

The OmicCircos usages by examples

Ying Hu and Chunhua Yan

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1 Introduction

The OmicCircos is to generate high-quality circular plots for visualizing variations in genomic data. The data can be gene or chromosome position-based values for mutation, copy number variation, expression, and methylation. OmicCircos is capable of displaying variations in scatterplot, line, and text label. The relationships between genomic features can be presented in polygon and curve. By utilizing the statistical and graphic functions in R/Bioconductor environment, OmicCircos is also able to draw boxplot, histogram, and heatmap from multiple sample data.

In this vignette, we will introduce the package plotting functions using simulation data sets and TCGA gene expression and copy number variation (cnv) data sets (<http://www.cancergenome.nih.gov/>). A quick way to load the vignette examples is:

```
1 vignette("OmicCircos")
```

2 Input file formats

There are four input data files in the package: segment data, mapping data, linking data and linking polygon data.

2.1 segment data

The first input file `segment` data lays out the foundation for a circular graph. Column 1 should be the segment or chromosome name. Columns 2 and 3 are the start and end points of the segment. Columns 4 and 5 are optional and used as segment or other segment name and description. The package comes with the segment data for human (hg18 and hg19) or mouse (mm9 and mm10). Let's start by loading the package

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3 # load the hg18 segment data
4 data(UCSC.hg18.chr);
5 # display the first six rows of the data
6 head(UCSC.hg18.chr);
```

```
## Loading required package: GenomicRanges
## Loading required package: BiocGenerics
## Loading required package: parallel
##
## Attaching package: 'BiocGenerics'
##
## The following objects are masked from 'package:parallel':
##
##   clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
##   clusterExport, clusterMap, parApply, parCapply, parLapply,
##   parLapplyLB, parRapply, parSapply, parSapplyLB
##
## The following object is masked from 'package:stats':
##
##   xtabs
##
## The following objects are masked from 'package:base':
##
##   anyDuplicated, append, as.data.frame, as.vector, cbind,
##   colnames, do.call, duplicated, eval, evalq, Filter, Find, get,
##   intersect, is.unsorted, lapply, Map, mapply, match, mget,
##   order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##   rbind, Reduce, rep.int, rownames, sapply, setdiff, sort,
##   table, tapply, union, unique, unlist
##
## Loading required package: IRanges
## Loading required package: GenomeInfoDb
##
##   chrom chromStart chromEnd   name gieStain
## 1  chr1           0  2300000 p36.33   gneg
## 2  chr1      2300000  5300000 p36.32  gpos25
## 3  chr1      5300000  7100000 p36.31   gneg
## 4  chr1      7100000  9200000 p36.23  gpos25
## 5  chr1      9200000 12600000 p36.22   gneg
```

```
## 6 chr1 12600000 16100000 p36.21 gpos50
```

2.2 mapping data

The second input file `mapping data` is an R data frame which includes values to be drawn in the graph. Columns 1 and 2 are segment name and position respectively. And the third column is optional which can be the value or name. In the following example, the third column is the gene symbol.

```
1 options(stringsAsFactors = FALSE);
2 # load the OmicCircos-package
3 library(OmicCircos);
4 # load the gene expression data sets as the mapping data
5 data(TCGA.BC.gene.exp.2k.60);
6 # display the first six rows and the first six columns of the data
7 head(TCGA.BC.gene.exp.2k.60[,c(1:6)]);
```

##	chr	po	NAME	TCGA.A1.A0SK.01A	TCGA.A1.A0SO.01A	TCGA.A2.A04W.01A
## 1	1	939246	ISG15	-3.618	-2.286	-0.998
## 2	1	2533140	MMEL1	-2.832	-3.093	1.037
## 3	1	6446321	TNFRSF25	2.559	-0.660	-0.705
## 4	1	7832974	UTS2	1.708	-0.726	-0.102
## 5	1	7912164	TNFRSF9	-0.189	-0.768	1.020
## 6	1	9989253	RBP7	0.817	-0.463	-1.472

2.3 linking data

The third file `linking data` is for linking two points. Columns 1 and 4 are segment names for the two anchor points. Columns 2 and 5 are the point positions on the segments. Columns 3 and 6 are the names of the two points. Column 7 is optional and could be used for the link type description.

```
1 options(stringsAsFactors = FALSE);
2 # load the OmicCircos-package
3 library(OmicCircos);
4 # load the gene fusion data sets as the linked data
5 data(TCGA.BC.fus);
6 # display the first six rows and the first six columns of the data
7 head(TCGA.BC.fus[,c(1:6)]);
```

##	chr1	po1	gene1	chr2	po2	gene2
## 1	2	63456333	WDPCP	10	37493749	ANKRD30A
## 2	18	14563374	PARD6G	21	14995400	POTED
## 3	10	37521495	ANKRD30A	3	49282645	CCDC36
## 4	10	37521495	ANKRD30A	7	100177212	LRCH4
## 5	18	18539803	ROCK1	18	112551	PARD6G
## 6	12	4618159	C12orf4	18	1514414	PARD6G

2.4 linking polygon data

The last input file linking polygon data is for linking two sub-segments. In the data frame, columns 1, 2 and 3 are name, start and end points for first segment and columns 4, 5 and 6 are for the second segment in the same order. Here is an example by homological analysis between human and mouse. One row is one of the homological segments between the species.

```
1 options(stringsAsFactors = FALSE);
2 # load the OmicCircos-package
3 library(OmicCircos);
4 # load the genome comparison data sets as the linked data
5 data(UCSC.hs_cyto_mm);
6 # display the first six rows of the data
7 head(UCSC.hs_cyto_mm);
```

##	hs.chr	hs.start	hs.end	mm.chr	mm.start	mm.end
## 1	1	168564	19853547	4	155606060	137469648
## 2	1	19999359	19853547	14	119903262	124902244
## 3	1	20114146	42595152	4	137324970	117524887
## 4	1	42702806	42595152	2	156994785	117524887
## 5	1	42707481	43332813	4	117433768	116843734
## 6	1	43355885	43332813	7	100327829	116843734

3 The package functions

There are three main functions in the package: `sim.circos`, `seg.engle.po` and `circos`.

3.1 `sim.circos`

The `sim.circos` function is a simulation function that is used to generate the four input files. In the following example, there are 10 segments, 10 individuals, 10 links and 10 link.pgs. For each segment, the range of the point number is from 20 to 50. The values will be generated by $\text{rnorm}(1) + i$. The i is the ordinal number of the segments. The values are increased by the segment order.

```
1 options(stringsAsFactors = FALSE);
2 # load the OmicCircos-package
3 library(OmicCircos);
4 # set up the initial parameters
5 seg.num      <- 10;
6 ind.num      <- 20;
7 seg.po       <- c(20:50);
8 link.num     <- 10;
9 link.pg.num  <- 10;
10 # run sim.circos function
11 sim.out      <- sim.circos(seg=seg.num, po=seg.po, ind=ind.num, link=link.num, link.pg=
    link.pg.num);
12 # display the data set names
13 names(sim.out)
14 # display the segment data
15 head(sim.out$seg.frame[,c(1:3)])
```

```
##      seg.name seg.Start seg.End
## 1      chr1         0         1
## 2      chr1         1         2
## 3      chr1         2         3
## 4      chr1         3         4
## 5      chr1         4         5
## 6      chr1         5         6
```

```
1 # display the mapping data
2 head(sim.out$seg.mapping[,c(1:5)])
```

```
##      seg.name seg.po name1  name2  name3
## 1      chr1      1 1.788    1.3 -0.814
## 2      chr1      2 1.891 -0.627 -0.604
## 3      chr1      3 1.905  0.934  2.133
## 4      chr1      4 2.473 -1.221  1.823
## 5      chr1      5 1.001  0.759  0.096
## 6      chr1      6 0.878  1.177  0.504
```

```
1 # display the linking data
2 head(sim.out$seg.link)
```

```
##      seg1 po1 name1 seg2 po2 name2 name3
## 1 chr7  23    n1 chr3   7    n1    n1
## 2 chr6  27    n2 chr8  26    n2    n2
## 3 chr7  26    n3 chr4   0    n3    n3
## 4 chr6  28    n4 chr2   5    n4    n4
## 5 chr7  40    n5 chr1  14    n5    n5
## 6 chr6  13    n6 chr6   1    n6    n6
```

```
1 # display the linking polygon data
2 head(sim.out$seg.link.pg)
```

```
##      seg1 start1 end1  seg2 start2 end2
## 1 chr4      19  30  chr8      0  14
## 2 chr7      14  38  chr4     20  30
## 3 chr5      22  19  chr8     16  25
## 4 chr4       5  27  chr1     16   1
## 5 chr9       2  14 chr10     16  13
## 6 chr3      23  31  chr4      6  14
```

3.2 segAnglePo

The segAnglePo function converts the segment pointer positions into angle values and returns a data frame.

