91999  0.98423
249  0.052957 0.060674
250  0.96074  0.97484
251  0.033503 0.025245

```

Expected number of loci in each replicate group

```

      AG04      AG06
108.6738 137.6873

```

By one of these methods, we finally acquire an annotated lociData object, with the annotations describing the co-ordinates of each segment.

We can use this lociData object, in combination with the alignmentData object, to plot the segmented genome.

```

> par(mfrow = c(2,1), mar = c(2,6,2,2))
> plotGenome(aD, hS, chr = ">Chr1", limits = c(1, 1e5),
+           showNumber = FALSE, cap = 50)
> plotGenome(aD, classSegs, chr = ">Chr1", limits = c(1, 1e5),
+           showNumber = FALSE, cap = 50)

```

Given the calculated likelihoods, we can filter the segmented genome by controlling on likelihood, false discovery rate, or familywise error rate

```

> loci <- selectLoci(classSegs, FDR = 0.05)
> loci

```

GRanges object with 122 ranges and 0 metadata columns:

```

      seqnames      ranges strand
      <Rle>      <IRanges> <Rle>
[1]    >Chr1    [ 665,  967]      *
[2]    >Chr1   [17055, 18728]      *
[3]    >Chr1   [42217, 42390]      *
[4]    >Chr1   [44710, 44811]      *
[5]    >Chr1   [76799, 76890]      *
...      ...      ...      ...
[118]  >Chr2 [144202, 144327]      *
[119]  >Chr2 [152150, 152173]      *
[120]  >Chr2 [169196, 169230]      *
[121]  >Chr2 [178344, 178636]      *
[122]  >Chr2 [179708, 180037]      *
-----

```

seqinfo: 2 sequences from an unspecified genome

An object of class "lociData"

122 rows and 4 columns

Slot "replicates"

AG06 AG06 AG04 AG04

Slot "groups":

```
[[1]]
```

```
[1] AG06 AG06 AG04 AG04
```

Levels: AG04 AG06

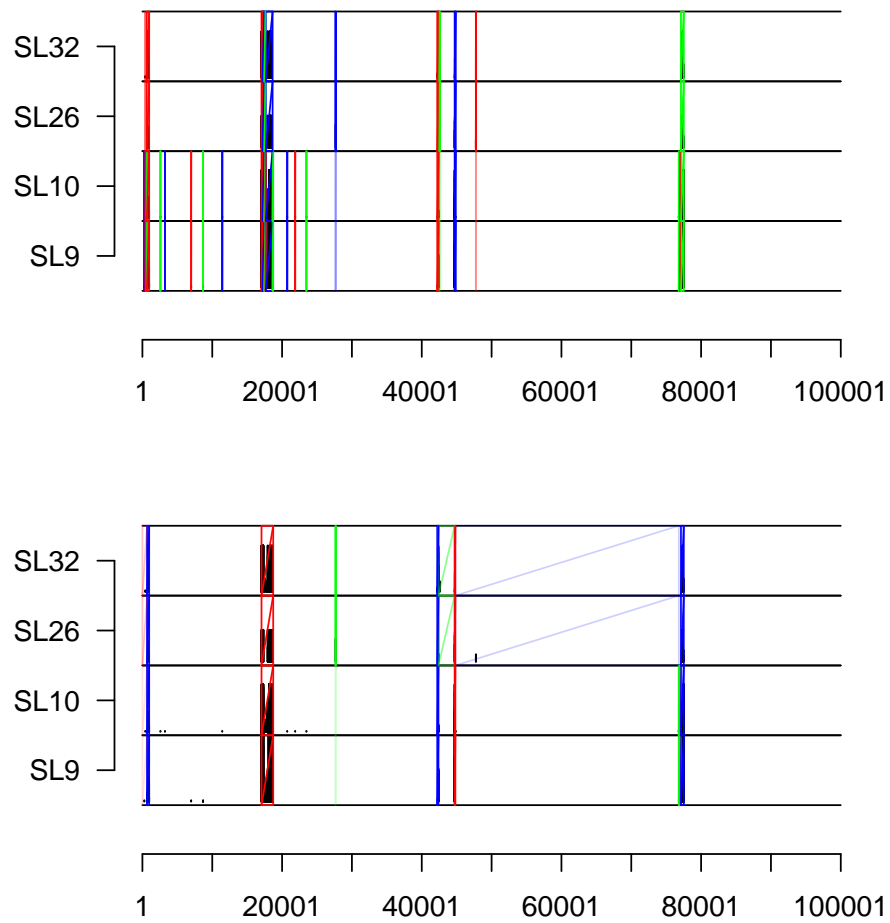


Figure 1: The segmented genome (first 10^5 bases of chromosome 1.

Slot "data":

	AG06.1	AG06.2	AG04.1	AG04.2
[1,]	54	45	65	83
[2,]	682	621	1405	1103
[3,]	31	9	48	56
[4,]	73	57	47	21
[5,]	6	19	0	0

117 more rows...

Slot "annotation":

data frame with 0 columns and 122 rows

Slot "locLikelihoods" (stored on log scale):

Matrix with 122 rows.

	AG04	AG06
1	0.95322	0.97731
2	0.98437	0.99646
3	0.96077	0.95554
4	0.96471	0.99799
5	0.09698	0.97094
...
118	0.8	0.96003

```

119 0.13351 0.96956
120 0.12181 0.98973
121 0.91999 0.98423
122 0.96074 0.97484

```

Expected number of loci in each replicate group

```

      AG04      AG06
81.43231 118.60032

```

The lociData objects can now be examined for differential expression with the baySeq package.

First we define the possible models of differential expression on the data. In this case, the models are of non-differential expression and pairwise differential expression.

