

# COSMIC 67

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## 1 Introduction

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The *COSMIC.67* package provides the curated mutations published with the COSMIC release version 67 (2013-10-24). Both variants found in coding and non-coding regions are included and offered as a single object of class 'CollapsedVCF' and a bgzipped and tabix-index 'VCF' file.

Additionally, the package contains the Cancer Gene Census, a list of genes causally linked to cancer.

## 2 Accessing and Using the Data

---

```
library(VariantAnnotation)
```

```
Loading required package: BiocGenerics
```

```
Loading required package: parallel
```

```
Attaching package: 'BiocGenerics'
```

```
The following objects are masked from 'package:parallel':
```

```
clusterApply, clusterApplyLB, clusterCall,  
clusterEvalQ, clusterExport, clusterMap, parApply,  
parCapply, parLapply, parLapplyLB, parRapply,  
parSapply, parSapplyLB
```

```
The following object is masked from 'package:stats':
```

*xtabs*

*The following objects are masked from 'package:base':*

*Filter, Find, Map, Position, Reduce, anyDuplicated,  
append, as.data.frame, as.vector, cbind, colnames,  
do.call, duplicated, eval, evalq, get, intersect,  
is.unsorted, lapply, mapply, match, mget, order,  
paste, pmax, pmax.int, pmin, pmin.int, rank, rbind,  
rep.int, rownames, sapply, setdiff, sort, table,  
tapply, union, unique, unlist*

*Loading required package: GenomicRanges*

*Loading required package: IRanges*

*Loading required package: GenomeInfoDb*

*Loading required package: Rsamtools*

*Loading required package: XVector*

*Loading required package: Biostrings*

*Attaching package: 'VariantAnnotation'*

*The following object is masked from 'package:base':*

*tabulate*

```
library(GenomicRanges)
```

```
data(package = "COSMIC.67")
```

```
data(cosmic_67, package = "COSMIC.67")
```

```
tp53_range = GRanges("17", IRanges(7565097, 7590856))
```

```
vcf_path = system.file("vcf", "cosmic_67.vcf.gz", package = "COSMIC.67")
```

```
cosmic_tp53 = readVcf(vcf_path, genome = "GRCh37", ScanVcfParam(which = tp53_range))
```

```
cosmic_tp53
```

```
class: CollapsedVCF
```

```
dim: 5892 0
```

```
rowData(vcf):
```

```
GRanges with 5 metadata columns: paramRangeID, REF, ALT, QUAL, FILTER
```

```
info(vcf):
```

```
DataFrame with 5 columns: GENE, STRAND, CDS, AA, CNT
```

```
info(header(vcf)):
```

	Number	Type	Description
GENE	1	String	Gene name
STRAND	1	String	Gene strand
CDS	1	String	CDS annotation
AA	1	String	Peptide annotation
CNT	1	Integer	How many samples have this mutation

```
geno(vcf):
```

```
SimpleList of length 0:
```

```
data(cgc_67, package = "COSMIC.67")
```

```
head(cgc_67)
```

	SYMBOL	ENTREZID	ENSEMBL
1	ABI1	10006	ENSG00000136754
2	ABL1	25	ENSG00000097007
3	ABL2	27	ENSG00000143322
4	ACSL3	2181	ENSG00000123983

```
5 CASC5      57082 ENSG00000137812
6 MLLT11     10962 ENSG00000213190
```

For details on the collection and curation of the original data, please see the webpage of the COSMIC project: <http://cancer.sanger.ac.uk/cancergenome/projects/cosmic/>.

