

# Package ‘motifStack’

October 8, 2014

**Type** Package

**Version** 1.8.1

**Date** 2013-6-16

**Title** Plot stacked logos for single or multiple DNA, RNA and amino acid sequence

**Author** Jianhong Ou, Michael Brodsky, Scot Wolfe and Lihua Julie Zhu

**Maintainer** Jianhong Ou <jianhong.ou@umassmed.edu>

**Imports** XML

**Depends** R (>= 2.15.1), methods, grImport, grid, MotIV, ade4

**Suggests** RUnit, BiocGenerics, MotifDb, RColorBrewer, BiocStyle

**biocViews** SequenceMatching, GenomicsSequence, Visualization

**Description** The motifStack package is designed for graphic representation of multiple motifs with different similarity scores. It works with both DNA/RNA sequence motif and amino acid sequence motif. In addition, it provides the flexibility for users to customize the graphic parameters such as the font type and symbol colors.

**License** GPL (>= 2)

**Lazyload** yes

## R topics documented:

motifStack-package . . . . .	2
colorset . . . . .	2
DNAmotifAlignment . . . . .	3
motifCloud . . . . .	4
motifSig-class . . . . .	5
motifSig-methods . . . . .	6
motifSignature . . . . .	6

motifStack . . . . .	7
ouNode-class . . . . .	8
pcm-class . . . . .	9
pcm-methods . . . . .	10
pfm-class . . . . .	11
pfm-methods . . . . .	12
plotMotifLogo . . . . .	13
plotMotifLogoA . . . . .	14
plotMotifLogoStack . . . . .	15
plotMotifLogoStackWithTree . . . . .	16
plotMotifStackWithPhylog . . . . .	17
plotMotifStackWithRadialPhylog . . . . .	18
plotXaxis . . . . .	20
plotYaxis . . . . .	21
readPCM . . . . .	21

**Index****22**

motifStack-package	<i>Plot stacked logos for single or multiple DNA, RNA and amino acid sequence</i>
--------------------	---

**Description**

motifStack is a package that is able to draw amino acid sequence as easy as to draw DNA/RNA sequence. motifStack provides the flexibility for users to select the font type and symbol colors. motifStack is designed for graphical representation of multiple motifs.

**Author(s)**

Jianhong Ou and Lihua Julie Zhu

Maintainer: Jianhong Ou <jianhong.ou@umassmed.edu>

colorset	<i>retrieve color setting for logo</i>
----------	--

**Description**

retrieve color setting for logo

**Usage**

```
colorset(alphabet="DNA", colorScheme=auto)
```

**Arguments**

alphabet	character, 'DNA', 'RNA' or 'AA'
colorScheme	'auto', 'charge', 'chemistry', 'classic' or 'hydrophobicity' for AA, 'auto' or 'basepairing' for DNA ro RNA

**Value**

A character vector of color scheme

**Examples**

```
col <- colorset("AA", "hydrophobicity")
```

DNAmotifAlignment	<i>align DNA motifs</i>
-------------------	-------------------------

**Description**

align DNA motifs for plotting motifs stack

**Usage**

```
DNAmotifAlignment(pfms, threshold=0.4, minimalConsensus=0, rcprefix="(RC)", revcomp=rep(TRUE, length
```

**Arguments**

pfms	a list of position frequency matrices, pfms must be a list of class pfm
threshold	information content cutoff threshold for useful postions
minimalConsensus	minimal length of consensus for alignment
rcprefix	the postfix for reverse complements
revcomp	a logical vector to indicates whether the reverse complemet should be involved into alignment

**Value**

a list of aligned motifs

**Examples**

```
pcms<-readPCM(file.path(find.package("motifStack"), "extdata"),"pcm$")
motifs<-lapply(pcms,pcm2pfm)
motifs<-DNAmotifAlignment(motifs)
```

**motifCloud***plot a DNA sequence logo cloud*

## Description

Plot a DNA sequence logo cloud

## Usage

```
motifCloud(motifSig, rcprefix=(RC),
           layout=c("rectangles", "cloud", "tree"),
           scale=c(6, .5), rot.per=.1,
           draw.box=TRUE, draw.freq=TRUE,
           box.col="gray", freq.col="gray",
           group.col=NULL, groups=NULL, draw.legend=FALSE,
           ic.scale=TRUE)
```