```
> groups(classSegs) <- list(NDE = c(1,1,1,1), DE = c("AG06", "AG06", "AG04", "AG04"))
```

Then we get empirical distributions on the parameter space of the data.

```
> classSegs <- getPriors(classSegs, cl = cl)
```

Then we get the posterior likelihoods of the data conforming to each model. Since the 'classSegs' object contains null regions as well as true loci, we will use the 'nullData = TRUE' option to distinguish between non-differentially expressed loci and non-expressed regions. By default, the loci likelihoods calculated earlier will be used to weight the initial parameter fit in order to detect null data.

```
> classSegs <- getLikelihoods(classSegs, nullData = TRUE, cl = cl)
```

.

We can examine the highest likelihood non-expressed ('null') regions

```
> topCounts(classSegs, NULL, number = 3)
```

	seqnames	start	end	width	strand	AG06.1	AG06.2	AG04.1	AG04.2	Likelihood	FDR.
1	>Chr1	754198	758593	4396	*	1	0	1	0	0.9389731	0.06102690
2	>Chr1	950796	958752	7957	*	2	4	3	2	0.9383700	0.06132845
3	>Chr1	148877	222440	73564	*	0	0	0	0	0.9364491	0.06206926

FWER.

```

1 0.0610269
2 0.1188958
3 0.1748908

```

The highest likelihood expressed but non-differentially expressed regions

```
> topCounts(classSegs, "NDE", number = 3)
```

	seqnames	start	end	width	strand	AG06.1	AG06.2	AG04.1	AG04.2	Likelihood	FDR.NDE
1	>Chr2	1554	14423	12870	*	7632	18470	20242	13836	0.9762245	0.02377548
2	>Chr1	446325	447437	1113	*	789	536	1291	1184	0.9323374	0.04571905
3	>Chr1	17055	18728	1674	*	682	621	1405	1103	0.9060522	0.06179530

FWER.NDE

```

1 0.02377548
2 0.08982939
3 0.17533791

```

And the highest likelihood differentially expressed regions

```
> topCounts(classSegs, "DE", number = 3)
```

	seqnames	start	end	width	strand	AG06.1	AG06.2	AG04.1	AG04.2	Likelihood	ordering
1	>Chr2	52652	53314	663	*	84	86	982	696	0.9987839	AG04>AG06
2	>Chr2	58131	59084	954	*	81	83	965	677	0.9985982	AG04>AG06
3	>Chr2	49137	50333	1197	*	101	116	992	719	0.9984145	AG04>AG06

FDR.DE FWER.DE

```

1 0.001216135 0.001216135
2 0.001308949 0.002616193
3 0.001401138 0.004197561

```

Finally, to be a good citizen, we stop the cluster we started earlier:

```
> if(!is.null(cl))
+   stopCluster(cl)
```

Session Info

```
> sessionInfo()
```

```
R version 3.2.1 (2015-06-18)
Platform: x86_64-apple-darwin10.8.0 (64-bit)
Running under: OS X 10.6.8 (Snow Leopard)
```

```
locale:
```

```
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
```

```
attached base packages:
```

```
[1] stats4      parallel    stats      graphics  grDevices  utils      datasets  methods
[9] base
```

```
other attached packages:
```

```
[1] segmentSeq_2.2.2      ShortRead_1.26.0      GenomicAlignments_1.4.1
[4] Rsamtools_1.20.4      Biostrings_2.36.2      XVector_0.8.0
[7] BiocParallel_1.2.19    baySeq_2.2.0           perm_1.0-0.0
[10] abind_1.4-3           GenomicRanges_1.20.5   GenomeInfoDb_1.4.1
[13] IRanges_2.2.5         S4Vectors_0.6.3        BiocGenerics_0.14.0
```

```
loaded via a namespace (and not attached):
```

```
[1] zlibbioc_1.14.0      lattice_0.20-33        hwriter_1.3.2          tools_3.2.1
[5] grid_3.2.1           Biobase_2.28.0         latticeExtra_0.6-26    lambda.r_1.1.7
[9] futile.logger_1.4.1   RColorBrewer_1.1-2     futile.options_1.0.0   bitops_1.0-6
[13] BiocStyle_1.6.0
```

References

- [1] Thomas J. Hardcastle and Krystyna A. Kelly. *baySeq: Empirical Bayesian Methods For Identifying Differential Expression In Sequence Count Data*. BMC Bioinformatics (2010).
- [2] Thomas J. Hardcastle and Krystyna A. Kelly and David C. Baulcombe. *Identifying small RNA loci from high-throughput sequencing data*. Bioinformatics (2012).