```
1 options(stringsAsFactors = FALSE);
2 # load the OmicCircos-package
3 library(OmicCircos);
4 # set up the initial parameters
5 seg.num      <- 10;
6 ind.num      <- 20;
7 seg.po       <- c(20:50);
8 link.num     <- 10;
9 link.pg.num  <- 10;
10 # run sim.circos function
11 sim.out      <- sim.circos(seg=seg.num, po=seg.po, ind=ind.num, link=link.num, link.pg=
    link.pg.num);
12 # To get the segment data
13 seg.f        <- sim.out$seg.frame;
14 # rename the segment names
15 seg.name     <- paste("chr", 1:seg.num, sep="");
16 # run segAnglePo function
17 db           <- segAnglePo(seg.f, seg=seg.name);
18 db[,2]       <- round(as.numeric(db[,2]), 3);
19 db[,3]       <- round(as.numeric(db[,3]), 3);
20 # To display the result
21 db
```

```
##      seg.name angle.start angle.end seg.sum.start seg.sum.end seg.start
## [1,] "chr1"    "270"        "297.651" "0"          "27"         "0"
## [2,] "chr2"    "299.651"    "331.398" "27"         "58"         "0"
## [3,] "chr3"    "333.398"    "377.434" "58"         "101"        "0"
```

```
## [4,] "chr4" "379.434" "427.566" "101" "148" "0"
## [5,] "chr5" "429.566" "470.53" "148" "188" "0"
## [6,] "chr6" "472.53" "502.229" "188" "217" "0"
## [7,] "chr7" "504.229" "528.807" "217" "241" "0"
## [8,] "chr8" "530.807" "551.289" "241" "261" "0"
## [9,] "chr9" "553.289" "599.373" "261" "306" "0"
## [10,] "chr10" "601.373" "628" "306" "332" "0"
## seg.end
## [1,] "27"
## [2,] "31"
## [3,] "43"
## [4,] "47"
## [5,] "40"
## [6,] "29"
## [7,] "24"
## [8,] "20"
## [9,] "45"
## [10,] "26"
```

In the above example, there are 10 segments, one segment per row. Column 1 is segment name. Columns 2, 3 are the start and end angles of the segment. Column 4 and 5 are the accumulative start and end points. Column 6 and 7 are the start and end point for the segment. The plotting is clockwise and the start pointer is at 12 oclock (270 degree) only.

3.3 circos

The circos is the main function to draw different shapes of the circle. The shapes can be scatterplots, lines and polygons. The function supports multiple samples to draw boxplot, histogram, multiple lines and heatmap. The detail usages of the function are in next sections.

4 Plotting parameters

4.1 basic plotting

The input data sets were generated by `sim.circos` function.

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3 options(stringsAsFactors = FALSE);
4 set.seed(1234);
5
6 # initial
7 seg.num    <- 10;
8 ind.num    <- 20;
9 seg.po     <- c(20:50);
10 link.num   <- 10;
11 link.pg.num <- 10;
12
13 sim.out <- sim.circos(seg=seg.num, po=seg.po, ind=ind.num, link=link.num,
14   link.pg=link.pg.num);
15
16 seg.f     <- sim.out$seg.frame;
17 seg.v     <- sim.out$seg.mapping;
18 link.v    <- sim.out$seg.link
19 link.pg.v <- sim.out$seg.link.pg
20 seg.num   <- length(unique(seg.f[,1]));
21
22 # name segment (option)
23 seg.name <- paste("chr", 1:seg.num, sep="");
24 db       <- segAnglePo(seg.f, seg=seg.name);
25 # set transparent colors
26 colors   <- rainbow(seg.num, alpha=0.5);
```

To create square figure and get perfect circle is by the same values in width, height and in the same values in the numbers of lines of the margin.

```
1 par(mar=c(2, 2, 2, 2));
2 #
3 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
4 #
5 circos(R=400, cir=db, type="chr", col=colors, print.chr.lab=T, W=4, scale=T);
6 circos(R=360, cir=db, W=40, mapping=seg.v, col.v=3, type="l", B=T, col=colors[1],
7   lwd=0.1, scale=T);
8 circos(R=320, cir=db, W=40, mapping=seg.v, col.v=3, type="ls", B=F, col=colors[3],
9   lwd=0.1, scale=T);
10 circos(R=280, cir=db, W=40, mapping=seg.v, col.v=3, type="lh", B=T, col=colors[7],
11   lwd=0.1, scale=T);
12 circos(R=240, cir=db, W=40, mapping=seg.v, col.v=19, type="ml", B=F, col=colors, lwd
13   =0.1, scale=T);
14 circos(R=200, cir=db, W=40, mapping=seg.v, col.v=19, type="ml2", B=T, col=colors, lwd
15   =0.1);
16 circos(R=160, cir=db, W=40, mapping=seg.v, col.v=19, type="ml3", B=F, cutoff=5, lwd=0
17   .1);
18 circos(R=150, cir=db, W=40, mapping=link.v, type="link", lwd=2, col=colors);
19 circos(R=150, cir=db, W=40, mapping=link.pg.v, type="link.pg", lwd=2, col=colors);
```