## Arguments

<b>motifSig</b>	an object of class <a href="#">motifSig</a>
<b>rcprefix</b>	postfix for reverse-complement motif names, default: (RC)
<b>layout</b>	layout of the logo cloud, rectangles, cloud or tree
<b>scale</b>	A vector of length 2 indicating the range of the size of the sequence logo.
<b>rot.per</b>	proportion sequence logo with 90 degree rotation. Only work for "cloud" layout
<b>draw.box</b>	draw box for each sequence logo or not
<b>draw.freq</b>	label frequency of each signature or not
<b>box.col</b>	color of box for each sequence logo
<b>freq.col</b>	color of frequency label
<b>group.col</b>	color setting for groups
<b>groups</b>	a named vectors of motif groups
<b>draw.legend</b>	draw group color legend or not
<b>ic.scale</b>	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

## Value

none

## Examples

```

if(interactive()){
library("MotifDb")
matrix.fly <- query(MotifDb, "Dmelanogaster")
motifs <- as.list(matrix.fly)
motifs <- motifs[grep("Dmelanogaster-FlyFactorSurvey-", names(motifs), fixed=TRUE)]
names(motifs) <- gsub("Dmelanogaster_FlyFactorSurvey_", "", 
  gsub("_FBgn[0-9]+$", "", 
  gsub("[^a-zA-Z0-9]", "_",
  gsub("(_[0-9]+)$", "", names(motifs)))))}
motifs <- motifs[unique(names(motifs))]
pfms <- sample(motifs, 50)
jaspar.scores <- MotIV::readDBScores(file.path(find.package("MotIV"), "extdata", "jaspar2010_PCC_SWU.scores"))
d <- MotIV::motifDistances(pfms)
hc <- MotIV::motifHclust(d)
phylog <- hclust2phylog(hc)
leaves <- names(phylog$leaves)
pfms <- pfms[leaves]
pfms <- lapply(names(pfms), function(.ele, pfms){new("pfm", mat=pfms[[.ele]], name=.ele)}, pfms)
motifSig <- motifSignature(pfms, phylog, groupDistance=0.1)
motifCloud(motifSig)
}

```

**motifSig-class**      *Class "motifSig"*

## Description

An object of class "motifSig" represents the output of function [motifSignature](#)

## Objects from the Class

Objects can be created by calls of the form `new("motifSig", signature, freq, nodelist, gpcol)`.

## Slots

`signatures` list object of class "pfm"  
`freq` code"numeric" signature frequency  
`nodelist` list object of class "[ouNode](#)"  
`gpcol` code"character" signature group color sets

## Methods

**signatures** `signature(object = "motifSig")` return the signatures of motifSig  
**frequency** `signature(object = "motifSig")` return the frequency of motifSig  
**nodelist** `signature(object = "motifSig")` return the nodelist of motifSig  
**sigColor** `signature(object = "motifSig")` return the group color sets of motifSig

**motifSig-methods**      *"motifSig" methods*

## Description

methods for motifSig objects.

## Usage

```
## S4 method for signature motifSig
signatures(object)
## S4 method for signature motifSig
frequence(object)
## S4 method for signature motifSig
nodelist(object)
## S4 method for signature motifSig
sigColor(object)
```

## Arguments

**object**      An object of class motifSig.

## Methods

**signatures** `signature(object = "motifSig")` return the signatures of motifSig  
**frequence** `signature(object = "motifSig")` return the frequency of motifSig  
**nodelist** `signature(object = "motifSig")` return the nodelist of motifSig  
**sigColor** `signature(object = "motifSig")` return the group color sets of motifSig

**motifSignature**      *get signatures from motifs*

## Description

extract signatures from multiple motifs by distance calculated from STAMP

## Usage

```
motifSignature(pfms, phylog, groupDistance, rcprefix="(RC)",
min.freq=2, trim=0.2, families=list())
```

**Arguments**

pfms	a list of objects of class pfm
phylog	an object of class phylog
groupDistance	maximal distance of motifs in the same group
rcompostfix	postfix for reverse-complement motif names, default: (RC)
min.freq	signatures with frequency below min.freq will not be plotted
trim	minimal information content for each position of signature
families	for each family, the motif number in one signature should only count as 1

**Value**

an Object of class [motifSig](#)

**Examples**

```
if(interactive()){
library("MotifDb")
matrix.fly <- query(MotifDb, "Dmelanogaster")
motifs <- as.list(matrix.fly)
motifs <- motifs[grepl("Dmelanogaster-FlyFactorSurvey-", names(motifs), fixed=TRUE)]
names(motifs) <- gsub("Dmelanogaster_FlyFactorSurvey_", "", 
  gsub("_FBgn[0-9]+$", "", 
  gsub("[^a-zA-Z0-9]", "_",
  gsub("(_[0-9]+)$", "", names(motifs)))))}
motifs <- motifs[unique(names(motifs))]
pfms <- sample(motifs, 50)
jaspar.scores <- MotIV::readDBScores(file.path(find.package("MotIV"), "extdata", "jaspar2010_PCC_SWU.scores"))
d <- MotIV::motifDistances(pfms)
hc <- MotIV::motifHclust(d)
phylog <- hclust2phylog(hc)
leaves <- names(phylog$leaves)
pfms <- pfms[leaves]
pfms <- lapply(names(pfms), function(.ele, pfms){new("pfm", mat=pfms[[.ele]], name=.ele)}, pfms)
motifSig <- motifSignature(pfms, phylog, groupDistance=0.1)
}
```