## 3 Data Provenance

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### 3.1 COSMIC Mutations

The following steps are performed for importing and processing of the VCF data:

1. Downloading of the VCF files 'CosmicCodingMuts\_v67\_20131024.vcf.gz' and 'CosmicNonCodingVariants\_v67\_20131024.vcf.gz' from 'ftp://ngs.sanger.ac.uk/production/cosmic/' to 'inst/raw/
2. Importing of both files to R using 'readVcf'.
3. Sorting of the seqlevels and adding 'seqinfo' data for the toplevel chromosomes of 'GRCh37'.
4. Merging of both objects, sorting according to genomic position.
5. Converting the object to class `VariantAnnotation::VRanges`.
6. Converting the 'character' columns to 'factors'.
7. Saving the merged object to 'data/cosmic\_v67\_vcf.rda'.
8. Exporting the merged object as a bgzipped and tabix-indexed 'VCF' to 'inst/vcf/cosmic\_v67.vcf.gz'.

### 3.2 Cancer Gene Census

The following steps are performed for importing and processing of the Cancer Gene Census data:

1. Downloading of the 'cancer\_gene\_census.tsv' file from [ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/data\\_export](ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/data_export) to 'inst/raw'.
2. Import of the files as a data frame.
3. Annotation of the 'HGNC' and 'ENSEMBLID' identifiers, using the 'ENTREZ gene ID' as query with the 'org.Hs.eg.db' object.
4. Saving the object to 'data/cgc\_67.rda'.

## 4 Data Source

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The mutation data was obtained from the Sanger Institute Catalogue Of Somatic Mutations In Cancer web site, <http://www.sanger.ac.uk/cosmic>

Bamford et al (2004):

The COSMIC (Catalogue of Somatic Mutations in Cancer) database and website.

Br J Cancer, 91,355-358.

For details on the usage and redistribution of the data, please see [ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/GUIDELINES\\_ON\\_THE\\_USE\\_OF\\_THIS\\_DATA.txt](ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/GUIDELINES_ON_THE_USE_OF_THIS_DATA.txt).

## 5 References

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- <http://cancer.sanger.ac.uk/cancergenome/projects/cosmic/>
- [http://nar.oxfordjournals.org/content/39/suppl\\_1/D945.long](http://nar.oxfordjournals.org/content/39/suppl_1/D945.long)

- [ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/GUIDELINES\\_ON\\_THE\\_USE\\_OF\\_THIS\\_DATA.txt](ftp://ftp.sanger.ac.uk/pub/CGP/cosmic/GUIDELINES_ON_THE_USE_OF_THIS_DATA.txt)

## 6 Session Info

---

```
R version 3.1.0 RC (2014-04-02 r65358)
Platform: i386-w64-mingw32/i386 (32-bit)

locale:
[1] LC_COLLATE=C
[2] LC_CTYPE=English_United States.1252
[3] LC_MONETARY=English_United States.1252
[4] LC_NUMERIC=C
[5] LC_TIME=English_United States.1252

attached base packages:
[1] parallel stats graphics grDevices utils datasets
[7] methods base

other attached packages:
[1] VariantAnnotation_1.10.0 Rsamtools_1.16.0
[3] Biostrings_2.32.0 XVector_0.4.0
[5] GenomicRanges_1.16.0 GenomeInfoDb_1.0.0
[7] IRanges_1.21.45 BiocGenerics_0.10.0
[9] knitr_1.5

loaded via a namespace (and not attached):
[1] AnnotationDbi_1.26.0 BBmisc_1.5
[3] BSgenome_1.32.0 BatchJobs_1.2
[5] Biobase_2.24.0 BiocParallel_0.6.0
[7] BiocStyle_1.2.0 DBI_0.2-7
[9] GenomicAlignments_1.0.0 GenomicFeatures_1.16.0
[11] RCurl_1.95-4.1 RSQLite_0.11.4
[13] Rcpp_0.11.1 XML_3.98-1.1
[15] biomaRt_2.20.0 bitops_1.0-6
[17] brew_1.0-6 codetools_0.2-8
[19] digest_0.6.4 evaluate_0.5.3
[21] fail_1.2 foreach_1.4.2
[23] formatR_0.10 highr_0.3
[25] iterators_1.0.7 plyr_1.8.1
[27] rtracklayer_1.24.0 sendmailR_1.1-2
[29] stats4_3.1.0 stringr_0.6.2
[31] tools_3.1.0 zlibbioc_1.10.0
```