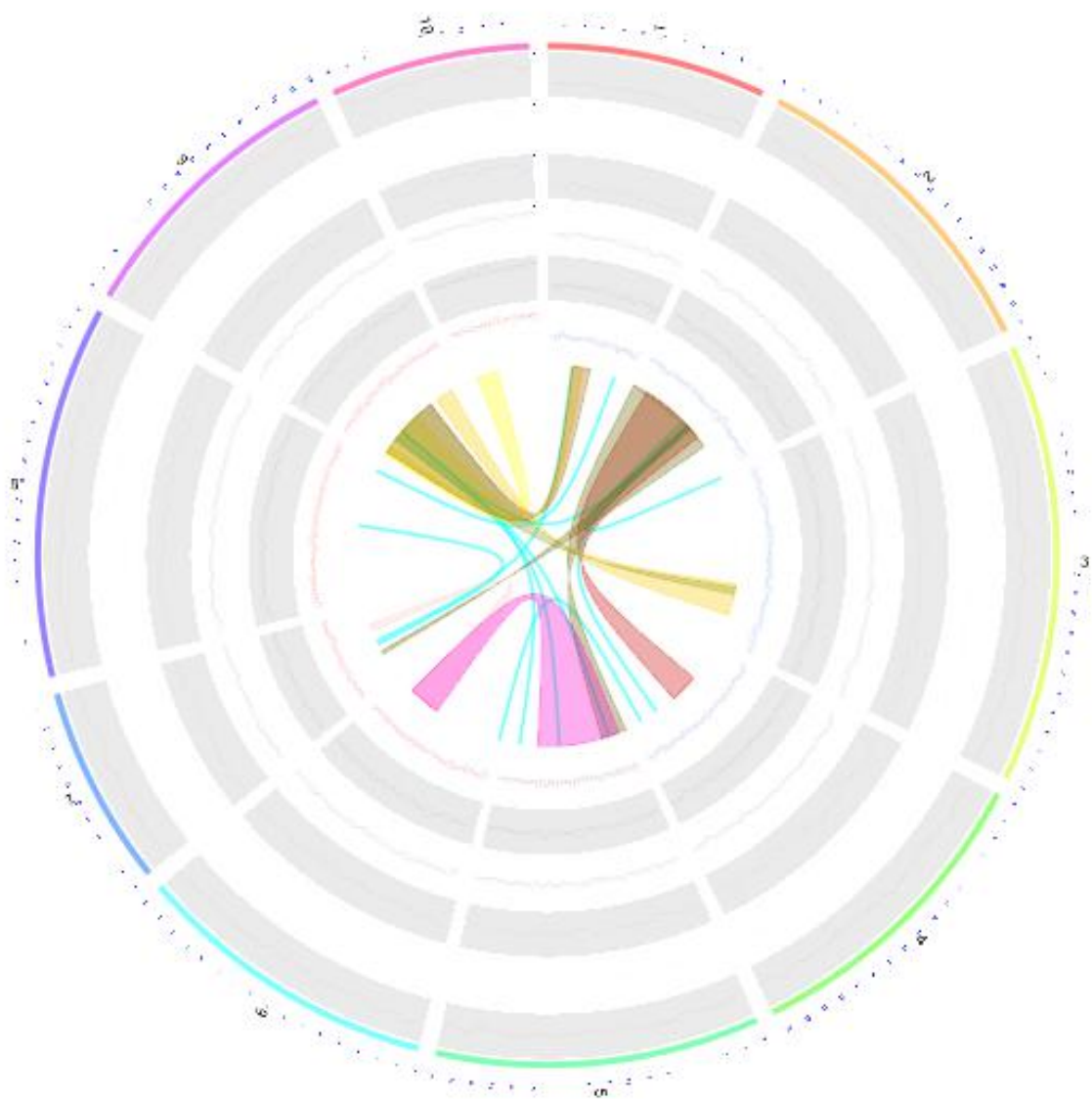


Figure 1

In figure 2, from outside to inside, the first track is the boxplot by the samples from column 8 (col.v=8) to the last column sample with the scale. Track 2 is the histogram (in horizontal) for multiple samples. Track 3 is the scatter plots for multiple samples. Tracks 4, 5 and 6 are the histogram (in vertical), scatter plot and vertical line by one sample without the scale.

```

1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3 set.seed(1234);
4
5 ## initial
6 seg.num      <- 10;
7 ind.num      <- 20;
8 seg.po       <- c(20:50);
9 link.num     <- 10;
10 link.pg.num  <- 10;
11
12 sim.out <- sim.circos(seg=seg.num, po=seg.po, ind=ind.num, link=link.num,
13   link.pg=link.pg.num);
14
15 seg.f      <- sim.out$seg.frame;
16 seg.v      <- sim.out$seg.mapping;
17 link.v     <- sim.out$seg.link
18 link.pg.v  <- sim.out$seg.link.pg
19 seg.num    <- length(unique(seg.f[,1]));
20
21 ## select segments
22 seg.name <- paste("chr", 1:seg.num, sep="");
23 db      <- segAnglePo(seg.f, seg=seg.name);
24
25 colors  <- rainbow(seg.num, alpha=0.5);

```

```

1 par(mar=c(2, 2, 2, 2));
2
3 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
4 circos(R=400, type="chr", cir=db, col=colors, print.chr.lab=T, W=4, scale=T);
5 circos(R=360, cir=db, W=40, mapping=seg.v, col.v=8, type="box", B=T, col=colors[1],
6   lwd=0.1, scale=T);
7 circos(R=320, cir=db, W=40, mapping=seg.v, col.v=8, type="hist", B=T, col=colors[3],
8   lwd=0.1, scale=T);
9 circos(R=280, cir=db, W=40, mapping=seg.v, col.v=8, type="ms", B=T, col=colors[7],
10   lwd=0.1, scale=T);
11 circos(R=240, cir=db, W=40, mapping=seg.v, col.v=3, type="h", B=F, col=colors[2],
12   lwd=0.1);
13 circos(R=200, cir=db, W=40, mapping=seg.v, col.v=3, type="s", B=T, col=colors, lwd=0.1
14   );
15 circos(R=160, cir=db, W=40, mapping=seg.v, col.v=3, type="b", B=F, col=colors, lwd=0.1
16   );
17 circos(R=150, cir=db, W=40, mapping=link.v, type="link", lwd=2, col=colors);
18 circos(R=150, cir=db, W=40, mapping=link.pg.v, type="link.pg", lwd=2, col=colors);

```

In figure 3, from outside to inside, track 1 is the three lines for quantile values by the samples from column 8 (col.v=8) with the scale. The middle line is for the median, the outside line is for 90% (or 75% if using type=<93>quant75<94>) and the inside line is for 1-90%. Track 2 is the circle points with the center=median and radius=variance. Track 3 is the circle plot with the center equal to the mean and scaled value (for example, the range from 0 to 3). Tracks 4 is the heatmap. Track 5 is the circle plot with the center=median and radius=standard deviation. Track 6 is the 95%

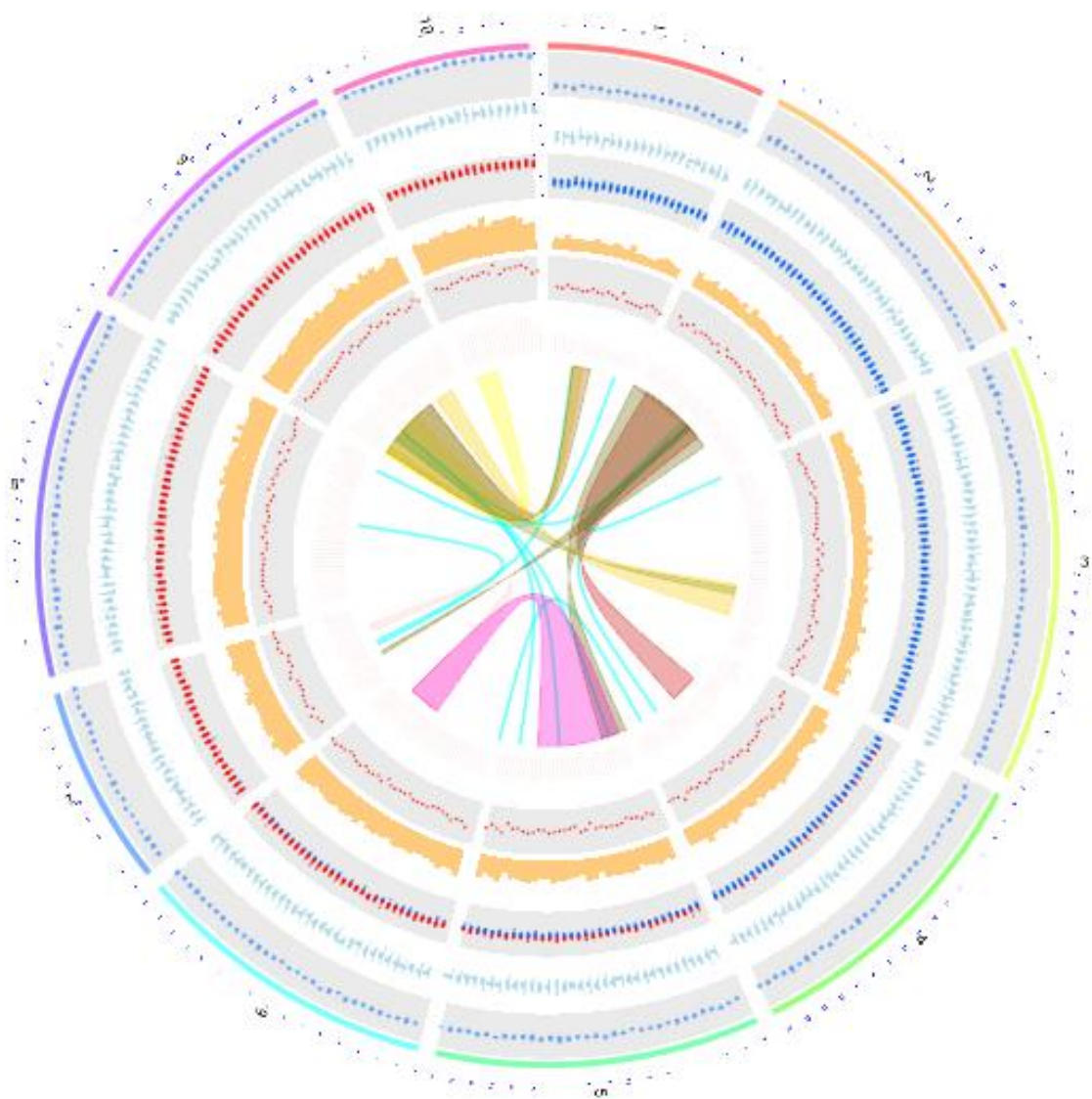


Figure 2

confidence interval of the samples.