**motifStack**

*plot a DNA sequence logo stack*

**Description**

Plot a DNA sequence logo stack

**Usage**

```
motifStack(pfms, layout=c("stack", "treeview", "phylog", "radialPhylog"), ...)
```

## Arguments

pfms	a list of objects of class <a href="#">pfm</a>
layout	layout of the logo stack, stack, treeview or radialPhylog
...	any parameters could to pass to <a href="#">plotMotifLogoStack</a> , <a href="#">plotMotifLogoStackWithTree</a> , <a href="#">plotMotifStackWithPhylog</a> or <a href="#">plotMotifStackWithRadialPhylog</a>

## Value

return a list contains pfms and phylog

## Examples

```
if(interactive()){
library("MotifDb")
matrix.fly <- query(MotifDb, "Dmelanogaster")
motifs <- as.list(matrix.fly)
motifs <- motifs[grep("Dmelanogaster-FlyFactorSurvey-", names(motifs), fixed=TRUE)]
names(motifs) <- gsub("Dmelanogaster_FlyFactorSurvey_", "", 
  gsub("_FBgn[0-9]+$", "", 
  gsub("[^a-zA-Z0-9]", "_",
  gsub("(_[0-9]+)+$", "", names(motifs)))))}
motifs <- motifs[unique(names(motifs))]
pfms <- sample(motifs, 50)
pfms <- lapply(names(pfms), function(.ele, pfms){new("pfm", mat=pfms[[.ele]], name=.ele)}, pfms)
motifStack(pfms, "radialPhylog")
}
```

ouNode-class

*Class ouNode*

## Description

An object of class "ouNode" represents a motif node in a cluster tree

## Objects from the Class

Objects can be created by calls of the form `new("ouNode", left, right, parent, distl, distr, sizel, sizer)`.

## Slots

- left:** character indicates the name of left leave
- right:** character indicates the name of right leave
- parent:** character indicates the name of parent node
- distl:** numeric indicates the distance of left leave
- distr:** numeric indicates the distance of right leave
- sizel:** numeric indicates the size of left leave
- sizer:** numeric indicates the size of right leave

## Examples

```
new("ouNode", left="A", right="B", parent="Root", distl=1, distr=2, sizel=1, sizer=1)
```

---

pcm-class

*Class "pcm"*

---

## Description

An object of class "pcm" represents the position count matrix of a DNA/RNA/amino-acid sequence motif. The entry stores a matrix, which in row i, column j gives the counts of observing nucleotide/or amino acid i in position j of the motif.

## Objects from the Class

Objects can be created by calls of the form `new("pcm", mat, name, alphabet, color, background)`.

## Slots

- `mat` Object of class "matrix" The position count matrix
- `name` code"character" The motif name
- `alphabet` "character" The sequence alphabet. "DNA", "RNA", "AA" or "others".
- `color` a "character" vector. The color setting for each symbol
- `background` a "numeric" vector. The background frequency.

## Methods

- addBlank** signature(x="pcm", n="numeric", b="logical") add space into the position count matrix for alignment. b is a bool value, if TRUE, add space to the 3' end, else add space to the 5' end. n indicates how many spaces should be added.
- coerce** signature(from = "pcm", to = "matrix"): convert object pcm to matrix
- getIC** signature(x = "pcm",) Calculate information content profile for position frequency matrix.
- matrixReverseComplement** signature(x = "pcm") get the reverse complement of position frequency matrix.
- trimMotif** signature(x = "pfm", t= "numeric") trim motif by information content.
- plot** signature(x = "pcm") Plots the sequence logo of the position count matrix.

## Examples

```
pcm <- read.table(file.path(find.package("motifStack"), "extdata", "bin_SOLEXA.pcm"))
pcm <- pcm[,3:ncol(pcm)]
rownames(pcm) <- c("A","C","G","T")
motif <- new("pcm", mat=as.matrix(pcm), name="bin_SOLEXA")
plot(motif)
```

---

pcm-methods	<i>"pcm" methods</i>
-------------	----------------------

---

## Description

methods for pcm objects.

## Usage

```
## S4 method for signature pcm,numeric,logical
addBlank(x,n,b)
## S4 method for signature pcm,ANY
getIC(x,p="missing")
## S4 method for signature pcm
matrixReverseComplement(x)
## S4 method for signature pcm,ANY
plot(x,y="missing",...)
## S4 method for signature pcm,ANY
pcm2pfm(x,background="missing")
## S4 method for signature matrix,ANY
pcm2pfm(x,background="missing")
## S4 method for signature matrix,numeric
pcm2pfm(x,background)
## S4 method for signature data.frame,ANY
pcm2pfm(x,background="missing")
## S4 method for signature data.frame,numeric
pcm2pfm(x,background)
## S4 method for signature pcm,numeric
trimMotif(x,t)
```

## Arguments

- x An object of class pfm. For getIC, if parameter p is followed, x should be an object of matrix. For pcm2pfm, x also could be an object of matrix.
- y Not use.
- p p is the background frequency.
- n how many spaces should be added.
- b logical value to indicate where the space should be added.
- background a "numeric" vector. The background frequency.
- t numeric value of information content threshold for trimming.
- ... Further potential arguments passed to plotMotifLogo.

## Methods

**addBlank** signature(x="pcm", n="numeric", b="logical") add space into the position count matrix for alignment. b is a bool value, if TRUE, add space to the 3' end, else add space to the 5' end. n indicates how many spaces should be added.

**coerce** signature(from = "pcm", to = "matrix"): convert object pcm to matrix

**getIC** signature(x = "pcm",) Calculate information content profile for position frequency matrix.

**matrixReverseComplement** signature(x = "pcm") get the reverse complement of position frequency matrix.

**plot** signature(x = "pcm") Plots the sequence logo of the position count matrix.

**trimMotif** signature(x = "pfm", t= "numeric") trim motif by information content.

## Examples

```
pcm <- read.table(file.path(find.package("motifStack"), "extdata", "bin_SOLEXA.pcm"))
pcm <- pcm[,3:ncol(pcm)]
rownames(pcm) <- c("A", "C", "G", "T")
motif <- new("pcm", mat=as.matrix(pcm), name="bin_SOLEXA")
getIC(motif)
matrixReverseComplement(motif)
as(motif,"matrix")
pcm2pfm(motif)
```

pfm-class

Class "pfm"

## Description

An object of class "pfm" represents the position frequency matrix of a DNA/RNA/amino-acid sequence motif. The entry stores a matrix, which in row i, column j gives the frequency of observing nucleotide/or amino acid i in position j of the motif.

## Objects from the Class

Objects can be created by calls of the form new("pfm", mat, name, alphabet, color, background).

## Slots

**mat** Object of class "matrix" The position frequency matrix

**name** code"character" The motif name

**alphabet** "character" The sequence alphabet. "DNA", "RNA", "AA" or "others".

**color** a "character" vector. The color setting for each symbol

**background** a "numeric" vector. The background frequency.

## Methods

**addBlank** signature(x="pfm", n="numeric", b="logical") add space into the position frequency matrix for alignment. b is a bool value, if TRUE, add space to the 3' end, else add space to the 5' end. n indicates how many spaces should be added.

**coerce** signature(from = "pfm", to = "matrix"): convert object pfm to matrix

**getIC** signature(x = "pfm",) Calculate information content profile for position frequency matrix.

**getIC** signature(x = "matrix", p = "numeric") Calculate information content profile for matrix. p is the background frequency

**matrixReverseComplement** signature(x = "pfm") get the reverse complement of position frequency matrix.

**trimMotif** signature(x = "pfm", t= "numeric") trim motif by information content.

**plot** signature(x = "pfm") Plots the sequence logo of the position frequency matrix.

## Examples

```
pcm <- read.table(file.path(find.package("motifStack"), "extdata", "bin_SOLEXA.pcm"))
pcm <- pcm[,3:ncol(pcm)]
rownames(pcm) <- c("A","C","G","T")
motif <- pcm2pfm(pcm)
motif <- new("pfm", mat=motif, name="bin_SOLEXA")
plot(motif)
```

## Description

methods for pfm objects.

## Usage

```
## S4 method for signature pfm,numeric,logical
addBlank(x,n,b)
## S4 method for signature pfm,ANY
getIC(x,p="missing")
## S4 method for signature matrix,numeric
getIC(x,p)
## S4 method for signature pfm
matrixReverseComplement(x)
## S4 method for signature pfm,ANY
plot(x,y="missing",...)
## S4 method for signature pfm,numeric
trimMotif(x,t)
```

## Arguments

x	An object of class pfm. For getIC, if parameter p is followed, x should be an object of matrix.
y	Not use.
p	p is the background frequency.
n	how many spaces should be added.
b	logical value to indicate where the space should be added.
t	numeric value of information content threshold for trimming.
...	Further potential arguments passed to plotMotifLogo.