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3 set.seed(1234);
4
5 ## initial
6 seg.num      <- 10;
7 ind.num      <- 20;
8 seg.po       <- c(20:50);
9 link.num     <- 10;
10 link.pg.num  <- 10;
11
12 sim.out <- sim.circos(seg=seg.num, po=seg.po, ind=ind.num, link=link.num,
13   link.pg=link.pg.num);
14
15 seg.f      <- sim.out$seg.frame;
16 seg.v      <- sim.out$seg.mapping;
17 link.v     <- sim.out$seg.link
18 link.pg.v  <- sim.out$seg.link.pg
19 seg.num    <- length(unique(seg.f[,1]));
20
21 ##
22 seg.name   <- paste("chr", 1:seg.num, sep="");
23 db        <- segAnglePo(seg.f, seg=seg.name);
24
25 colors    <- rainbow(seg.num, alpha=0.5);

1 par(mar=c(2, 2, 2, 2));
2 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
3
4 circos(R=400, type="chr", cir=db, col=colors, print.chr.lab=T, W=4, scale=T);
5 circos(R=360, cir=db, W=40, mapping=seg.v, col.v=8, type="quant90", B=F, col=colors,
6   lwd=0.1, scale=T);
7 circos(R=320, cir=db, W=40, mapping=seg.v, col.v=3, type="sv", B=T, col=colors[7], lwd
8   =0.1, scale=T);
9 circos(R=280, cir=db, W=40, mapping=seg.v, col.v=3, type="ss", B=F, col=colors[3], lwd
10  =0.1, scale=T);
11 circos(R=240, cir=db, W=40, mapping=seg.v, col.v=8, type="heatmap", lwd=3);
12 circos(R=200, cir=db, W=40, mapping=seg.v, col.v=3, type="s.sd", B=F, col=colors[4],
13   lwd=0.1);
14 circos(R=160, cir=db, W=40, mapping=seg.v, col.v=3, type="ci95", B=T, col=colors[4],
15   lwd=0.1);
16 circos(R=150, cir=db, W=40, mapping=link.v, type="link", lwd=2, col=colors);
17 circos(R=150, cir=db, W=40, mapping=link.pg.v, type="link.pg", lwd=2, col=colors);
18
19 the.col1=rainbow(10, alpha=0.3)[3];
20 highlight <- c(160, 410, 6, 2, 6, 10, the.col1, the.col1);
21 circos(R=110, cir=db, W=40, mapping=highlight, type="hl", lwd=2);
22
23 the.col1=rainbow(10, alpha=0.01)[3];
24 the.col2=rainbow(10, alpha=0.8)[1];
25 highlight <- c(160, 410, 6, 12, 7, 10, the.col1, the.col2);
26 circos(R=110, cir=db, W=40, mapping=highlight, type="hl", lwd=2);
```

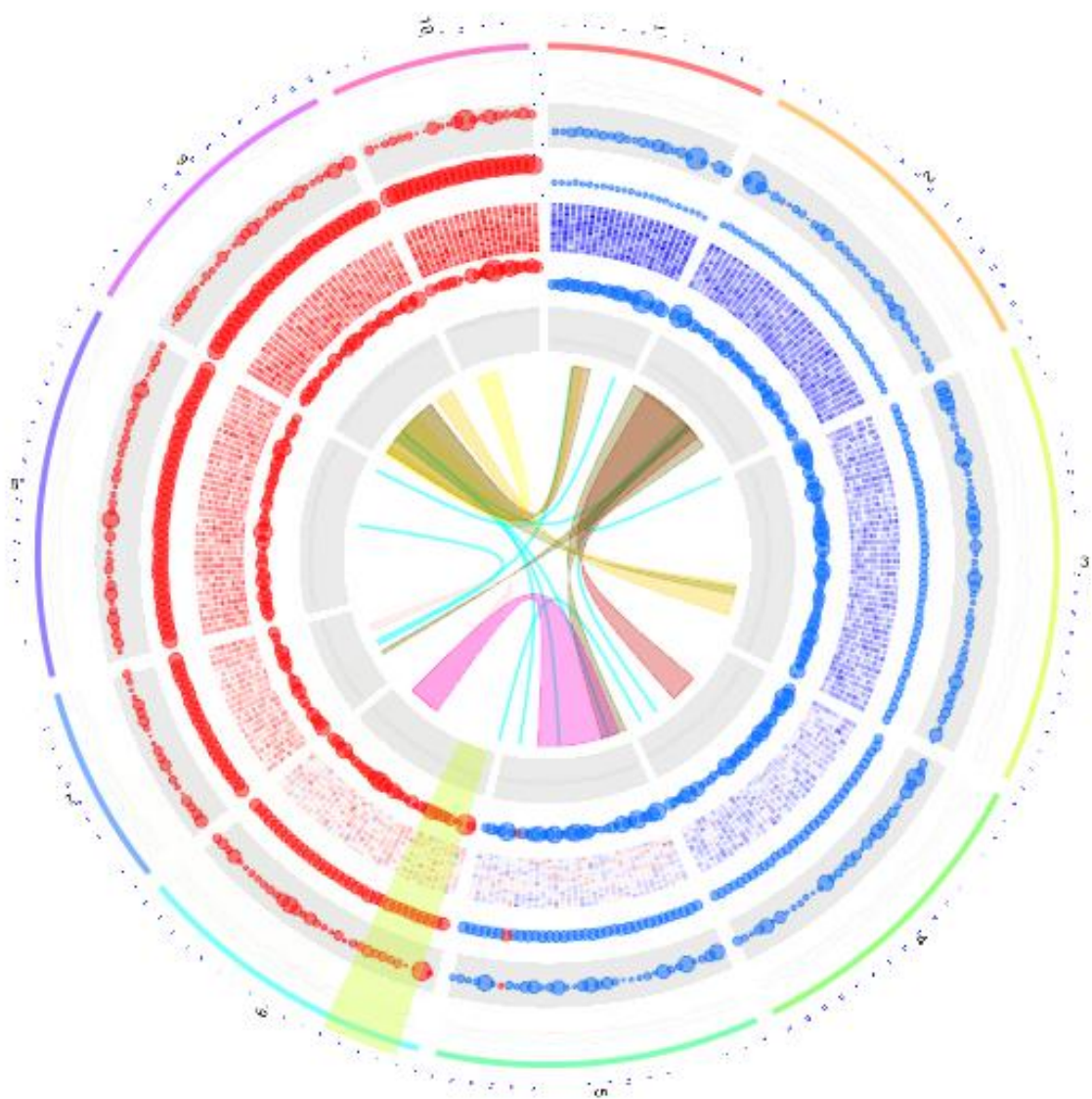


Figure 3

4.2 lables

An example of adding outside label.

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3
4 data("TCGA.PAM50_genefu_hg18");
5 data("TCGA.BC.fus");
6 data("TCGA.BC.cnv.2k.60");
7 data("TCGA.BC.gene.exp.2k.60");
8 data("TCGA.BC.sample60");
9 data("TCGA.BC_Her2_cnv_exp");
10
11 pvalue <- -1 * log10(TCGA.BC_Her2_cnv_exp[,5]);
12 pvalue <- cbind(TCGA.BC_Her2_cnv_exp[,c(1:3)], pvalue);
13
14 Her2.i <- which(TCGA.BC.sample60[,2] == "Her2");
15 Her2.n <- TCGA.BC.sample60[Her2.i,1];
16
17 Her2.j <- which(colnames(TCGA.BC.cnv.2k.60) %in% Her2.n);
18 cnv <- TCGA.BC.cnv.2k.60[,c(1:3,Her2.j)];
19 cnv.m <- cnv[,c(4:ncol(cnv))];
20 cnv.m[cnv.m > 2] <- 2;
21 cnv.m[cnv.m < -2] <- -2;
22 cnv <- cbind(cnv[,1:3], cnv.m);
23
24 Her2.j <- which(colnames(TCGA.BC.gene.exp.2k.60) %in% Her2.n);
25 gene.exp <- TCGA.BC.gene.exp.2k.60[,c(1:3,Her2.j)];
26 colors <- rainbow(10, alpha=0.5);
```

```
1 par(mar=c(2, 2, 2, 2));
2 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
3 circos(R=300, type="chr", cir="hg18", print.chr.lab=F, W=4);
4 circos(R=310, cir="hg18", W=20, mapping=TCGA.PAM50_genefu_hg18, type="label", side="
  out", col="black");
5 circos(R=250, cir="hg18", W=50, mapping=cnv, col.v=4, type="ml3", B=F, col=colors[7],
  cutoff=0, scale=T);
6 circos(R=200, cir="hg18", W=50, mapping=gene.exp, col.v=4, type="ml3", B=T, col=colors
  [3], cutoff=0, scale=T);
7 circos(R=140, cir="hg18", W=50, mapping=pvalue, col.v=4, type="l", B=F, col=colors[1],
  scale=T);
8 circos(R=132, cir="hg18", W=50, mapping=TCGA.BC.fus, type="link", lwd=2);
```