## Methods

**addBlank** signature(x="pfm", n="numeric", b="logical") add space into the position frequency matrix for alignment. b is a bool value, if TRUE, add space to the 3' end, else add space to the 5' end. n indicates how many spaces should be added.

**getIC** signature(x = "pfm",) Calculate information content profile for position frequency matrix.

**getIC** signature(x = "matrix", p = "numeric") Calculate information content profile for matrix. p is the background frequency

**matrixReverseComplement** signature(x = "pfm") get the reverse complement of position frequency matrix.

**plot** signature(x = "pfm") Plots the sequence logo of the position frequency matrix.

**trimMotif** signature(x = "pfm", t= "numeric") trim motif by information content.

## Examples

```
pcm <- read.table(file.path(find.package("motifStack"), "extdata", "bin_SOLEXA.pcm"))
pcm <- pcm[,3:ncol(pcm)]
rownames(pcm) <- c("A", "C", "G", "T")
motif <- pcm2pfm(pcm)
motif <- new("pfm", mat=motif, name="bin_SOLEXA")
getIC(motif)
matrixReverseComplement(motif)
addBlank(motif, 1, FALSE)
addBlank(motif, 3, TRUE)
as(motif, "matrix")
```

plotMotifLogo

*plot sequence logo*

## Description

plot amino acid or DNA sequence logo

**Usage**

```
plotMotifLogo(pf $\text{m}$ , motifName, p=rep(0.25, 4), font="Helvetica-Bold",
colset=c("#00811B", "#2000C7", "#FFB32C", "#D00001"),
xaxis=TRUE, yaxis=TRUE, xlab="position", ylab="bits",
xlcex=1.2, ylcex=1.2, ncex=1.2, ic.scale=TRUE)
```

**Arguments**

pf $\text{m}$	a position frequency matrices
motifName	motif name
p	background possibility
font	font of logo
colset	color setting for each logo letter
xaxis	draw x-axis or not
yaxis	draw y-axis or not
xlab	x-label, do nothing if set xlab as NA
ylab	y-label, do nothing if set ylab as NA
xlce $\text{x}$	cex value for x-label
ylce $\text{x}$	cex value for y-label
ncex	cex value for motif name
ic.scale	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

**Value**

none

**Examples**

```
pcm<-matrix(runif(40,0,100),nrow=4,ncol=10)
pf $\text{m}$ <-pcm2pf $\text{m}$ (pcm)
rownames(pf $\text{m}$ )<-c("A", "C", "G", "T")
plotMotifLogo(pf $\text{m}$ )
```

**plotMotifLogoA**

*plot sequence logo without plot.new*

**Description**

plot amino acid or DNA sequence logo in a given canvas

**Usage**

```
plotMotifLogoA(pf $\text{m}$ , font="Helvetica-Bold", ic.scale=TRUE)
```

**Arguments**

pfm	an object of pfm
font	font of logo
ic.scale	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

**Value**

none

**Examples**

```
pcm<-matrix(runif(40,0,100),nrow=4,ncol=10)
pfm<-pcm2pfm(pcm)
rownames(pfm)<-c("A","C","G","T")
motif <- new("pfm", mat=pfm, name="bin_SOLEXA")
plotMotifLogoA(motif)
```

plotMotifLogoStack      *plot sequence logos stack*

**Description**

plot sequence logos stack

**Usage**

plotMotifLogoStack(pfms, ...)

**Arguments**

pfms	a list of position frequency matrices, pfms must be a list of class pfm
...	other parameters can be passed to plotMotifLogo function

**Value**

none

**Examples**

```
pcm1<-matrix(c(0,50,0,50,
 100,0,0,0,
 0,100,0,0,
 0,0,100,0,
 0,0,0,100,
 50,50,0,0,
 0,0,50,50), nrow=4)
pcm2<-matrix(c(50,50,0,0,
```

```

0,100,0,0,
0,50,50,0,
0,0,0,100,
50,50,0,0,
0,0,50,50), nrow=4)
rownames(pcm1)<-c("A", "C", "G", "T")
rownames(pcm2)<-c("A", "C", "G", "T")
pfms<-list(p1=new("pfm", mat=pcm2pfm(pcm1), name="m1"),
            p2=new("pfm", mat=pcm2pfm(pcm2), name="m2"))
pfms<-DNAmotifAlignment(pfms)
plotMotifLogoStack(pfms)