This is an example of the inside label.

```
1 par(mar=c(2, 2, 2, 2));
2 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
3 circos(R=300, type="chr", cir="hg18", col=T, print.chr.lab=F, W=4);
4 circos(R=290, cir="hg18", W=20, mapping=TCGA.PAM50_genefu_hg18, type="label", side="in
  ", col="blue");
5 circos(R=310, cir="hg18", W=50, mapping=cnv, col.v=4, type="ml3", B=T, col=colors[7],
  cutoff=0, scale=T);
6 circos(R=150, cir="hg18", W=50, mapping=gene.exp, col.v=4, type="ml3", B=T, col=colors
  [3], cutoff=0, scale=T);
7 circos(R=90, cir="hg18", W=50, mapping=pvalue, col.v=4, type="l", B=F, col=colors[1],
  scale=T);
```

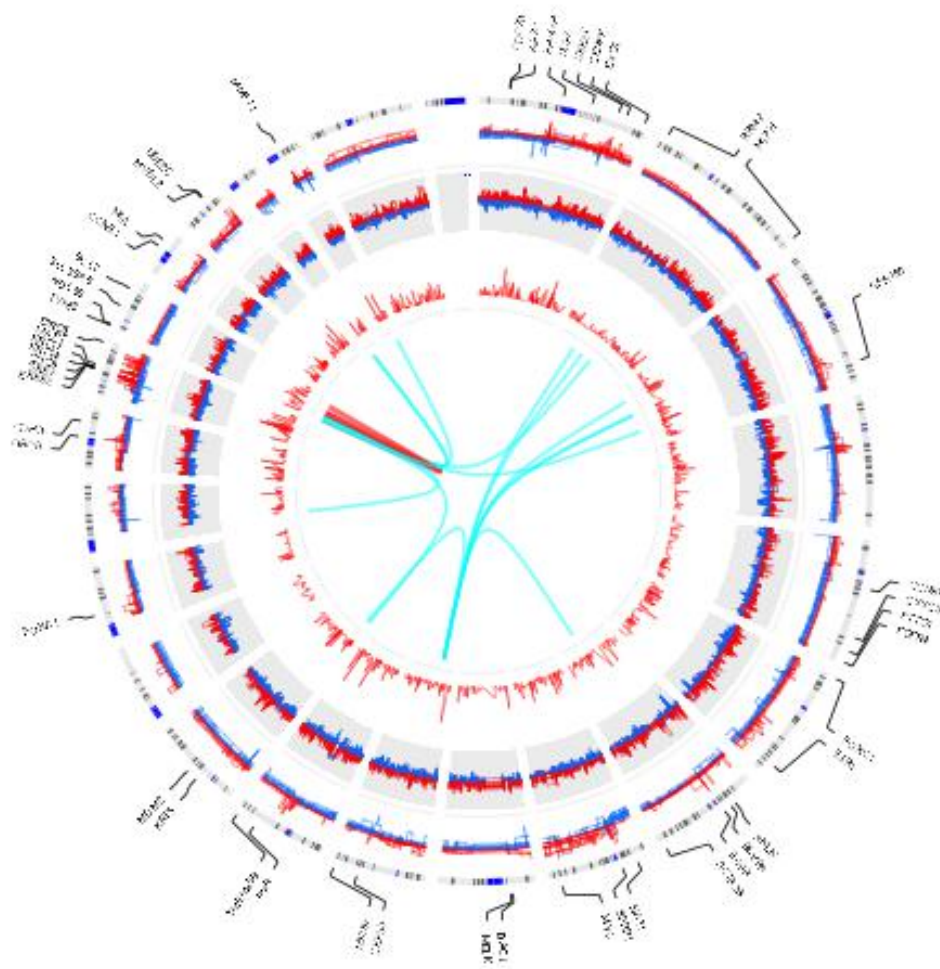



Figure 4


```
8 | circos(R=82, cir="hg18", W=50, mapping=TCGA.BC.fus, type="link", lwd=2);
```

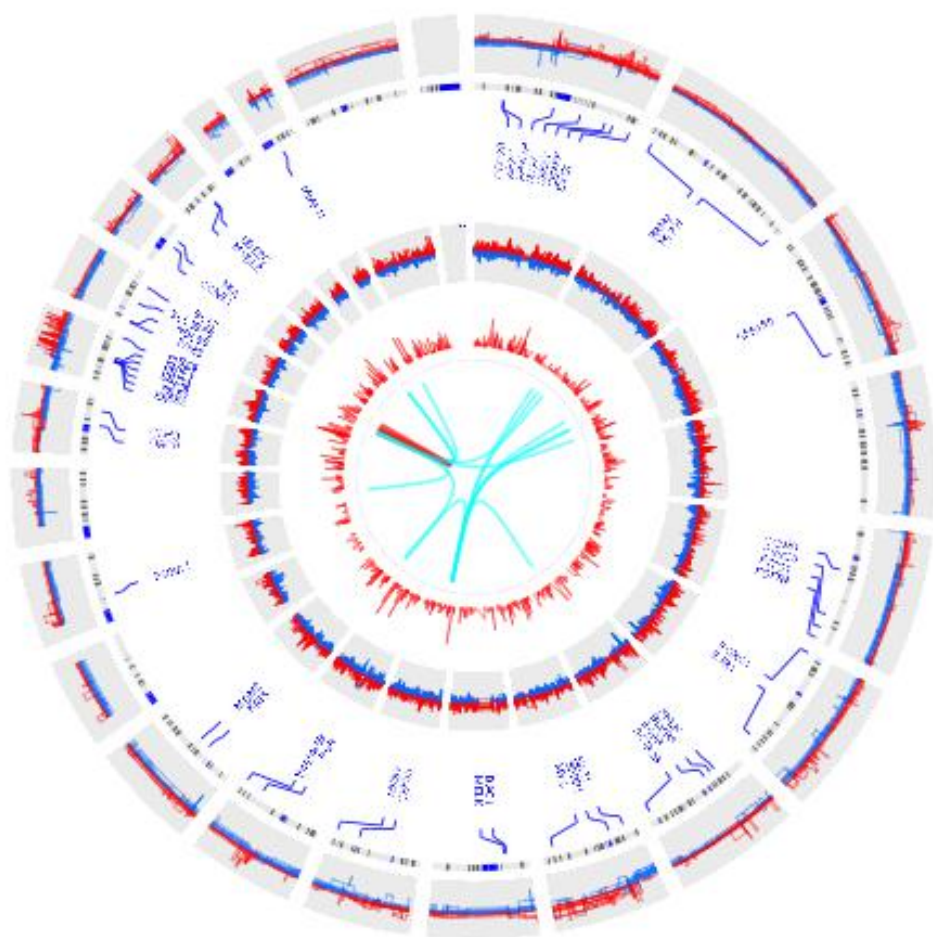


Figure 5

4.3 cluster and heatmap legend

An example of the clustered heatmap with the cluster structure and the color bar.

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3
4 data("TCGA.PAM50_genefu_hg18");
5 data("TCGA.BC.fus");
6 data("TCGA.BC.cnv.2k.60");
7 data("TCGA.BC.gene.exp.2k.60");
8 data("TCGA.BC.sample60");
9 data("TCGA.BC_Her2_cnv_exp");
10
11 pvalue <- -1 * log10(TCGA.BC_Her2_cnv_exp[,5]);
12 pvalue <- cbind(TCGA.BC_Her2_cnv_exp[,c(1:3)], pvalue);
13
14 Her2.i <- which(TCGA.BC.sample60[,2] == "Her2");
15 Her2.n <- TCGA.BC.sample60[Her2.i,1];
16
17 Her2.j <- which(colnames(TCGA.BC.cnv.2k.60) %in% Her2.n);
18 cnv <- TCGA.BC.cnv.2k.60[,c(1:3, Her2.j)];
19 cnv.m <- cnv[,c(4:ncol(cnv))];
20 cnv.m[cnv.m > 2] <- 2;
21 cnv.m[cnv.m < -2] <- -2;
22 cnv <- cbind(cnv[,1:3], cnv.m);
23
24 Her2.j <- which(colnames(TCGA.BC.gene.exp.2k.60) %in% Her2.n);
25 gene.exp <- TCGA.BC.gene.exp.2k.60[,c(1:3, Her2.j)];
26
27 colors <- rainbow(10, alpha=0.5);

1 par(mar=c(2, 2, 2, 2));
2
3 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
4
5 circos(R=400, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T);
6 circos(R=400, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T);
7 circos(R=300, cir="hg18", W=100, mapping=gene.exp, col.v=4, type="heatmap2", cluster
  =T, col.bar=T, lwd=0.01);
8 circos(R=220, cir="hg18", W=80, mapping=cnv, col.v=4, type="ml3", B=F, lwd=1,
  cutoff=0);
9 circos(R=140, cir="hg18", W=80, mapping=pvalue, col.v=4, type="l", B=T, lwd=1,
  col=colors[1]);
10 circos(R=130, cir="hg18", W=10, mapping=TCGA.BC.fus, type="link", lwd=2);
```

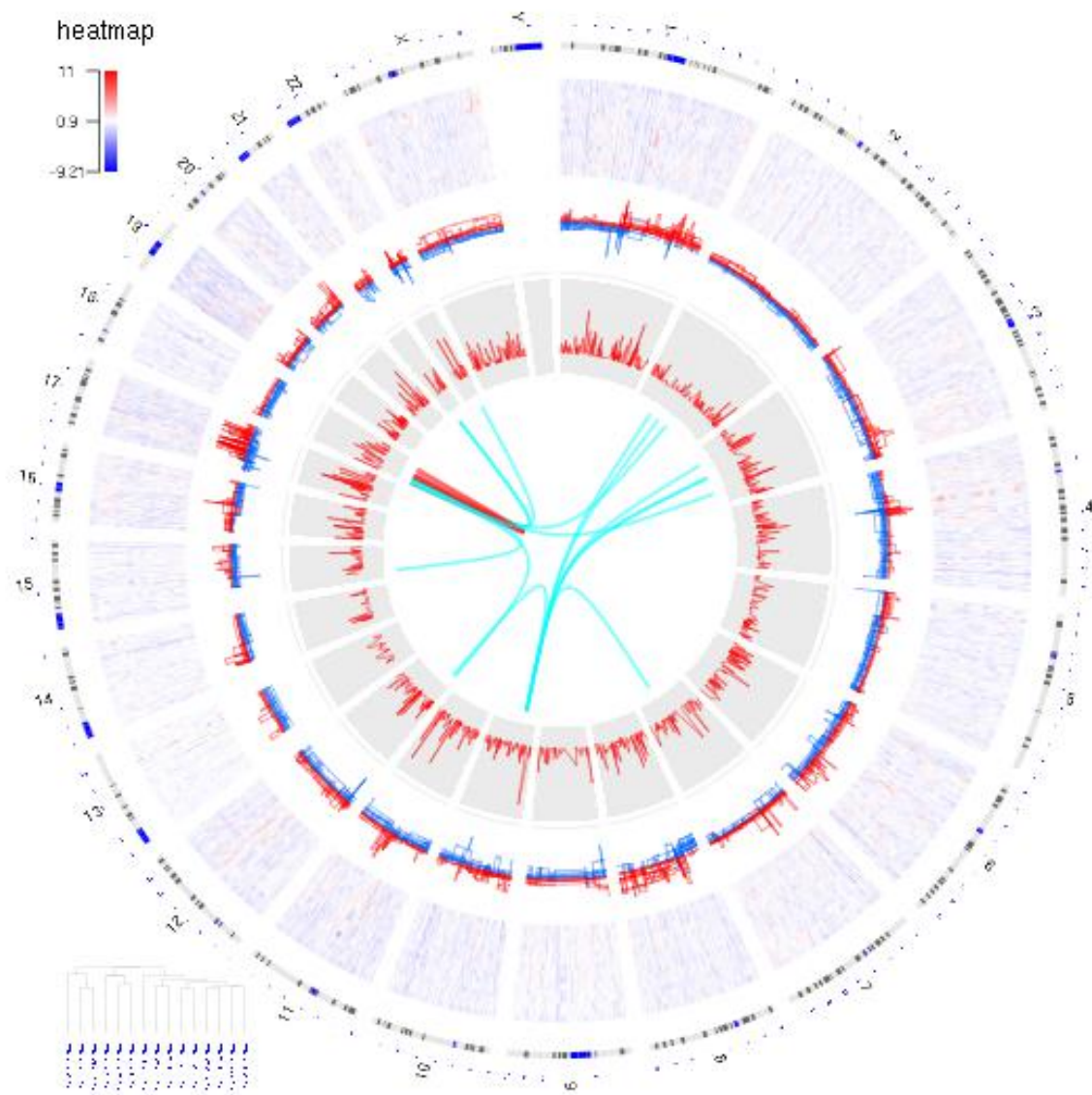


Figure 6

4.4 traditional plotting and OmicCircos

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3
4 data("TCGA.BC.fus");
5 data("TCGA.BC.cnv.2k.60");
6 data("TCGA.BC.gene.exp.2k.60");
7 data("TCGA.BC.sample60");
8
9 ## gene expression data for PCA
10 exp.m <- TCGA.BC.gene.exp.2k.60[,c(4:ncol(TCGA.BC.gene.exp.2k.60))];
11 cnv <- TCGA.BC.cnv.2k.60;
12 type.n <- unique(TCGA.BC.sample60[,2]);
13 colors <- rainbow(length(type.n), alpha=0.5);
14
15 ## sub-type colors
16 pca.col <- rep(NA, nrow(TCGA.BC.sample60));
17 for (i in 1:length(type.n)){
18   n <- type.n[i];
19   n.i <- which(TCGA.BC.sample60[,2] == n);
20   n.n <- TCGA.BC.sample60[n.i,1];
21   g.i <- which(colnames(exp.m) %in% n.n);
22   pca.col[g.i] <- colors[i];
23 }
24
25 ## run PCA
26 exp.m <- na.omit(exp.m);
27 pca.out <- prcomp(t(exp.m), scale = TRUE);
28
29 ## subtype cnv
30 cnv.i <- c();
31 for (i in 1:length(type.n)){
32   n <- type.n[i];
33   n.i <- which(TCGA.BC.sample60[,2] == n);
34   n.n <- TCGA.BC.sample60[n.i,1];
35   cnv.i <- which(colnames(cnv) %in% n.n);
36 }
37
38 ## PCA is plotting.
39 plot(pca.out$x[,1]*5, pca.out$x[,2]*5, pch=19, col=pca.col, main="",
40       cex=2, xlab="PC1", ylab="PC2", ylim=c(-200, 460), xlim=c(-200,460));
41 legend(200,0, c("Basal","Her2","LumA","LumB"), pch=19, col=colors[c(2,4,1,3)], cex=1,
42        title = "Gene Expression (PCA)", box.col="white");
43
44 ## It is going to plot the circos.
45 circos(xc=280, yc=280, R=168, cir="hg18", W=4, type="chr", print.chr.lab=T);
46 R.v <- 135;
47 for (i in 1:length(type.n)){
48   n <- type.n[i];
49   n.i <- which(TCGA.BC.sample60[,2] == n);
50   n.n <- TCGA.BC.sample60[n.i,1];
51   cnv.i <- which(colnames(cnv) %in% n.n);
52   cnv.v <- cnv[,cnv.i];
53   cnv.v[cnv.v > 2] <- 2;
```

```

17   cnv.v[cnv.v < -2] <- -2;
18   cnv.m <- cbind(cnv[,c(1:3)], cnv.v);
19   circos(xc=280, yc=280, R=R.v, cir="hg18", W=34, mapping=cnv.m, col.v=4, type="ml3",
20         B=F, lwd=0.5, cutoff=0);
21   R.v <- R.v - 25;
22 }
23 legend(-80,460, c("1 Basal", "2 Her2", "3 LumA", "4 LumB", "(center)"), cex=1,
24       title = "CNV (OmicCircos)", box.col="white");