```

---

**plotMotifLogoStackWithTree***plot sequence logos stack with hierarchical cluster tree***Description**

plot sequence logos stack with hierarchical cluster tree

**Usage**`plotMotifLogoStackWithTree(pfms, hc, treewidth=1/8, trueDist=FALSE, ...)`**Arguments**

<code>pfms</code>	a list of position frequency matrices, pfms must be a list of class pfm
<code>hc</code>	an object of the type produced by stats::hclust
<code>treewidth</code>	the width to show tree
<code>trueDist</code>	logical flags to use hclust height or not.
<code>...</code>	other parameters can be passed to plotMotifLogo function

**Value**

none

**Examples**

```

#####Input#####
pcms<-readPCM(file.path(find.package("motifStack"), "extdata"), "pcm$")
motifs<-lapply(pcms, pcm2pfm)

#####Clustering#####
jaspar.scores <- MotIV::readDBScores(file.path(find.package("MotIV"), "extdata", "jaspar2010_PCC_SWU.scores"))
d <- MotIV::motifDistances(sapply(motifs, function(.ele) .ele@mat))
hc <- MotIV::motifHclust(d)

##reorder the motifs for plotMotifLogoStack

```

```

motifs<-motifs[hc$order]
##do alignment
motifs<-DNAmotifAlignment(motifs)
##plot stacks
plotMotifLogoStack(motifs, nce=1.0)
plotMotifLogoStackWithTree(motifs, hc=hc)

```

**plotMotifStackWithPhylog***plot sequence logo stacks with a ape4-style phylogenetic tree***Description**

plot sequence logo stacks with a ape4-style phylogenetic tree

**Usage**

```

plotMotifStackWithPhylog(phylog, pfms=NULL,
f.phylog = 0.3, f.logo = NULL, cleaves =1, cnodes =0,
labels.leaves = names(phylog$leaves), clabel.leaves=1,
labels.nodes = names(phylog$nodes), clabel.nodes = 0, ic.scale=TRUE)

```

**Arguments**

<code>phylog</code>	an object of class <code>phylog</code>
<code>pfms</code>	a list of objects of class <code>pfm</code>
<code>f.phylog</code>	a size coefficient for tree size (a parameter to draw the tree in proportion to leaves label)
<code>f.logo</code>	a size coefficient for the motif
<code>cleaves</code>	a character size for plotting the points that represent the leaves, used with <code>par("cex")*cleaves</code> . If zero, no points are drawn
<code>cnodes</code>	a character size for plotting the points that represent the nodes, used with <code>par("cex")*cnodes</code> . If zero, no points are drawn
<code>labels.leaves</code>	a vector of strings of characters for the leaves labels
<code>clabel.leaves</code>	a character size for the leaves labels, used with
<code>labels.nodes</code>	a vector of strings of characters for the nodes labels
<code>clabel.nodes</code>	a character size for the nodes labels, used with <code>par("cex")*clabel.nodes</code> . If zero, no nodes labels are drawn
<code>ic.scale</code>	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

**Value**

`none`

**See Also**[plot.phylog](#)**Examples**

```
if(interactive()){
library("MotifDb")
matrix.fly <- query(MotifDb, "Dmelanogaster")
motifs <- as.list(matrix.fly)
motifs <- motifs[grepl("Dmelanogaster-FlyFactorSurvey-", names(motifs), fixed=TRUE)]
names(motifs) <- gsub("Dmelanogaster_FlyFactorSurvey_", "", 
  gsub("_FBgn[0-9]+$", "", 
  gsub("[^a-zA-Z0-9]", "_",
  gsub("(_[0-9]+)+$", "", names(motifs)))))}
motifs <- motifs[unique(names(motifs))]
pfms <- sample(motifs, 50)
jaspar.scores <- MotIV::readDBScores(file.path(find.package("MotIV"), "extdata", "jaspar2010_PCC_SWU.scores"))
d <- MotIV::motifDistances(pfms)
hc <- MotIV::motifHclust(d)
phylog <- hclust2phylog(hc)
leaves <- names(phylog$leaves)
pfms <- pfms[leaves]
pfms <- lapply(names(pfms), function(.ele, pfms){new("pfm", mat=pfms[[.ele]], name=.ele)}, pfms)
pfms <- DNAmotifAlignment(pfms, minimalConsensus=3)
plotMotifStackWithPhylog(phylog, pfms, f.phylog=0.3, cleaves = 0.5, clabel.leaves = 0.7)
}
```