```

It is an example, PCA plotting is at the center of the circos.

```

1 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
2
3 legend(680,800, c("Basal","Her2","LumA","LumB"), pch=19, col=colors[c(2,4,1,3)], cex=0
4       .5,
5       title = "Gene Expression (PCA)", box.col="white");
6
7 legend(5,800, c("1 Basal", "2 Her2", "3 LumA", "4 LumB", "(center)"), cex=0.5,
8       title = "CNV (OmicCircos)", box.col="white");
9
10 circos(xc=400, yc=400, R=390, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T);
11 R.v <- 330;
12 for (i in 1:length(type.n)){
13   n <- type.n[i];
14   n.i <- which(TCGA.BC.sample60[,2] == n);
15   n.n <- TCGA.BC.sample60[n.i,1];
16   cnv.i <- which(colnames(cnv) %in% n.n);
17   cnv.v <- cnv[,cnv.i];
18   cnv.v[cnv.v > 2] <- 2;
19   cnv.v[cnv.v < -2] <- -2;
20   cnv.m <- cbind(cnv[,c(1:3)], cnv.v);
21   circos(xc=400, yc=400, R=R.v, cir="hg18", W=60, mapping=cnv.m, col.v=4, type="ml3",
22         B=F, lwd=1, cutoff=0, scale=T);
23   R.v <- R.v - 60;
24 }
25
26 points(pca.out$x[,1]*3.6+400, pca.out$x[,2]*3.6+400, pch=19, col=pca.col, cex=2);

```

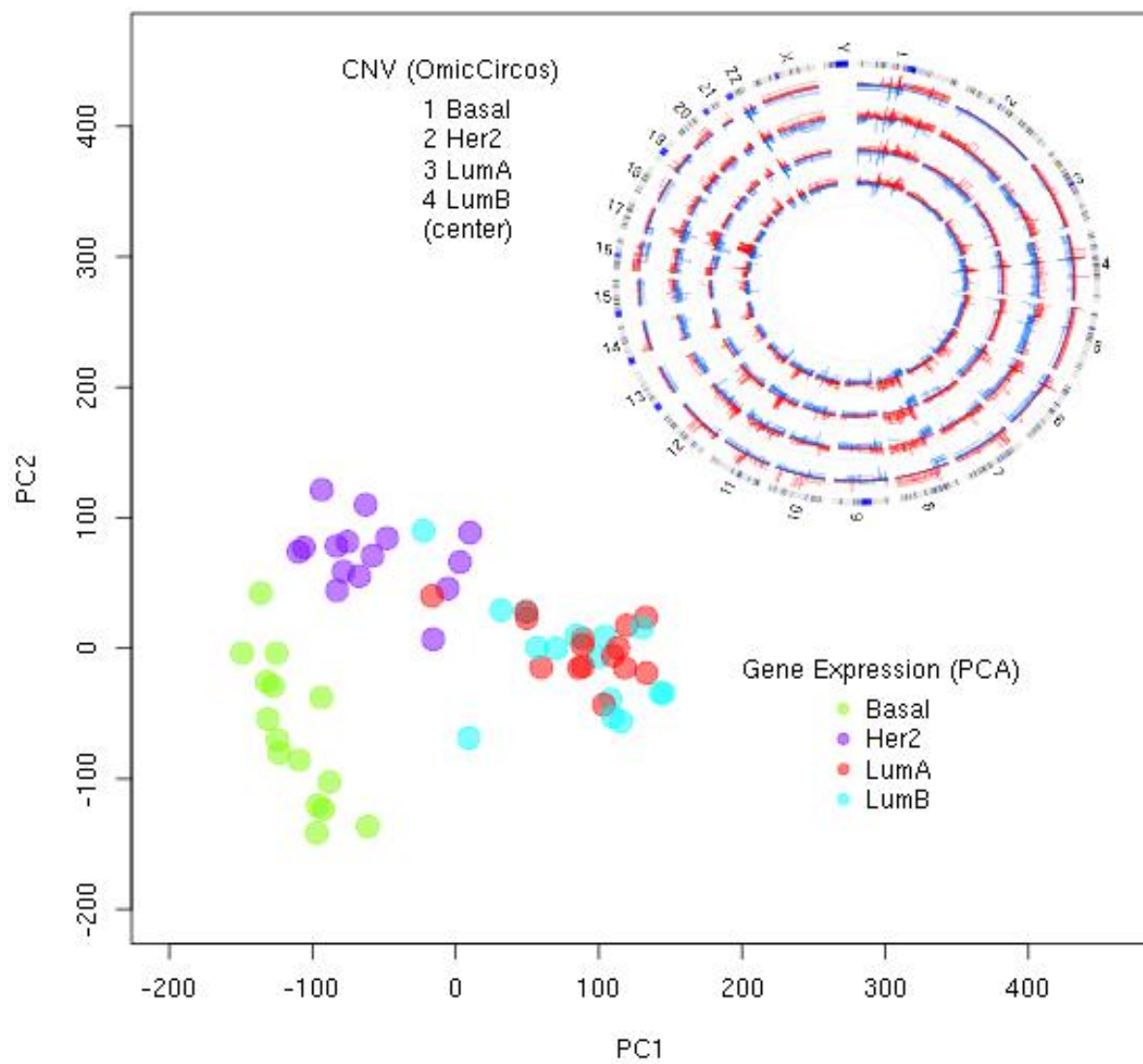


Figure 7

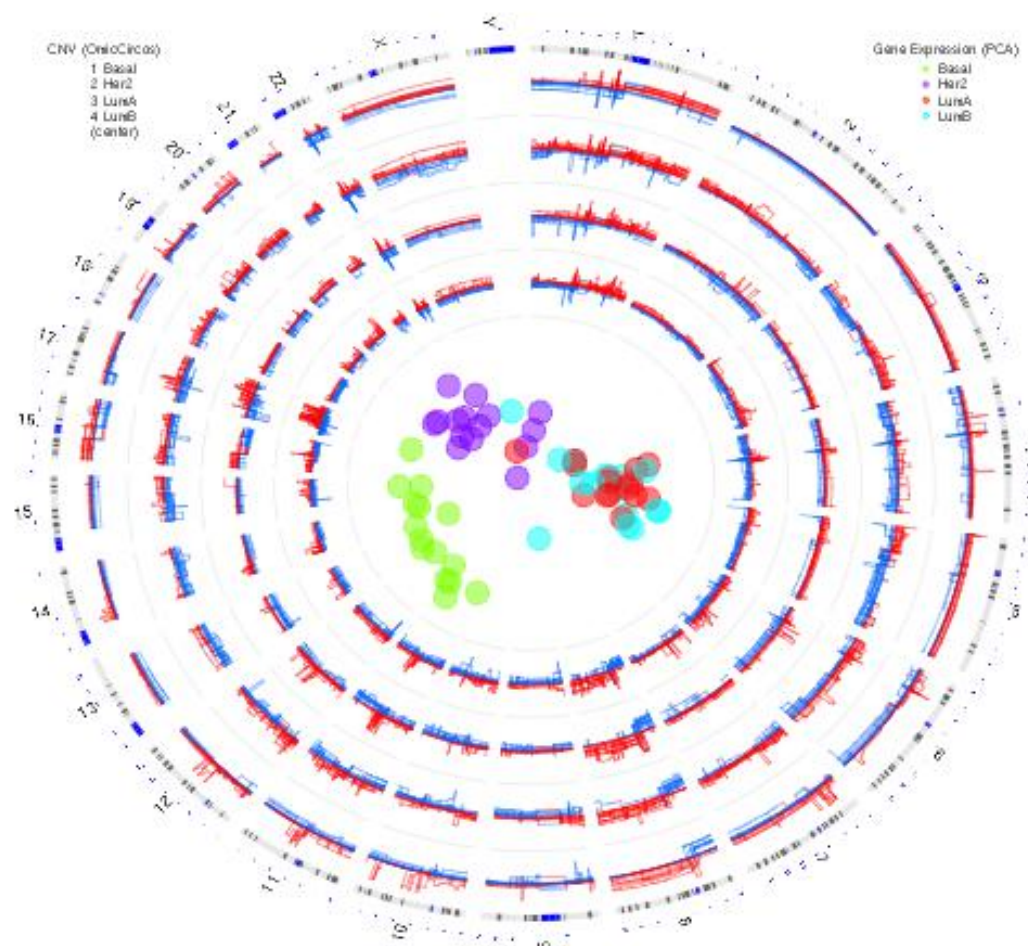


Figure 8

4.5 zoom

```
1 options(stringsAsFactors = FALSE);
2 library(OmicCircos);
3
4 data("TCGA.PAM50_genefu_hg18");
5 data("TCGA.BC.fus");
6 data("TCGA.BC.cnv.2k.60");
7 data("TCGA.BC.gene.exp.2k.60");
8 data("TCGA.BC.sample60");
9 data("TCGA.BC_Her2_cnv_exp");
10 data("TCGA.PAM50_genefu_hg18");
11
12 pvalue <- -1 * log10(TCGA.BC_Her2_cnv_exp[,5]);
13 pvalue <- cbind(TCGA.BC_Her2_cnv_exp[,c(1:3)], pvalue);
14
15 Her2.i <- which(TCGA.BC.sample60[,2] == "Her2");
16 Her2.n <- TCGA.BC.sample60[Her2.i,1];
17
18 Her2.j <- which(colnames(TCGA.BC.cnv.2k.60) %in% Her2.n);
19 cnv <- TCGA.BC.cnv.2k.60[,c(1:3, Her2.j)];
20 cnv.m <- cnv[,c(4:ncol(cnv))];
21 cnv.m[cnv.m > 2] <- 2;
22 cnv.m[cnv.m < -2] <- -2;
23 cnv <- cbind(cnv[,1:3], cnv.m);
24
25 gene.exp <- TCGA.BC.gene.exp.2k.60[,c(1:3, Her2.j)];
26
27 colors <- rainbow(10, alpha=0.5);

1 par(mar=c(2, 2, 2, 2));
2
3 plot(c(1,800), c(1,800), type="n", axes=F, xlab="", ylab="", main="");
4 # In figure 7, the chromosome 1 to chromosome 22 are going to be plotted from the
5 # angle 0 (12 Oclock)
6 # to 180 degree (6 Oclock).
7 zoom <- c(1, 22, 939245.5, 154143883, 0, 180);
8 circos(R=400, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T, zoom=zoom);
9 circos(R=300, cir="hg18", W=100, mapping=gene.exp, col.v=4, type="heatmap2", cluster=
10 T, col.bar=T, col.bar.po = "bottomright", lwd=0.01, zoom=zoom);
11 circos(R=220, cir="hg18", W=80, mapping=cnv, col.v=4, type="ml3", B=F, lwd=1,
12 cutoff=0, zoom=zoom);
13 circos(R=140, cir="hg18", W=80, mapping=pvalue, col.v=4, type="l", B=T, lwd=1,
14 col=colors[1], zoom=zoom);
15 circos(R=130, cir="hg18", W=10, mapping=TCGA.BC.fus, type="link", lwd=2, zoom=zoom);
16
17 # zoom in links by using the hightlight functions
18 # highlight
19 the.col1=rainbow(10, alpha=0.5)[1];
20
21 # The highline region is radium from 140 to 400 and from position 282412.5 to
22 # 133770314.5 in chromosome 11.
23 highlight <- c(140, 400, 11, 282412.5, 11, 133770314.5, the.col1, the.col1);
24 circos(R=110, cir="hg18", W=40, mapping=highlight, type="hl", lwd=2, zoom=zoom);
25 the.col2=rainbow(10, alpha=0.5)[6];
```

```

21 highlight <- c(140, 400, 17, 739525, 17, 78385909, the.col2, the.col2);
22 circos(R=110, cir="hg18", W=40, mapping=highlight, type="hl", lwd=2, zoom=zoom);
23 ## highlight link
24 highlight.link1 <- c(400, 400, 140, 376.8544, 384.0021, 450, 540.5);
25 circos(cir="hg18", mapping=highlight.link1, type="highlight.link", col=the.col1, lwd
    =1);
26 highlight.link2 <- c(400, 400, 140, 419.1154, 423.3032, 543, 627);
27 circos(cir="hg18", mapping=highlight.link2, type="highlight.link", col=the.col2, lwd
    =1);
28
29 # The chromosome 11 region is going plotting from 180 (6 O'clock) to 270 degree (9
    O'clock).
30 zoom <- c(11, 11, 282412.5, 133770314.5, 180, 270);
31 circos(R=400, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T, zoom=zoom);
32 circos(R=300, cir="hg18", W=100, mapping=gene.exp, col.v=4, type="heatmap2", cluster=
    T, lwd=0.01, zoom=zoom);
33 circos(R=220, cir="hg18", W=80, mapping=cnv, col.v=4, type="ml3", B=F, lwd=1,
    cutoff=0, zoom=zoom);
34 circos(R=140, cir="hg18", W=80, mapping=pvalue, col.v=4, type="l", B=T, lwd=1,
    col=colors[1], zoom=zoom);
35
36 # The chromosome 17 region is going plotting from 180 (6 O'clock) to 270 degree (9
    O'clock).
37
38 gene.names <- c("ERBB2", "CDC6");
39 PAM50.17 <- which(TCGA.PAM50_genefu_hg18[,3]==gene.names);
40 TCGA.PAM50 <- TCGA.PAM50_genefu_hg18[PAM50.17,];
41
42 # zoom in chromosome 17
43 zoom <- c(17, 17, 739525, 78385909, 274, 356);
44 circos(R=400, cir="hg18", W=4, type="chr", print.chr.lab=T, scale=T, zoom=zoom);
45 circos(R=300, cir="hg18", W=100, mapping=gene.exp, col.v=4, type="heatmap2", cluster=
    T, lwd=0.01, zoom=zoom);
46 circos(R=220, cir="hg18", W=80, mapping=cnv, col.v=4, type="ml3", B=F, lwd=1,
    cutoff=0, zoom=zoom);
47 circos(R=140, cir="hg18", W=80, mapping=pvalue, col.v=4, type="l", B=T, lwd=1,
    col=colors[1], zoom=zoom);
48 circos(R=410, cir="hg18", W=40, mapping=TCGA.PAM50, type="label", side="out", col="
    blue", zoom=zoom);

```

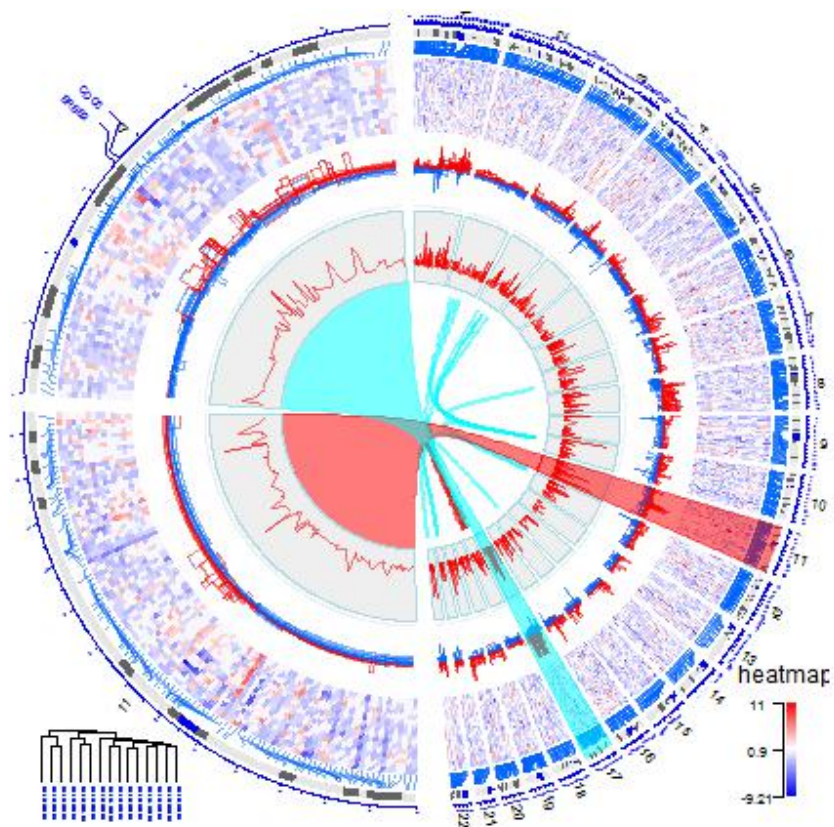


Figure 9