**plotMotifStackWithRadialPhylog***plot sequence logo stacks with a radial phylogenetic tree***Description**

plot sequence logo stacks with a radial phylogenetic tree

**Usage**

```
plotMotifStackWithRadialPhylog(phylog, pfms=NULL,
  circle=1, circle.motif=NA, cleaves=1, cnodes=0,
  labels.leaves=names(phylog$leaves), clabel.leaves=1,
  labels.nodes=names(phylog$nodes), clabel.nodes=0,
  draw.box=FALSE,
  col.leaves=rep("black", length(labels.leaves)),
  col.leaves.bg=NULL, col.leaves.bg.alpha=1,
  col.bg=NULL, col.bg.alpha=1,
  col.inner.label.circle=NULL, inner.label.circle.width="default",
  col.outer.label.circle=NULL, outer.label.circle.width="default",
  clockwise =FALSE, init.angle=if(clockwise) 90 else 0,
```

```
angle=360, pfmNameSpliter=";", rcpostfix = "(RC)", motifScale=c("linear","logarithmic"), ic.scale=TRUE)
```

### Arguments

<code>phylog</code>	an object of class phylog
<code>pfms</code>	a list of objects of class pfm
<code>circle</code>	a size coefficient for the outer circle
<code>circle.motif</code>	a size coefficient for the motif circle
<code>cleaves</code>	a character size for plotting the points that represent the leaves, used with <code>par("cex")*cleaves</code> . If zero, no points are drawn
<code>cnodes</code>	a character size for plotting the points that represent the nodes, used with <code>par("cex")*cnodes</code> . If zero, no points are drawn
<code>labels.leaves</code>	a vector of strings of characters for the leaves labels
<code>clabel.leaves</code>	a character size for the leaves labels, used with
<code>labels.nodes</code>	a vector of strings of characters for the nodes labels
<code>clabel.nodes</code>	a character size for the nodes labels, used with <code>par("cex")*clabel.nodes</code> . If zero, no nodes labels are drawn
<code>draw.box</code>	if TRUE draws a box around the current plot with the function <code>box()</code>
<code>col.leaves</code>	a vector of colors for leaves labels
<code>col.leaves.bg</code>	a vector of colors for background of leaves labels
<code>col.leaves.bg.alpha</code>	alpha value [0, 1] for the colors of background of leaves labels
<code>col.bg</code>	a vector of colors for background
<code>col.bg.alpha</code>	a alpha value [0, 1] of colors for background
<code>col.inner.label.circle</code>	a vector of colors for inner circle of leaves labels
<code>inner.label.circle.width</code>	width for inner circle of leaves labels
<code>col.outer.label.circle</code>	a vector of colors for outer circle of leaves labels
<code>outer.label.circle.width</code>	width for outer circle of leaves labels
<code>clockwise</code>	a logical value indicating if slices are drawn clockwise or counter clockwise
<code>init.angle</code>	number specifying the starting angle (in degrees) for the slices. Defaults to 0 (i.e., '3 o'clock') unless clockwise is true where init.angle defaults to 90 (degrees), (i.e., '12 o'clock')
<code>angle</code>	number specifying the angle (in degrees) for phylogenetic tree. Defaults 360
<code>pfmNameSpliter</code>	splitter when name of pfms contain multiple node of labels.leaves
<code>rcpostfix</code>	the postfix for reverse complements
<code>motifScale</code>	the scale of logo size
<code>ic.scale</code>	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

**Value**

none

**See Also**

[plot.phylog](#)

**Examples**

```
if(interactive()){
  library("MotifDb")
  matrix.fly <- query(MotifDb, "Dmelanogaster")
  motifs <- as.list(matrix.fly)
  motifs <- motifs[grep("Dmelanogaster-FlyFactorSurvey-", names(motifs), fixed=TRUE)]
  names(motifs) <- gsub("Dmelanogaster_FlyFactorSurvey_", "", 
    gsub("_FBgn[0-9]+$", "", 
    gsub("[^a-zA-Z0-9]", "_",
    gsub("(_[0-9]+)+$", "", names(motifs)))))}
  motifs <- motifs[unique(names(motifs))]
  pfms <- sample(motifs, 50)
  jaspar.scores <- MotIV::readDBScores(file.path(find.package("MotIV"), "extdata", "jaspar2010_PCC_SWU.scores"))
  d <- MotIV::motifDistances(pfms)
  hc <- MotIV::motifHclust(d)
  phylog <- hclust2phylog(hc)
  leaves <- names(phylog$leaves)
  pfms <- pfms[leaves]
  pfms <- lapply(names(pfms), function(.ele, pfms){new("pfm", mat=pfms[[.ele]], name=.ele)}, pfms)
  pfms <- DNAmotifAlignment(pfms, minimalConsensus=3)
  library(RColorBrewer)
  color <- brewer.pal(12, "Set3")
  plotMotifStackWithRadialPhylog(phylog, pfms, circle=0.9, cleaves = 0.5, clabel.leaves = 0.7,
    col.bg=rep(color, each=5), col.leaves=rep(color, each=5))
}
```

**plotXaxis**

*plot x-axis*

**Description**

plot x-axis for the sequence logo

**Usage**

```
plotXaxis(pfm, p=rep(0.25, 4))
```

**Arguments**

pfm	position frequency matrices
p	background possibility

**Value**

none

---

plotYaxis

*plot y-axis*

---

**Description**

plot y-axis for the sequence logo

**Usage**

```
plotYaxis(pfM, ic.scale=TRUE)
```

**Arguments**

pfM	position frequency matrices
ic.scale	logical If TRUE, the height of each column is proportional to its information content. Otherwise, all columns have the same height.

**Value**

none

---

readPCM

*read pcm from a path*

---

**Description**

read position count matrix from a path

**Usage**

```
readPCM(path=". ", pattern=NULL)
```

**Arguments**

path	a character vector of full path names
pattern	an optional regular expression

**Value**

A list of [pcm](#) objects

**Examples**

```
pcms<-readPCM(file.path(find.package("motifStack"), "extdata"),"pcm$")
```

# Index

## \*Topic classes

motifSig-class, 5  
motifSig-methods, 6  
ouNode-class, 8  
pcm-class, 9  
pcm-methods, 10  
pfm-class, 11  
pfm-methods, 12

## \*Topic package

motifStack-package, 2  
  
addBlank (pfm-methods), 12  
addBlank, pcm, numeric, logical-method  
    (pfcm-methods), 10  
addBlank, pfm, numeric, logical-method  
    (pfpm-methods), 12  
  
colorset, 2  
  
DNAmotifAlignment, 3  
  
frequence (motifSig-methods), 6  
frequence, motifSig-method  
    (motifSig-methods), 6  
  
getIC (pfm-methods), 12  
getIC, matrix, numeric-method  
    (pfpm-methods), 12  
getIC, pcm, ANY-method (pcm-methods), 10  
getIC, pfm, ANY-method (pfm-methods), 12  
  
matrixReverseComplement (pfm-methods),  
    12  
matrixReverseComplement, pcm-method  
    (pfcm-methods), 10  
matrixReverseComplement, pfm-method  
    (pfpm-methods), 12  
motifCloud, 4  
motifSig, 4, 7  
motifSig (motifSig-methods), 6  
motifSig-class, 5  
  
motifSig-methods, 6  
motifSignature, 5, 6  
motifStack, 7  
motifStack-package, 2  
  
nodelist (motifSig-methods), 6  
nodelist, motifSig-method  
    (motifSig-methods), 6  
  
ouNode, 5  
ouNode (ouNode-class), 8  
ouNode-class, 8  
  
pcm, 21  
pcm (pcm-methods), 10  
pcm-class, 9  
pcm-methods, 10  
pcm2pfm (pcm-methods), 10  
pcm2pfm, data.frame, ANY-method  
    (pfcm-methods), 10  
pcm2pfm, data.frame, numeric-method  
    (pfcm-methods), 10  
pcm2pfm, matrix, ANY-method  
    (pfcm-methods), 10  
pcm2pfm, matrix, numeric-method  
    (pfpm-methods), 10  
pcm2pfm, pcm, ANY-method (pcm-methods), 10  
pfm, 8  
pfm (pfm-methods), 12  
pfm-class, 11  
pfm-methods, 12  
plot, pcm, ANY-method (pcm-methods), 10  
plot, pfm, ANY-method (pfm-methods), 12  
plot.phylog, 18, 20  
plotMotifLogo, 13  
plotMotifLogoA, 14  
plotMotifLogoStack, 8, 15  
plotMotifLogoStackWithTree, 8, 16  
plotMotifStackWithPhylog, 8, 17  
plotMotifStackWithRadialPhylog, 8, 18

plotXaxis, 20  
plotYaxis, 21  
  
readPCM, 21  
  
sigColor (motifSig-methods), 6  
sigColor, motifSig-method  
    (motifSig-methods), 6  
signatures (motifSig-methods), 6  
signatures, motifSig-method  
    (motifSig-methods), 6  
  
trimMotif (pcm-methods), 10  
trimMotif, pcm, numeric-method  
    (pcm-methods), 10  
trimMotif, pfm, numeric-method  
    (pfm-methods), 12